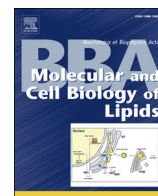




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## 1 Review

Q1 The impairment of cholesterol metabolism in Huntington disease<sup>☆</sup>Q2 Valerio Leoni<sup>a,b,\*</sup>, Claudio Caccia<sup>b</sup>4 <sup>a</sup> Laboratory of Clinical Chemistry, Ospedale Causa Pia Luvini, Cittiglio, AO Ospedale di Circolo e Fondazione Macchi, Varese, Italy5 <sup>b</sup> Laboratory of Clinical Pathology and Medical Genetics, Foundation IRCCS Institute of Neurology Carlo Besta, Milano, Italy

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## A B S T R A C T

Huntington disease (HD), an autosomal dominant neurodegenerative disorder caused by an abnormal expansion 18 of CAG trinucleotide repeat in the *Huntingtin* (*HTT*) gene, is characterized by extensive neurodegeneration of 19 striatum and cortex and severe diffuse atrophy at MRI. 20

The expression of genes involved in the cholesterol biosynthetic pathway and the amount of cholesterol, 21 lanosterol, lathosterol and 24S-hydroxycholesterol were reduced in murine models of HD. In case of HD- 22 patients, the decrease of plasma 24OHC follows disease progression proportionally to motor and neuropsychiatric 23 dysfunction and MRI brain atrophy, together with lanosterol and lathosterol (markers of cholesterol synthesis), 24 and 27-hydroxycholesterol. A significant reduction of total plasma cholesterol was observed only in advanced 25 stages. 26

It is likely that mutant *HTT* decreases the maturation of SREBP and the up-regulation LXR and LXR-targeted genes 27 (SREBP, ABCG1 and ABCG4, HMGCoA reductase, ApoE) resulting into a lower synthesis and transport of choles- 28 terol from astrocytes to neurons via ApoE. In primary oligodendrocytes, mutant *HTT* inhibited the regulatory 29 effect of PGC1 $\alpha$  on cholesterol metabolism and on the expression of MBP. 30

*HTT* seems to play a regulatory role in lipid metabolism. The impairment of the cholesterol metabolism was 31 found to be proportional to the CAG repeat length and to the load of mutant *HTT*. A dysregulation on PGC1 $\alpha$  32 and mitochondria dysfunction may be involved in an overall reduction of acetyl-CoA and ATP synthesis, contrib- 33 uting to the cerebral and whole body cholesterol impairment. This article is part of a Special Issue entitled Brain 34 Lipids 35

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## 1. Cholesterol

Q3

Cholesterol is a structural element of mammal cellular membrane 42 and regulates the fluidity of lipid bilayers. It is the precursor of bile 43 acids, steroid hormones and oxysterols. The cellular needs are covered 44 by de novo synthesis or by the uptake from circulating lipoproteins. 45 All the cells are able to synthesize, release and take up cholesterol to 46 maintain their cholesterol homeostasis: some produce an excess of 47 cholesterol to provide other cells, some others need exogenous choles- 48 terol because of limited synthetic capacity. In humans, under normal 49 conditions, about the 60% of the body's cholesterol is synthesized 50 (about 700 mg/day) and the remaining is provided by the diet. Choles- 51 terol, together with the other lipids, is absorbed by small intestine, 52 loaded on chylomicrons and delivered to the liver. The exogenous cell 53 supply is covered by very low density lipoprotein (VLDL)–low density 54 lipoprotein (LDL) cycle. Since an excess of free cholesterol is toxic to 55 the cells, a number of strategies have been evolved either to export 56 it (via lipoproteins), to store it in an esterified form or release it after 57 oxidation into oxysterols. 58

A major fraction of the exceeding is exported by the reverse choles- 59 terol transport mechanism involving the high density lipoprotein (HDL) 60 and the ATP-binding cassette (ABC)-transporter family. 61

**Abbreviations:** 24OHC, 24S-Hydroxycholesterol; 27OHC, 27-Hydroxycholesterol; ABC, ATP-binding cassette transporter; ACAT, Acyl-Coa:cholesterol acyltransferase; AD, Alzheimer disease; ApoE, Apolipoprotein E; BACHD, Bacterial artificial chromosome HD; BDNF, Brain-derived neurotrophic factor; Cav1, Caveolin-1; CNS, Central nervous system; CSF, Cerebrospinal fluid; CYP7A1, Cholesterol 7 $\alpha$ -hydroxylase; CYP27A1, Sterol 27-hydroxylase; CYP46A1, Cholesterol 24-hydroxylase; CYP51, Lanosterol 14- $\alpha$  demethylase; DHCR24, 24-Dehydrocholesterol reductase; ER, Endoplasmic reticulum; HD, Huntington disease; HDL, High density lipoproteins; HMGCoA, 3 $\alpha$ -Hydroxy-3-methylglutarylcoenzyme A; HMGCoAR, HMGCoA reductase; HMGCoAS, HMGCoA synthetase; *HTT*, *Huntingtin*; *Insig*, Insulin induced gene; LDL, Low density lipoproteins; LDL-R, LDL-receptor; LRP, LDL-related protein; LXR, Liver X receptor; MBP, Myelin basic protein; MM, Mitochondrial membrane; MRI, Magnetic resonance imaging; MS, Multiple sclerosis; NPC, Niemann–Pick type C; NS, Neural stem; PGC1 $\alpha$ , Peroxisome proliferator-activated receptor-gamma co-activator 1 alpha; PLP, Proteolipid protein; PPAR $\gamma$ , Peroxisome proliferator-activated receptor gamma; SCAP, SREBP cleavage-activating protein; ST, Striatum; TCA, Tricarboxylic acid; SRE, Sterol responsive element; SREBP, Sterol responsive element binding protein; VLDL, Very low density lipoprotein; YAC, Yeast artificial chromosome

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\* Corresponding author at: Laboratory of Clinical Chemistry, Ospedale Causa Pia Luvini, AO Ospedale di Circolo e Fondazione Macchi, via Luvini 21033 Cittiglio (VA). Tel.: +39 0332 607285, +39 333 3818519 (mobile).

E-mail addresses: [valerioleoni@hotmail.com](mailto:valerioleoni@hotmail.com), [valerio.leoni@ospedale.varese.it](mailto:valerio.leoni@ospedale.varese.it) (V. Leoni).

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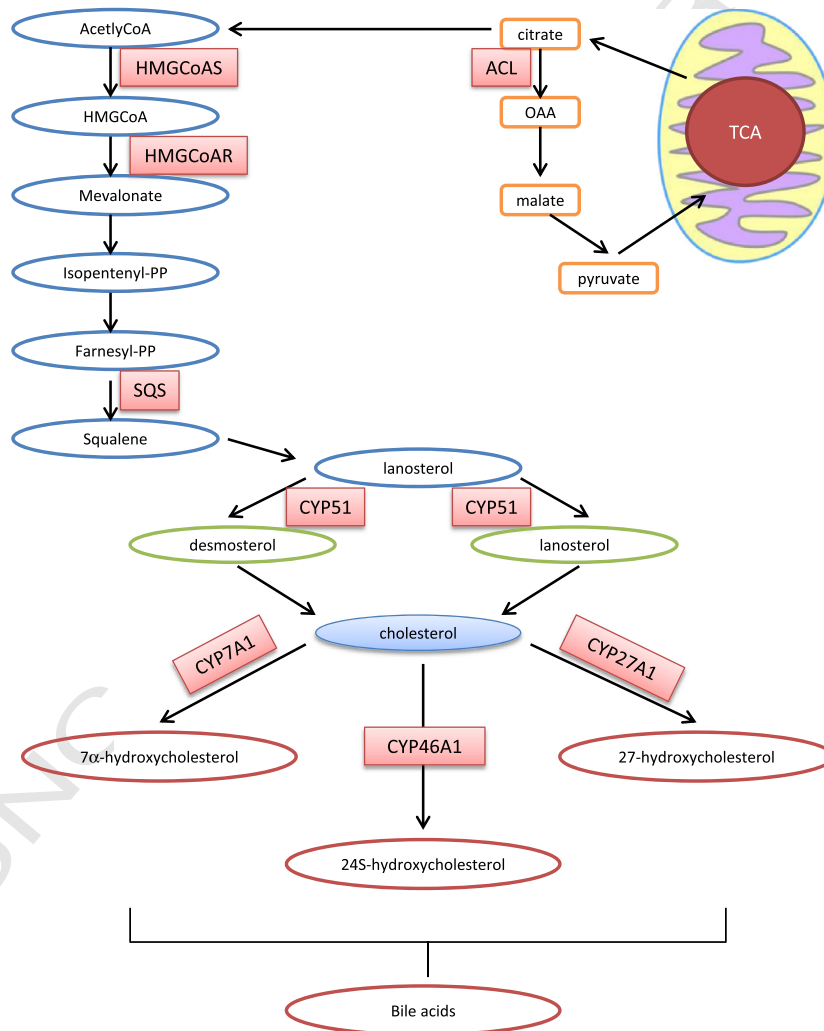
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Cholesterol is synthesized from acetyl-CoA that is converted into 3-hydroxy-3-methylglutaril-CoA (HMGCoA) through two condensation steps (Figs. 1 and 2). Microsomal HMGCoA reductase (HMGCoAR) catalyzes the reduction of HMGCoA into mevalonate in endoplasmic reticulum (ER). Mevalonate is phosphorylated into isopentenyl-pyrophosphate and other isoprenoids and then by condensation of six units is formed squalene which is cyclized into the parental sterol, lanosterol. Two alternative ways proceed with the cholesterol synthesis: the Block pathway (desmosterol as the main intermediate) and the Kandutsch–Russell pathway (lanosterol and 7-dehydrocholesterol, the two main intermediates). The quantification of the cholesterol precursors lanosterol, lanosterol and desmosterol is considered as surrogate marker for tissue or whole body cholesterol synthesis [1–3].

The HMGCoAR reaction is recognized as the rate limiting step of cholesterol synthesis [4]. Cholesterol and oxysterols are directly involved in a negative feedback mechanism of the enzyme regulation both at the protein and the transcriptional level. Oxysterols modulate lipid synthesis by acting on sterol responsive element (SRE) binding proteins (SREBPs). These transcription factors regulate lipid homeostasis in vertebrate cells

by activation of more than 30 genes involved in the synthesis and up- 81  
take of cholesterol, fatty acids, triglycerides and phospholipids as well 82  
as NADPH [5]. SREBPs are expressed as inactive 120 kDa precursors 83  
(pSREBPs) integral to the ER membrane. When intracellular cholesterol 84  
levels are low, pSREBPs are translocated from the ER to the Golgi by an 85  
escort protein, SREBP cleavage-activating protein (SCAP), where they 86  
are cleaved into a 67 kDa active transcription factors, not membrane 87  
bound. These shorter mature SREBPs (mSREBPs) enter the nucleus 88  
and modulate transcription of genes containing a SRE in the promoter 89  
region. When intracellular cholesterol levels are in excess, SCAP, which 90  
has a cholesterol-sensing domain, binds insulin induced gene (Insig) 91  
and the Insig-SCAP-pSREBP is retained in the ER reducing cholesterol 92  
synthesis [6,7]. SREBPs exist in three isoforms: SREBP-1A activates 93  
cholesterol, fatty acid and triglycerides synthesis, SREBP-1C enhances 94  
fatty acid synthesis and SREBP-2 is primarily involved in cholesterol 95  
synthesis [6]. 96

About 1 g of cholesterol is eliminated from the body every day. Ap- 97  
proximately half of this is excreted into the feces after conversion into 98  
bile acids; the remainder is excreted as non-metabolized cholesterol 99



**Fig. 1.** Simplified diagram of cholesterol metabolism in the cells. The formation of acetyl-CoA is the first step of cholesterol and fatty acids synthesis. The acetyl-CoA enters in cytosol in form of citrate by the tricarboxylate transport system. ATP-citrate-lyase (ACL) converts citrate into acetyl-CoA and oxaloacetate in an ATP-driven reaction. HMGCoAS catalyzes the condensation of 3 acetyl-CoA into HMGCoA. The rate limiting step occurs at the HMGCoAR followed by mevalonate formation. Phosphorylation is required to solubilize the isoprenoid intermediates in the pathway (the PP abbreviation stands for pyrophosphate). Intermediates in the pathway are used for the synthesis of prenylated proteins, dolichol, coenzyme Q and the side chain of Heme A. Pyrophosphated isoprenoids are condensed and cyclized by squalene synthetase (SQS) then the first sterol, lanosterol is formed. Two alternative pathways (Block and Kandutsch–Russell) lead to cholesterol formation. Liver CYP7A1 converts cholesterol into 7 $\alpha$ -hydroxycholesterol (7 $\alpha$ OHC), the main precursor of the neutral bile acid pathway. Cholesterol 27-hydroxylase (CYP27A1), expressed in different cell types, converts cholesterol into 27-hydroxycholesterol (27OHC), precursor of the acidic bile acid pathway. Neuronal specific cholesterol 24-hydroxylase (CYP46A1) is responsible for 24S-hydroxycholesterol (24OHC) formation.

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