



Review

A combined literature and in silico analysis enlightens the role of the NDRG family in the gut

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ABSTRACT

Background: The N-Myc Downstream-Regulated Gene (*NDRG*) family comprises four members that function in cellular processes like proliferation and differentiation. While *NDRG1* and *NDRG2* are extensively studied, knowledge regarding *NDRG3* and *NDRG4*, despite its recognition as a well-established early-detection marker for colorectal cancer (Cologuard®), is sparse.

Scope of review: To summarize expression, biomarker potential and functional mechanisms of the *NDRGs* in the developing, mature and cancerous gut, we combine current literature and *in silico* analyses from the TCGA-database, GTEx Project, E14.5 mouse intestine and enteric neural crest cells, and an RNA-sequencing time-series of human embryonic colonic samples.

Major conclusions: This study reveals that all members display a differential expression pattern in the gut and that *NDRG1*, *NDRG2* and *NDRG4* (1) can serve as biomarker for colorectal cancer and (2) have tumor suppressive properties mainly affecting cell proliferation and epithelial-mesenchymal transition.

General significance: Similar effects of the *NDRGs* on the key-hallmarks of cancer, could implicate analogous functions in other tissue/cancer types.

1. Introduction

The N-myc downstream-regulated gene (*NDRG*) family is composed of four members: *NDRG1*, *NDRG2*, *NDRG3* and *NDRG4*, and owes its name to the discovery of the first member, *NDRG1*, as being repressed by the *C-Myc* and *N-Myc* oncogenes [1]. In humans and mice, the four genes are all located on different chromosomes. Various aliases have been designated to each family member and each gene is transcribed into multiple alternatively spliced mRNA transcripts that encode several protein isoforms [2]. The encoded proteins of this family are all characterized by an NDR region and an α/β hydrolase fold, but lack the

catalytic motif required to be enzymatically active [3,4]. The *NDRG* proteins which share about 52-65% sequence homology with an identical C-terminal sequence “MEVSC” and only very few sequence differences primarily in the C- and N-terminal regions, have been shown to be highly conserved in a variety of species [5,6]. More detailed information about the general knowledge on the *NDRG* family members, with respect to structure, origin and signaling in physiological processes can be found in our former review by Melotte et al. [2].

Previously, we and various independent groups identified *NDRG4* promoter CpG island methylation in fecal DNA as an accurate early-detection marker for colorectal cancer (CRC) [7–16]. The

Abbreviations: BDM1, Brain developmental-related molecule 1; Cav-1, Caveolin-1; CRC, Colorectal cancer; DAC, 5-aza-2'-deoxycytine; DFS, Disease free survival; EMT, Epithelial-mesenchymal transition; ENCC, Enteric neural crest cells; ENS, Enteric nervous system; EW, Embryonic week; FDR, False discovery rate; FIT, Fecal immunochemical test; GEO, Gene expression omnibus; GI, Gastrointestinal; GTEx, Genotype-tissue expression project; HMSNL, Hereditary motor and sensory neuropathy-Lom; IBD, Inflammatory bowel disease; LN, Lymph node; *NDRG*, N-myc downstream-regulated gene; OS, Overall survival; RPKM, Reads per kilobase million; SMAP8, Smooth muscle-associated protein 8; SNARE, Soluble NSF attachment protein receptor; SNP, Single nucleotide polymorphism; TSA, Trichostatin A; TCGA, The cancer genome atlas; TSS, Transcription start site; UC, Ulcerative colitis; VAMP, Vesicle-associated membrane protein

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biomarker performance of *NDRG4* was further exploited by Exact Sciences, who integrated *NDRG4* promoter methylation into a multi-target stool DNA test: Cologuard® (Madison, USA) [16]. This FDA-approved stool DNA test detects significantly more CRCs and advanced precancerous lesions than the leading fecal immunochemical test (FIT) and is currently used in the USA for CRC screening [7]. Despite being incorporated into the Cologuard®, almost nothing is known about the function(s) of *NDRG4* in the (diseased) intestinal tract. Whereas *NDRG1* and *NDRG2* are important in various cellular processes (e.g. proliferation, differentiation, apoptosis), little is known about *NDRG3* and *NDRG4*.

Here, we performed a literature search (Pubmed, Web of Science, Embase and Medline) and supported this literature search with *in silico* data extracted from (i) publicly available transcriptome databases [17,18], (ii) previously generated transcriptome data of E14.5 mouse intestine and enteric neural crest cells (ENCCs) [19] and (iii) an RNA-sequencing time-series of human embryonic colonic samples, to gain more insights into the expression pattern, biomarker potential and common pathways of the *NDRGs* in the intestinal tract.

Our review indicates that *NDRG1*, *NDRG2* and *NDRG4* can act as biomarkers for CRC. Although there is no evidence that the different family members are major drivers in the development of CRC, data suggest that, except for *NDRG3*, each *NDRG* member has a protective role during intestinal carcinogenesis.

2. Expression pattern of the *NDRGs* in the developing and adult gut

To understand the functional importance of the *NDRG* family members in the intestinal tract, we first elaborate on their structure and summarize their (overlapping) intestinal-specific expression pattern in mammalian species like rat, mouse and human.

In humans, *NDRG1* is located on chromosome 8q24.3, while the mouse ortholog is located on chromosome 15D2. The encoded canonical protein sequence of human *NDRG1* (*DRG1*, *RTP*, *RIT42*, *CAP43*) consists of 394 amino acids (AA), resulting in a 43 kDa protein. The two other isoforms are made up of 328 and 313 AA, with a respective mass of 35.5 and 34 kDa. In mouse and rat, only one isoform has been found, with a similar length as the human canonical protein sequence and minor AA differences.

The expression of the *NDRG1* gene usually coincides in tissues where *N-myc* is expressed, but only initiates at E9.5 as the intestinal epithelium begins to differentiate and *N-myc* expression decreases [20]. As further intestinal development requires retinoic acid-dependent activation of *NDRG1* and suppression of the *Wnt*/ β -catenin pathway [21], *NDRG1* mRNA expression steadily increases in the mouse and human intestinal epithelium from E10.5 onwards [20]. Further intestinal maturation in rodents and humans seems to be controlled by circulating levels of glucocorticoids and cortisol, respectively [22], giving rise to similar levels of *NDRG1* in the adult small and large intestine [5,22–24]. Moreover, the expression of *NDRG1* mRNA and protein is induced by glucocorticoid treatment (i.e. by dexamethasone) and strongly overlaps with a gradual increase from crypt cells towards surface cells [20,22,24,25]. On a sub-cellular level, *NDRG1* is primarily localized in the cytoplasm of epithelial cells, where it is often associated with the basolateral membrane adjacent to adherens junctions and desmosomes [20,24,26].

NDRG2 is located on chromosome 14q11,2 and 14C1 in human and mouse, respectively. The main functional form of human, mouse and rat *NDRG2* (*SYLD*, *NDR2*) is either a 41 kDa protein, composed of 371 AA, or a 39 kDa protein of 357 AA. In addition, there are four other human isoforms with a length ranging from 328 to 367 AA and a variable mass between 36 and 40.3 kDa.

In early developmental stages (E8.5–9.5) the poorly differentiated intestinal epithelium expresses low levels of *NDRG2* mRNA and protein. From E10.5 towards adulthood, the levels of *NDRG2* mRNA and protein

very slightly increase in the mucous forming, lining epithelial cells [20,27], leading to barely detectable levels in adult gut tissue. Even though most studies observed a slightly higher level of *NDRG2* mRNA in the (distal) colon compared to the small intestine [5,28,29], contradictory observations regarding *NDRG2* protein expression have been described. Yamamoto et al. detected a marginally higher signal in colon than in the small intestine [30], whereas Hu et al. detected opposite results with weak to moderate *NDRG2* immunoreactivity in the small intestine but not in the colon [31].

Human chromosome 20q11.21–11.32 and mouse chromosome 2H1 contain the *NDRG3* gene, which can be transcribed into three identified protein isoforms. The canonical sequence of *NDRG3* (no known aliases) encodes a 41.5 kDa protein of 375 AA in various mammalian species. The second and third isoform are only found in humans and encode a 40 kDa protein of 363 AA and a markedly lighter protein (31.5 kDa) of only 286 AA.

Only three studies include a brief description about the expression of *NDRG3*. In fact, *in situ* mRNA hybridization (ISH) and Northern Blotting reveal that *NDRG3* expression is already activated at E9.5 and that *NDRG3* is widely expressed during embryogenesis. However no *NDRG3* mRNA is detected in the developing intestinal tract [20]. During further development, overall *NDRG3* mRNA and protein levels increase slightly, although they are hardly detectable in matured mouse and human gut tissues [5,30].

The *NDRG4* gene is situated on human chromosome 16q21–q22.1 and mouse chromosome 8D1. For *NDRG4* (*SMAP8*, *BDM1*), the canonical protein sequence is referred to as *NDRG4B^{var}*, consisting of 352 AA (39 kDa). The two other main isoforms are *NDRG4B* and *NDRG4H* which contain 339 AA (37 kDa) and 371 AA (41 kDa), respectively. In addition, 5 other isoforms can be produced by alternative splicing, with sequences ranging from 339 to 391 AA (37–43 kDa). In mouse, we recently identified three isoforms, corresponding to human *NDRG4B^{var}* (long isoform), *NDRG4B* (short isoform) and *NDRG4H* [32]. Furthermore, rat tissues express an additional fourth isoform of 45kDa, encoded by the rat ortholog: *SMAP8/BDM1* [33].

So far, the pattern of *NDRG4* expression during embryonic gut development has not yet been described. In adult tissues however, current literature is inconsistent regarding the intestinal-specific expression pattern of *NDRG4* which can be attributed to the use of different, a-specific commercially-available antibodies [32]. We and Chu et al. previously described that *NDRG4* is expressed within epithelial cells of the gut [12,34] and Qu et al. found high *NDRG4* protein levels in smooth muscle cells of the stomach [35]. However, we recently observed that the anti-*NDRG4* antibody used in these studies was not specifically targeting *NDRG4* [32]. The experimental application of an antibody that specifically targets *NDRG4* has demonstrated that *NDRG4* expression in the gut is restricted to the cytoplasm of neuronal cell bodies and nerve fibers belonging to the nervous system of the gut, i.e. the enteric nervous system (ENS) [32].

2.1. *In silico* expression analysis

To clarify the discrepancies concerning the *NDRG* expression pattern in the developing/mature gut observed in the limited number of publications, we evaluated expression data from the (i) GTEx Project (06/19/2017), (ii) previously generated transcriptome data of E14.5 mouse intestine and ENCCs (available in the Gene Expression Omnibus (GEO); GSE34208) [19] and (iii) an RNA seq time-series of human embryonic full-thickness samples of proximal colon (Embryonic Week (EW) 12, EW14 and EW16; data generated by the group of Prof. Thapar (UCL, London) and deposited to GEO).

Compared to its family members, *NDRG1* is highly expressed in EW12, EW14 and EW16 human colonic samples, with levels (in reads per kilobase million, RPKM) being 10, 8 and 12 times higher than for *NDRG2*, 3 and 4, respectively (Fig. 1A). In addition, *NDRG1* shows the highest and intercomparable expression in sections of the mature small

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