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Antigen-specific CD4⁺ effector T cells: Analysis of factors regulating clonal expansion and cytokine production Clonal expansion and cytokine production by CD4⁺ effector T cells

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ABSTRACT

In order to fully understand T cell-mediated immunity, the mechanisms that regulate clonal expansion and cytokine production by CD4⁺ antigen-specific effector T cells in response to a wide range of antigenic stimulation needs clarification. For this purpose, panels of antigen-specific CD4⁺ T cell clones with different thresholds for antigen-induced proliferation were generated by repeated stimulation with high- or low-dose antigen. Differences in antigen sensitivities did not correlate with expression of TCR, CD4, adhesion or costimulatory molecules. There was no significant difference in antigen-dependent cytokine production by TG40 cells transfected with TCR obtained from either high- or low-dose-responding T cell clones, suggesting that the affinity of TCRs for their ligands is not primary determinant of T cell antigen reactivity. The proliferative response of all T cell clones to both peptide stimulation and to TCR β crosslinking revealed parallel dose-response curves. These results suggest that the TCR signal strength of effector T cells and threshold of antigen reactivity is determined by an intrinsic property, such as the TCR signalosome and/or intracellular signaling machinery. Finally, the antigen responses of high- and low-peptide-responding T cell clones reveal that clonal expansion and cytokine production of effector T cells occur independently of antigen concentration. Based on these results, the mechanisms underlying selection of high "avidity" effector and memory T cells in response to pathogen are discussed.

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The activation and differentiation of effector T cells in response to antigenic stimulation is distinct from that of naïve T cells [1,2]. During the course of the first infection, naïve T cells are primed with pathogen-derived peptides presented in the context of MHC by professional APC. These Ag-primed effector cells proliferate and differentiate into effector cells in regional lymph nodes [3]. Consequently, this process induces a set of pathogen-specific effector T cells that differ in their sensitivity and reactivity, or "avidity", to pathogen-derived peptides. These effector T cells then immigrate to the infection site, where they interact with target cells [4].

At the site of the infection, pathogens are not uniformly distributed; that is, the amount of antigen presented by individual APC to immigrated pathogen-reactive T cells is not uniform. Furthermore, antigenic stimulation by pathogen-presenting APC to T cells is dynamic during the course of infection [5]. Under these circumstances, pathogen-specific effector T cells, whose sensitivities and reactivities to pathogen epitopes are heterogeneous, encounter different strengths of antigen stimulation. In order to fully understand T cell-mediated immunity, analysis of altered T cell effector function in an environment of changing antigen load is important [6].

The goal of the present report is to determine the mechanisms that regulate clonal expansion and cytokine production by CD4⁺ antigenspecific effector T cells in response to a wide range of antigenic stimulation. To achieve this, we have generated a panel of OVA-specific T cell clones whose optimal proliferative responses occur at peptide concentrations that vary between 3.9 and 1000 nM. The expression levels of TCR^β, CD4 or cell surface adhesion and costimulatory receptors on these T cells does not correlate with their peptide dose-dependent proliferative response. When TCR $\alpha\beta$ genes were cloned from high- or lowdose-responding T cell clones and expressed on TCR-negative TG40 T cells, there was no significant difference in their reactivity to various concentrations of OVA-peptide. This result suggests that differences in affinities of TCRs used by individual T cell clones are not responsible for differences in peptide-dose-dependent proliferation. In addition, the proliferative responses of these clones to TCR crosslinking with anti-TCR antibody parallel their response to antigen stimulation. These results strongly suggest that the sensitivity and reactivity of CD4⁺ effector T cells to their antigenic targets may be largely determined by intrinsic properties of the effector T cell. Furthermore, while high

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"avidity" effector T cell clones reveal high-antigen-dose suppression, resulting in a biphasic proliferation dose-response curve, their cytokine production pattern was monophasic. Thus these results support a model in which clonal expansion and cytokine production by effector T cells is regulated independently at various antigen conditions.

Materials and methods

Preparation and assay of OVA_{323–339}-specific T cell lines and clones. BALB/c mice were immunized in the footpad with 100 mg (56 nM) OVA323-329 peptide in CFA. After two weeks, inguinal and popliteal lymph nodes were harvested from 2 mice and were cultured with serial dilutions of OVA₃₂₃₋₃₂₉ (from 0.97 to 1000 nM). Viable T cells were collected weekly and re-stimulated with irradiated (30 Gy) BALB/c spleen cells and the same concentration of OVA323-329 used in the initial culture. Three independent antigen-specific T cell lines were generated and were cloned by limiting dilution after 2, 4, and 8 restimulations, respectively. For their proliferative response to antigen, 2×10^5 cells were incubated with titrated amounts of OVA peptide and 5×10^5 irradiated BALB/c splenocytes for 51 h at 37 °C. For anti-TCR mAb crosslinking, 2×10^5 cells were stimulated in wells pre-coated with 50 ml of serially diluted TCRβspecific mAb and cultured for 36 h at 37 °C. [³H]TdR was added to the cultures for the last 12 h of incubation. For IFN- γ production, supernatants were removed from cultures 48 h after stimulation; IFN- γ was quantified by ELISA.

Flow cytometry. Immunofluorescence staining was carried out as previously described [7]. Fluorescent-conjugated Abs specific for TCR β , CD4, CD28, LFA-1, ICOS, CTLA4, 4-1BB, and OX40 purchased from PharMingen (San Diego, CA. USA). Cytometric analyses

were performed on a FACSCalibur using CellQuest software (Becton Dickinson, Mountain View, CA. USA).

Isolation of full-length TCR α and β chains from CD4⁺ T cell clones and transfection into TCR-negative T cell hybridoma. cDNA gene encoding the TCR α and TCR β chains from individual CD4⁺ T cell clones were cloned, sequenced, and synthesized as previously described [8]. Products were cloned into the multi-cloning site of pMX-IRES-GFP [9]. The TCR-negative T cell hybridoma, TG40 was retrovirally transduced as previously described [10]. Infected cells were stained with PE-labeled TCR β mAb, and the positive fraction (TCR⁺, GFP⁺) was sorted on a FACSVantage (BD Biosciences). For activation of infected TG40 cells, 1 × 10⁵ cells were incubated with titrated amounts of OVA peptide and 5 × 10⁵ irradiated T cell-depleted BALB/c splenocytes for 24 h. Supernatants were then collected, and IL-2 production was quantified using the IL-2 indicator cell, CTLL-2.

Preparation of OVA₃₂₃₋₃₃₉/I-A^d tetramers. OVA₃₂₃₋₃₃₉/I-A^d tetramers were prepared using baculovirus as previously described [11]. For proliferative responses to OVA₃₂₃₋₃₃₉/I-A^d tetramer, 2×10^4 cells were added to wells pre-coated with serially diluted OVA₃₂₃₋₃₃₉/I-A^d tetramer. Cells were cultured for 51 h at 37 °C, [³H]TdR was added for the last 12 h of incubation.

Results and discussions

Generation of T cell clones responding to different concentrations of OVA-peptide

The proliferative responses of lymph-node (LN) cells from OVAprimed mice were assayed by stimulating with titrated OVA-peptide. As shown in Fig. 1A, these cells proliferated to peptide stimu-



Fig. 1. Proliferative responses of OVA-primed LN cells and OVA-specific T cell lines and clones. (A) Proliferative responses of OVA-primed BALB/c lymph node cells to serial titrations of OVA peptide. (B) Proliferative responses of long-term T cell lines, which were generated by restimulation for 4 times *in vitro* with 1000, 250, 62.5, or 3.9 nM OVA peptide. (C) Proliferative responses of T cell clones obtained from long-term T cell lines by limiting dilution. Bars represent group means ± SD. Similar results were obtained for three additional experiments.

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