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We asked if this association was also observed in a large French ALS population. We also studied the possible relation of *HFE* polymorphisms with the clinical characteristics of the disease.

2. Methods

2.1. Subjects

Clinical data and blood samples were obtained from 824 patients with SALS and 447 controls. The diagnosis of sporadic ALS was made in one of the French ALS Study Group's centres. All patients met the El Escorial World Federation criteria for the diagnosis of definite or probable ALS [20]. For each patient, clinical data comprised gender, site-of-onset (limb or bulbar), age-at-onset and, if available, disease duration. The site-of-onset was defined as either bulbar or limb-onset. Bulbar-onset was defined as symptoms first occurring at the bulbar level with dysphagia, dysphonia or dysarthria. Limb-onset was defined as symptoms first occurring in the limbs. The age-at-onset was defined as the time at which first motor weakness was reported by the patient. The duration of ALS was defined as the interval between onset (first symptoms) and death or tracheotomy. Controls were matched for age, sex, and ethnicity. All participants gave written informed consent.

In order to enlarge the control population, we analysed also French controls from the literature. We used the review by Merryweather-Clarke et al. [21] to select five French series and pooled controls from these series.

2.2. Genotyping

DNA was extracted by standard procedures from peripheral blood. PCR reactions were carried out in a total of 50 µl containing 200 ng of genomic DNA, 10 pmol of each primers for C282Y or 20 pmol of each primers for H63D, 1.5 mM MgCl₂, 200 µM of dNTPs, and 1.75 U AmpliTaq polymerase (PROMEGA). The PCR conditions were 1 cycle at 92 °C for 5 min, 30 cycles at 92 °C for 1 min, 56 °C for 1 min, 72 °C for 1 min, and 1 cycle at 72 °C for 7 min. Amplification primers were as follows : C282Y, forward primer 5'-TGGCAAGGGTAAACAGATCC-3', reverse primer 5'-CTCAGGCACTCTCTCAACC-3'; H63D, forward primer 5'-ACATGGTTAAGGCCTGTTGC-3', reverse primer 5'-GCCACATCTGGCTTGAAATT-3'. The *HFE* polymorphisms were detected in PCR products by restriction enzyme digestion with Rsa1 for C282Y (ROCHE Diagnostics) and Mbo1 for H63D (PROMEGA France) [22]. Restriction reactions were carried out for 3 h at 37 °C using 30 µl of the PCR products and 15 U of Mbo1 or 12 U of Rsa1. The restriction fragments were run on a 2% resophor gel. The 387-bp PCR product digested with Rsa1 showed two fragments of 247- and 140-bp in normal DNA. The C282Y mutation created a new Rsa1 restriction site and the 387-bp PCR product digested showed three fragments of 247, 111 and 29 bp in mutated DNA. The 208-bp PCR product of normal DNA was cut into two fragments of 138 and 70 bp with Mbo1 digestion. The H63D mutation destroyed the Mbo1 site and the mutated DNA was not cut by the restriction enzyme.

2.3. Statistical analysis

Analysis of *HFE* polymorphisms was performed by an investigator blind to the disease status (ALS or control). Allele and genotype frequencies were counted and tested for Hardy–Weinberg equilibrium. A correction for multiple tests was applied to adjust the p values by accounting for the 2 loci (C282Y and H63D) analysed in the study. Differences were considered as significant when $p < 0.025$.

We compared the frequencies of alleles and genotypes of the two polymorphisms in controls and patients using the χ^2 test. We also compared the frequency of the *HFE* polymorphisms in our controls group to the control group from literature using χ^2 test [21].

Table 1
Genotype and allele frequency data for the C282Y polymorphism.

	Controls		Patients	
	Frequency (%)	Numbers	Frequency (%)	Numbers
<i>Genotype</i>				
CC	88.4	395	93.7	772
CY	11.4	51	6.2	51
YY	0.2	1	0.1	1
Total	100	447	100	824
<i>Allele</i>				
C	94.1	841	96.8	1595
Y	5.9	53	3.2	53
Total	100	894	100	1648

Chi-square test for genotypes (grouping CY and YY): $\chi^2 = 10.927$; $p = 0.001$ (odds ratio: 1.95 [IC 95%: 1.3–2.92]); chi-square test for alleles: $\chi^2 = 10.671$; $p = 0.001$ (odds ratio: 1.89 [IC 95%: 1.29–2.81]).

Haplotypes of the two variants (C282Y and H63D) were calculated using a maximum likelihood algorithm [23, 24]. We also tested the distribution of the two polymorphisms according to the presence of the APOE $\epsilon 4$ allele [25], in all patients and in each gender group.

t-tests evaluated the influence of gender, site of onset and both polymorphisms on the age-of-onset. We evaluated the distribution of genders, site-of-onset and age-of-onset between D allele carriers and non carriers and between Y carriers and non carriers using χ^2 tests or t-tests. Kaplan Meier methods estimated survival curves. Comparison of survival data between groups defined by gender, site-of-onset, and *HFE* variants and according to age-of-onset was performed using the log-rank test or Cox analysis when appropriate. The different factors having a significant influence on survival were explored with multivariate Cox proportional hazard models. We included only patients without missing data in all statistical survival analyses and $p < 0.05$ was considered as significant. The power of this study was sufficient to detect a difference of 5% between controls and patients, accepting an alpha risk of 5% and a beta risk of 20%. Statistical analysis was performed with JMP statistical software version 7.0.2 (SAS Institute, Cary, North Carolina).

3. Results

Gender, site-of-onset and age-at-onset were known for 758 patients (92%), female represented 41.4% ($N = 314$) of the SALS group and bulbar-onset was determined in 28.9% ($N = 219$). Median age-at-onset was 61.4 [20.1–89.1] years. The mean age-at-onset for bulbar-onset patients was significantly higher than the age-at-onset for limb-onset patients (mean: 64.4 ± 11.0 years vs 58.1 ± 12.9 years respectively; $p < 0.0001$) and higher in female (61.9 ± 12.1 years vs 58.4 ± 12.9 years; $p = 0.0002$).

Table 2
Genotypes and allele frequency data for the H63D polymorphism.

	Controls		Patients	
	Frequency (%)	Numbers	Frequency (%)	Numbers
<i>Genotype</i>				
HH	69.8	312	73.0	601
HD	27.3	122	25.2	208
DD	2.9	13	1.8	15
Total	100	447	100	824
<i>Allele</i>				
H	83.4	746	85.6	1410
D	16.6	148	14.4	238
Total	100	894	100	1648

Chi-square test for genotypes: $\chi^2 = 2.423$; $p = 0.29$; chi-square test for alleles: $\chi^2 = 2.009$; $p = 0.16$.

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