Alexandria Engineering Journal (2016) xxx, xxx-xxx



Alexandria University

Alexandria Engineering Journal

www.elsevier.com/locate/aej



ORIGINAL ARTICLE

Empirical modeling of drying kinetics and microwave assisted extraction of bioactive compounds from *Adathoda vasica* and *Cymbopogon citratus*

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Received 17 August 2014; revised 14 November 2015; accepted 20 December 2015

KEYWORDS

Microwave assisted extraction; Drying kinetics; Empirical modeling; Extraction; Food; Medicinal plants

Abstract To highlight the shortcomings in conventional methods of extraction, this study investigates the efficacy of Microwave Assisted Extraction (MAE) toward bioactive compound recovery from pharmaceutically-significant medicinal plants, Adathoda vasica and Cymbopogon citratus. Initially, the microwave (MW) drying behavior of the plant leaves was investigated at different sample loadings, MW power and drying time. Kinetics was analyzed through empirical modeling of drying data against 10 conventional thin-layer drying equations that were further improvised through the incorporation of Arrhenius, exponential and linear-type expressions. 81 semi-empirical Midilli equations were derived and subjected to non-linear regression to arrive at the characteristic drying equations. Bioactive compounds recovery from the leaves was examined under various parameters through a comparative approach that studied MAE against Soxhlet extraction. MAE of A. vasica reported similar yields although drastic reduction in extraction time (210 s) as against the average time of 10 h in the Soxhlet apparatus. Extract yield for MAE of C. citratus was higher than the conventional process with optimal parameters determined to be 20 g sample load, 1:20 sample/solvent ratio, extraction time of 150 s and 300 W output power. Scanning Electron Microscopy and Fourier Transform Infrared Spectroscopy were performed to depict changes in internal leaf morphology. © 2015 Faculty of Engineering, Alexandria University. Production and hosting by Elsevier B.V. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

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Peer review under responsibility of Faculty of Engineering, Alexandria University.

http://dx.doi.org/10.1016/j.aej.2015.12.020

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Please cite this article in press as: P. Simha et al., Empirical modeling of drying kinetics and microwave assisted extraction of bioactive compounds from *Adathoda vasica* and *Cymbopogon citratus*, Alexandria Eng. J. (2016), http://dx.doi.org/10.1016/j.aej.2015.12.020

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Nomenclature R^2 $a, a_1, b, b_1, c, g, k, k_1, k_2, n, n_1$ empirical coefficients coefficient of determination D_{eff} effective moisture diffusivity (m² s⁻¹) E_{RMS} root mean square error D_o pre-exponential factor (m² s⁻¹) RSS residual sum of squares E_a activation energy (W g^{-1}) drying time (min) pre-exponential factor (min⁻¹) W_c k_o weights of the crude extract (g) half the thickness of the sample (m) W_o weight of dry powdered sample (g) I. sample mass (g) m number of mathematical models used for empirical M Greek letters constant $(\min^{-1} m^{-2} s^{-1})$ modeling MC_i moisture content at specific time (g water/g dry chi-square MC_o moisture content at initial time (g water/g dry Subscripts solids) initial MRmoisture ratio i specific time N number of model constants ththeoretical P microwave output power (W)

1. Introduction

Auto-oxidation of food during transportation, storage and processing is the primary mechanism for its physical (taste, color) and nutritional deterioration [12]. There is growing interest in the food science community on bioactive components such as natural antioxidants and their potential to replace synthetic compounds¹ in various food applications. Since time immemorial, we have looked toward plants as a source of inspiration and for harnessing their medicinal properties; indeed, their contributions toward improving human health have been substantial [24]. In this vein, both Adathoda vasica and Cymbopogon citratus are well-recognized plants in the Indian system of oriental medicine [8,3]. The contemporary remedial applications of A. vasica include the treatment of asthma, chronic bronchitis, rheumatic inflammatory swellings, antispasmodics, piles, cold [6,14]. Identified A. vasica alkaloids, primarily, vasicine and vasicinone have been acknowledged to be bioactive with their combination producing both in vitro and in vivo bronchodilatory activity commensurate to that of the ophylline [2,29]. Likewise, C. citratus (lemongrass) has been used in various pharmaceutical applications as spasmolytics, analgesics, antipyretics, and diuretics and in tranquilizers. Essential oil extract from lemongrass leaves has high citral content and is utilized as a raw material in the production of vitamin A, beta carotene, ionone, etc. [20].

In lieu of the shortcomings of conventional methods² toward bioactive component recovery from plants, research effort has been directed toward the applicability of newer techniques that utilize enzymes, microwaves, supercritical fluids and ultrasounds for assisting the extraction. In addition to their high safety-low cost aspect these processes also benefit from allowing extraction to occur without affecting the quality of the final product or degrading unsaturated compounds in the extract [4]. Particularly, Microwave Assisted Extraction

(MAE) has shown promise with several studies demonstrating its feasibility for the extraction of compounds from medicinal plants [15,5,7]. MAE enhances traditional solvent extraction by subjecting the solvent to rapid heat generation and direct interaction with electromagnetic radiation. The process results in the creation of a high pressure difference that causes the plant cellular structure to break and the solvent to penetrate within its matrix [22].

A comprehensive understanding of both, the drying and extraction steps is necessary to design the extraction process, avoid the denaturation of the product and determine feasible operating parameters. Given the limited availability of this literature for *A. vasica* and *C. citratus*, this study investigates the effect of various process parameters on the leaf extraction yield of bioactive compounds therein. An empirical correlation was developed for the examined drying parameters by improvising upon a selected conventional drying model. A comparative experimental procedure was followed to distinguish conventional Soxhlet extraction and MAE which were performed following the microwave drying of the leaves.

2. Materials and methods

Fresh *A. vasica* and *C. citratus* leaves were sourced from the VIT University campus at Vellore, India (12°55′12.79″N – 79°7′59.9″E; 216 m above sea level). The leaves were washed with distilled water and stored at *ca* 10 °C. Both plant specimens were validated by the Plant Biotechnology Division³ at VIT University, Vellore, India. Ten replicated measurements were performed to determine the leaf thickness using a micrometer (Leica stage micrometer-MA285, Germany) and initial moisture content by standard vacuum drying method. Leaf thickness was found to be 0.5 mm and 0.75 mm and moisture content was 70.72% and 68.75% (w.b.), for *A. vasica* and *C. citratus* respectively. All solvents and chemicals used in the study were purchased from SD Fine Chemicals, Mumbai, India, and were of analytical grade.

¹ *Inter alia*, synthetic antioxidants such as butylated hydroxyanisole have been banned in Japan and *tert*-butyl hydroquinone is no longer allowed for food applications in the EU and Canada.

² Such as hydrodistillation, enfleurage, maceration, pressing and distillation.

³ http://www.vit.ac.in/academics/schools/sbst

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