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### Identification of amino acids in mitochondrially encoded proteins that correlate with lifespan 2

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## ABSTRACT

Animals show a huge diversity in their lifespan that can vary from a few weeks to over a hundred years in verte- 22 brates. Size is a key element in this variation and the positive correlation between size and maximum lifespan can 23 be observed in each class of vertebrate. Some groups and species clearly stand out in this size-lifespan relation- 24 ship and the ones with exceptionally long lifespan have been studied to understand the biological causes of their 25 low aging rate. Among the potential explanations of animals' lifespan variations, mitochondria and 26 mitochondrially encoded genes have drawn attention because of their importance in the aging process. To under-27 stand both the extent of lifespan variations and their dependence to genes and amino acid variations in mito-28 chondrial genes and DNA (mtDNA), we analyze in a systematic way all 13 proteins encoded by mitochondria 29 in all vertebrates for which we had information on weight, maximum lifespan and mtDNA sequence. This com- 30 parison allows us to visualize positions, and even specific amino acids, in these sequences that correlate with 31 lifespan. With this approach, we draw a map of 356 amino acid residues, at 296 positions within the sequence, 32 that correlate with longer or shorter lifespan. We also compared this map with the human mitochondrial poly-33 morphism to determine its potential as a predictive tool. 34

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#### 1. Introduction 40

1.1. Mitochondria and its role in ATP and ROS production 41

Mitochondria are essential organelles of eukaryotic cells and have 42 been linked to aging and longevity in numerous publications (Balaban 43 44 et al., 2005; Harman, 1972; Loeb et al., 2005). They produce the energy of the cell by oxidative phosphorylation of ADP into ATP through the 45transfer of electron in the respiratory chain. An incomplete coupling in 46this process, mostly at the complex I of the electron transfer chain 4748 (Barja and Herrero, 1998), leads to electron leakage that can interact with oxygen to create superoxide radicals(Brand, 2000). Superoxide 49 dismutase can then turn superoxide into hydrogen peroxide, which 5051can lead to the production of hydroxyl radicals (Murphy, 2009). All these reactive oxygen species (ROS) are involved in the oxidation of 52all types of macromolecules in the cell and are believed to be one of 5354the main causes of aging (Barja, 2013; Beckman and Ames, 1998; 55Cadenas and Davies, 2000; Harman, 1972). They also damage

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mitochondrial proteins and membrane, leading to an increase in ROS 56 production (Remmen and Richardson, 2001). The implication of mito- 57 chondria in the regulation of apoptosis is also referred to play a key 58 role in the aging process of animals (Green and Reed, 1998). 59

### 1.2. Mitochondria, lifespan and comparative biology

Comparative studies of various vertebrates show that the efficiency 61 of the electron transfer chain (ETC) and the ROS production are tightly 62 linked with lifespan (Pamplona and Costantini, 2011). Longer living 63 species show a lower production of ROS, a lower activity of antioxidant 64 enzymes (Perez-Campo et al., 1998), a lower degree of fatty acid 65 unsaturation (Barja, 2002) and a lower amount of mtDNA oxidation 66 (Barja and Herrero, 2000). Considering that all the proteins encoded 67 by the mitochondrial genome are part of the ETC, the correlation of 68 lifespan with mitochondrial genome has been measured in various as- 69 pects. The composition of mtDNA and mitochondrially encoded pro-70 teins (MEPs) show the most significant correlation with longevity, 71 captured whether by the proportion of nucleotides in mtDNA (Jobson 72 et al., 2010; Lehmann et al., 2008; Min and Hickey, 2008) or the propor-73 tion of some amino acids (Lehmann et al., 2006; Moosmann, 2011; 74 Schindeldecker and Moosmann, 2013), especially methionine (Aledo 75 et al., 2012) and cysteine (Moosmann and Behl, 2008). These findings 76 show the strong role of MEPs in longevity but reduce mtDNA and 77

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Abbreviations: mtDNA, Mitochondrial DNA; MEPs, Mitochondrially encoded proteins; ROS, Reactive oxygen species; ACL (+ or -), Amino acid correlated with (longer or shorter) lifespan.

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MEPs to their composition in nucleotides and amino acids and are obliv ious to the primary and secondary structure of these MEPs.

## 80 1.3. Role of mitochondrial proteins' sequence in lifespan

Because of the importance of these proteins in ATP and ROS production, the amino acid sequence of the proteins encoded by mtDNA and their variations must play a role in the determination of lifespan. Such a relationship was shown by Rottenberg to depend on the rate of evolution for placental mammals (Rottenberg, 2007).

In our study, we have studied the evolution of MEPs and its correlation with longevity by comparing their amino acid sequences in vertebrates. Our goal was to identify the key positions and amino acids within the sequence of MEPs that play a role in aging and longevity and to chart a map of these amino acids.

### 91 2. Material and methods

### 92 2.1. Selection of species

We used the AnAge database (De Magalhães and Costa, 2009) 93 (genomics.senescence.info/species/), containing over 4000 species 94 95with information on both maximum lifespan (MLSP) and body mass (BM), and the PubMed database on mitochondrial genome se-96 quences with over 2000 vertebrate mtDNA sequences (www.ncbi. 97 nlm.nih.gov/genomes/OrganelleResource.cgi?opt=organelle&taxid= 98 7742), in order to obtain 324 species with all three information. These 99 100 species were tested to be significantly representative in terms of MLSP/BM distribution compared with the 4000 vertebrate species 101 present in AnAge (data not shown). 102

### 103 2.2. Phylogenetic analyses

The 13 protein-coding genes (A6, A8, C1, C2, C3, Cb, N1, N2, N3, N4, 104 N4L, N5, N6) of the mitochondrial genomes of the 324 species were 105aligned separately using the muscle (version 3.7, default parameters) 106 (Edgar, 2004). Most genes were present in all species (N2 was missing 107 from Oncorhynchus kisutch, N5 from Sphenodon punctatus and N6 108 from Castor canadensis). Alignments were both analyzed separately 109and concatenated in a supermatrix resulting in a total of 14 alignments. 110 The supermatrix had 4005 columns, corresponding to as many amino 111 112 acids. The best evolution model was selected for each alignment using Prottest v3.0 (Darriba et al., 2011). The mitochondrial replacement ma-113 trix MtMAM + I + G was strongly favored for all genes but N6 for all 114 criteria (AIC, BIC and variations). The posterior probability was even 115 close to 1.00 for 9 of the 13 (A6, C1, C2, C3, Cb, N1, N2, N4, N5). In 116 117 each case, the second best model was MtMAM + G. For N6, the favored model was JTT + I + G, with only contender JTT + G (respective poste-118 rior probabilities: 0.85/0.15 for AIC, 0.53/0.47 for BIC). Fourteen phylo-119genetic trees were reconstructed using PhyML v3.0 (Guindon et al., 1202010) with best of SPR and NNI moves, 5 random starting trees and 121122100 bootstrap replicates. We performed further analyses with the tree 123reconstructed from the concatenated alignment named T hereafter, as it minimized the sum of distances to the other trees. 124

## 125 2.3. Phylogenetic regression

To investigate the association between particular alignment sites and lifespan while correcting for body mass, we performed a phylogenetic regression of Lifespan against Body mass and amino acid distribution at that site. Namely, for each amino acid *aa* at each site *i*, we constructed a presence/absence vector of  $X_{aa,i}$  of that amino acid at that site and estimated the linear model:

 $log(Lifespan) \sim \ log(Body\,mass) + X_{aa,i} + \epsilon$ 

where  $\varepsilon$  is the centered Gaussian with covariance matrix  $\sigma^2 \Sigma(T)$  where133 $\sigma^2$  is unknown and  $\Sigma(T)$  is completely specified by T, as the covariance134matrix of a Brownian motion running on T (Felsenstein, 1985). Lifespan134and Body Mass were log-transformed to account for well-described al-135lometric relations between the two.136

This procedure resulted in 4005 \* 20 = 80,100 linear models and as 137 many coefficients  $a_{aa,i}$  to estimate. The coefficients were estimated 138 using the compar.gee function of the R ape package (version 2.8) 139 (Paradis et al., 2004). Visual inspection of the p-values did not show 140 signs of significant correlations between the tests and we therefore 141 used FDR correction (Benjamini and Hochberg, 1995) to correct for 142 multiple testing, resulting in a total of 356 significant coefficients with 143 a p-value under 0.01 [Supplementary Table 1]. 144

3. Results

### 3.1. Map of ACL

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As previously described (De Magalhães et al., 2007; Speakman, 147 2005), the maximum lifespan of vertebrates, with or without correction 148 for their body mass, has a log-normal distribution. Each class of verte-149 brates shows a similar distribution with a class-specific mean and vari-150 ance. Based on this observation, it is possible to compare the maximum 151 lifespan distribution of vertebrates for every amino acid at every posi-152 tion in the 13 MEPs. We could therefore estimate the extent to which 153 these amino acid residues are related to lifespan and draw a map of 154 the Amino acids Correlated with maximum Lifespan (ACL) whether 155 positively (ACL+) or negatively (ACL-). In Fig. 1, we illustrate by a 156 hypothetic example the way we have selected these ACL. This illustra-157 tion doesn't take into account the correction for phylogeny that defines 158 the p-value and correlation coefficient of each ACL.

The multiple alignments of the concatenated MEP sequences of 324 160 vertebrates give a matrix of 4005 positions [Supplementary Table 1]. 161 Among those, 296 positions have at least one amino acid that correlates, 162 positively or negatively, with maximum lifespan for a total of 356 ACL, 163 206 ACL + and 150 ACL -. There are 242 positions in the alignment se- 164 quence with a single ACL, 48 with 2 ACL and 6 with 3 ACL at a single po- 165 sition. Taken together, these ACL draw a map [Fig. 2] of the influent of **Q3** 



**Fig. 1.** Longevity distribution centered around 0 after correction for body mass. In this fictitious example, species are grouped by amino acid at a given position. The distribution of longevity for all species is featured in purple. The corresponding distribution for species exhibiting the consensus amino acid (green) is also centered around 0. By contrast, species with the ACL + amino acid (red) have a right-shifted distribution and thus longer lifespan. Similarly, species with the ACL – amino acid (blue) have a left-shifted distribution and thus longer lifespan. (For interpretation of the references to color in this figure legend, **Q2** the reader is referred to the web version of this article.)

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