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# Stellatolide H, a cytotoxic peptide lactone from a deep-sea sponge *Discodermia* sp.



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#### ABSTRACT

Stellatolide H (1) was isolated from a deep-sea sponge *Discodermia* sp. as the cytotoxic constituent. The planar structure of 1 was elucidated on the basis of the NMR spectroscopic and mass spectrometric data. The absolute configurations of the constituent amino acid residues were determined by the Marfey's method. Stellatolide H (1) is a peptide lactone of the callipeltin class with its N-terminus blocked by 3-hydroxy-6,8-dimethyldeca-(4*Z*,6*E*)-dienoic acid (Hdda).

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Sessile marine organisms in the deep sea build unique communities which are different from those in shallow waters. Only those physiologically adapted to the deep sea can survive there. Because secondary metabolites are compounds distributed species-specifically, it is reasonable to speculate that distinct natural products are present in the deep-sea organisms.<sup>2</sup> During our search for bioactive metabolites from deep-sea sponges, 3,4 we found that the extract of a sponge of the genus Discodermia, collected at a depth of 310 m in the East China Sea, exhibited cytotoxic activity. From the sponges of this genus a variety of biologically active natural products of polyketide synthase/non-ribosomal peptide synthetase origin have been discovered.<sup>5,6</sup> Bioassay-guided fractionation of the sponge afforded a peptide lactone of the callipeltin class named stellatolide H (1). In this paper, we report the isolation, structure elucidation, and biological activity of stellatolide H (1).

The EtOH extract of the sponge *Discodermia* sp. was partitioned between H<sub>2</sub>O and CHCl<sub>3</sub>, and the aqueous phase was further extracted with *n*-BuOH. The CHCl<sub>3</sub> and *n*-BuOH fractions, both of which exhibited cytotoxicity against HeLa cells, were combined

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and separated by ODS flash chromatography followed by reversed-phase HPLC to afford stellatolide H (1) as a colorless amorphous solid, together with the known cyclolithistide A.<sup>7,8</sup>

Stellatolide H (1) had the molecular formula of  $C_{66}H_{106}N_{14}O_{21}$ as determined by HRESIMS. Preliminary analysis of the <sup>1</sup>H and  $^{13}$ C NMR spectra measured in DMSO- $d_6$  (Table S1) revealed the presence of several amide protons and carbonyl carbons suggesting its peptide nature. Most of the NMR signals were doubled in this solvent, indicating the presence of conformational equilibrium. This signal duplication was not observed in CD<sub>3</sub>OH (Table 1). Further analyses of the 2D NMR data mostly in CD<sub>3</sub>OH disclosed the presence of seven usual amino acid residues, Ser, Gly, Leu, Nmethylalanine (NMeAla), N-methylglutamine (NMeGln), and two residues of Thr or allo-Thr (aThr) together with four unusual residues. The amidation of the side chain of the NMeGln residue was demonstrated by ROESY cross peaks between a pair of mutually coupled amide protons ( $\delta_H$  7.27 and 6.80) and the  $\beta$ -methylene protons at  $\delta_H$  1.67 (2H). There was a spin system reminiscent of a Ser residue with a deshielded  $\beta$ -methylene carbon ( $\delta_C$  71.4) which was coupled with an O-methyl proton signal at  $\delta_H$  3.37 in the HMBC spectrum, indicating the presence of an O-methylserine (OMeSer) residue. The 2,3-diaminobutanoic acid (Dab) residue exhibited a spin system similar to that of a Thr residue, but with more shielded signals for the  $\beta$ -carbon ( $\delta_C$  48.8). The presence of

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Table 1  $^{1}\text{H}$  NMR (600 MHz) and  $^{13}\text{C}$  NMR (150 MHz) data for stellatolide H (1) in CD<sub>3</sub>OH.

Position	$\delta_{C}$	δ <sub>H</sub> (J in Hz)	НМВС
Hdda 1	171.0		
1 2a	174.6 44.9	2.51 m	1, 3, 4
2b	44.5	2.41 m	1, 3, 4
3	66.6	5.02 m	•
4	131.0	5.31 dd (11.5, 9.6)	6
5	136.0	5.91 d (11.5)	3, 6-Me, 8
6	132.3		
6-Me	16.8	1.77 s	5, 6, 7
7	140.0	5.18 d (9.6)	5, 6-Me, 8, 8-Me, 9
8	35.5	2.34 m	6, 7, 8-Me, 9
8-Me	20.9	0.95 d (6.9)	7, 8, 9
9a	31.2	1.36 m	7, 8-Me, 9, 10
9b	12.2	1.25 <sup>a</sup>	7, 8-Me, 9, 10
10	12.2	0.84 t (7.5)	8, 9
<u>Ser</u>	$ND^\mathrm{b}$		
11	56.8	4.20 m	
12	62.8	4.38 m 3.85 dd (4.8, 11.0)	
13a 13b	02.8	3.65 dd (4.8, 11.0) 3.79 m	
NH		8.29 brs	
<u>Thr</u>		0.23 bis	
1111 14	$ND^\mathrm{b}$		
15	60.1	4.40 m	
16	68.0	4.35 m	
17	20.1	1.19 d (6.2)	15, 16
NH		8.01 brs	,
<u>Dab</u>			
18	$ND^\mathrm{b}$		
19	56.8	4.51 <sup>a</sup>	
20	48.8	3.76 m	
$NH_2$		$ND^\mathrm{b}$	
21	16.8	1.33 d (6.2)	19, 20
NH		$ND^\mathrm{b}$	
Me <sub>2</sub> Gln			
22	$ND^{b}$		
23	58.9	4.09 d (8.25)	23, 24, 24-Me
24	37.1	2.38 m	23
24-Me	16.6	1.25 <sup>a</sup>	24, 25
25	44.4	2.68 m	25-Me, 26
25-Me	13.1	1.26 <sup>a</sup>	24, 25, 26
26	182.1	7.00.1	
NH <sub>2</sub>		7.89 br s	
NH		7.07 br s ND <sup>b</sup>	
		ND	
<u>aThr</u>	NDb		
27 28	ND <sup>b</sup>	E 26 m	
28 29	57.2 71.7	5.26 m 5.58 m	54
30	14.8	1.17 d (6.2)	28, 29
NH	14.0	8.88 d (9.6)	26, 25
<u>OMeSer</u>		0.00 a (0.0)	
31	173.6		
32	55.2	4.47 m	31
33a	71.4	3.80 m	31, 32, OMe
33b	,	3.72 dd (6.9, 9.6)	32, OMe
OMe	59.4	3.37 s	33
NH		8.42 br	
<u>Gly</u>			
34	$ND^\mathrm{b}$		
35a	44.0	3.92 d (17.2)	31
35b		3.50 d (17.2)	
NH		9.03 br	
Leu			
36	$ND^b$		
37	49.2	4.75 <sup>a</sup>	
38a	40.6	1.26 <sup>a</sup>	37
38b		1.56 m	
39	25.8	1.62 m	
40	21.5	0.88 d (6.9)	38, 39, 41
41	23.3	0.92 d (6.9)	38, 39, 40
NH		7.20 br s	

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