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Investigation of potential breath biomarkers for the early diagnosis of breast cancer using gas chromatography–mass spectrometry

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ABSTRACT

Background: Breast cancer (BC) remains the most commonly diagnosed malignancy in women. We investigated 420straight aldehydes in the exhaled breath as potential early BC diagnostic biomarkers.21Methods: End-tailed breath were collected by Bio-VOC® sampler and assayed by gas chromatography–mass22spectrometry. Kruskal–Wallis one-way analysis of variance test and binary logistic regression were used for23data analysis. The diagnostic accuracies were evaluated by receiver operating characteristic curves. A predictive24model/equation was generated using the 4 biomarkers and validated by leave-one-out cross-validation.25Results: All four potential biomarkers demonstrated significant differences in concentrations between BC and
healthy controls (HC) (p < 0.05). The areas under the curves (AUCs) in HC vs BC_{I-II} model using hexanal, hepta-
2727nal, octanal, and nonanal were 0.816, 0.809, 0.731, and 0.830, respectively. The AUC for their combined use was
28280.934 (sensitivity 91.7%, specificity 95.8%) in the early diagnosis of BC. The predictive model/equation exhibited
2929good sensitivity (72.7%) and specificity (91.7%) in distinguishing between HC and BC (cross-validation: sensitiv-
3030ity 68.2% and specificity 91.7%).31

Conclusions: The diagnostic values of 4 exhaled straight aldehydes as early diagnostic biomarkers for BC were 32 successfully verified and the diagnostic accuracy improved in their combined use. 33

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39 1. Introduction

Breast cancer (BC) remains the most commonly diagnosed malignan-40 cy and the second leading cause of cancer-related deaths in women [1]. 41 42Early diagnosis is very important for reducing the BC mortality rate. However, the early diagnosis of BC is limited because the disease usually 43develops asymptomatically, while the conventional diagnostic tech-44 niques still have some shortcomings [2]. Therefore, there is still an urgent 4546 clinical need to improve or alternate these methods. The combination of molecular screening methods with these conventional methods may 47 help improve this situation [2]. Cancer cells release molecular bio-48 49 markers into the blood during their growth, which can provide prognostic symbols that could be useful in early cancer diagnosis through breath, 50blood, and urine analyses [3–6]. Breath analysis is noninvasive, easy to 5152perform, painless, and present no risks, thus it may allow the develop-53ment of convenient and safe method that could complement blood and 54urine tests [7].

Human exhaled breath (EB) mainly comprises N₂, CO₂, O₂, water 55 vapor, inert gases, volatile organic compounds (VOCs) and nonvolatile 56 substances [8,9]. These VOCs are either produced endogenously via 57 metabolism or absorbed exogenously from the environment. Some 58 additional compounds are produced and/or their concentrations are 59 changed in pathological states of the body [10]. Endogenous biomarkers 60 are commonly used for diagnostic purposes. 61

There is a considerable experimental evidence that the volatile com- 62 ponents in EB can be used to detect neoplasms [9,11,12]. The theoretical 63 basis for diagnosing BC based on breath analysis is speculative as follows. 64 BC is accompanied by increased oxidative stress (OS), which can lead to 65 DNA, proteins and lipid damages. The induction of cytochrome P450 en- 66 zymes leads to lipid peroxidation of polyunsaturated fatty acids in the 67 cell membranes and resulting in overexpression of volatile alkanes and 68 alkane-derivatives in the breath, and eventually affecting the abundance 69 of VOCs in the EB [13–15]. Besides, free radical-induced oxidation of 70 amino acids and proteins has also been demonstrated to result in the 71 generation of some hydrocarbons along with malonaldehyde [16–18]. 72

Only a limited number of studies have been performed to investigate 73 the feasibility of BC diagnosis based on breath analysis. The published 74 data show that the patterns of VOCs in the EB can distinguish patients 75 with and without BC [19–21]. Unfortunately, the reported potential 76

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breath biomarkers for BC differ among studies [22]. Therefore, there isstill a need to continue searching breath biomarkers for BC.

The growing interest in the discovery of breath biomarkers of the OS 7980 associated with lung cancer, especially straight aldehydes [23-25], led us to consider whether straight aldehydes are potential effective breath 81 biomarkers for BC. Besides, there have been articles reported that form-82 83 aldehyde [26] and heptanal [20] may be potential BC breath biomarkers. Aldehydes are secondary breakdown products of lipids oxidation [27, 84 85 28]. In particular, C5–C10 monofunctional aldehydes (including hexa-86 nal, heptanal, octanal, and nonanal) have been identified to be derived from lipid peroxidation [28]. The well-understood biochemical path-87 ways of these aldehydes simplify the evaluation of their diagnostic 88 values. In addition, volatile aldehydes have low solubility in the blood, 89 90 so they are excreted into the breath within minutes after their formation in tissues [9,29]. Furthermore, hexanal, heptanal, octanal and 91 nonanal were relatively stable than low molecular weight aldehydes 92 and they all could be detected by our method. Moreover, we found 93 94 that their levels seem to be different between HC and BC in our preliminary experiment results. Therefore, among a variety of straight alde-95 hydes, we choose to study hexanal, heptanal, octanal and nonanal. 96

97 Elevated levels of aldehydes imply enhanced OS and they have been 98 proposed as a measure for cancer status diagnosis [30–32]. Moreover, 99 the pattern of aldehydes appeared to be unique to each form of cancer [31]. Hexanal, heptanal, some alkanes, alkane derivatives, and benzene 100 derivatives [33], pentanal, hexanal, octanal, and nonanal [24] were re-101 ported to be potential breath biomarkers for lung cancer. Furthermore, 102formaldehyde [26] and heptanal [20] was also reported as potential 103 104 breath biomarkers for BC. Therefore, we and other researchers have proposed that exhaled aldehydes are potential diagnostic biomarkers 105for various human cancers, including BC [20,22,24,26,33,34]. 106

107 Although Fuchs et al. [24] found out that exhaled aldehydes could be 108used as lung cancer biomarkers, they only focus on lung cancer. Peng 109et al. [35] achieved the discrimination between 'healthy' and 'cancerous' breath in a variety of human cancers (including lung, breast, colorectal, 110 and prostate cancers) by a nanosensor array. To the best of our knowl-111 edge, no previous studies have evaluated in detail the clinical value of 112 identifying BC using specific volatile straight aldehydes in EB by simple 113 direct gas chromatography-mass spectrometry (GC-MS) analysis with-114 out any preconcentration. 115

Therefore, we designed a systematic study to investigate and identify novel potential breath aldehyde biomarkers for the early diagnosis of BC. Based on literature survey [22–24,31,32] and preliminary experiment results, we choose hexanal, heptanal, octanal and nonanal (which were relatively stable than low molecular weight aldehydes, and were often reported as potential tumor biomarkers, as well as their biochemical pathways were well understood) as our target compounds.

123 2. Materials and methods

124 2.1. Chemicals and calibration standards

Hexanal (98%) was obtained from Sigma-Aldrich. Heptanal (97%), octanal (99%), and nonanal (96%) were from Aladdin Chemistry. Absolute ethanol (analytical grade) was purchased from Chengdu Tianhua Chemical Industry. The standard aldehyde solutions were prepared in absolute ethanol.

The standard stock aldehyde solution mixtures (hexanal: 3.98 \times 130 10^{-2} mol/l, 2.39×10^{-2} mol/l, 1.59×10^{-2} mol/l, 7.96×10^{-3} mol/l, 131 1.99×10^{-3} mol/l, 9.95×10^{-4} mol/l, 7.96×10^{-5} mol/l; heptanal: 1323.61 \times 10 $^{-2}$ mol/l, 2.17 \times 10 $^{-2}$ mol/l, 1.44 \times 10 $^{-2}$ mol/l, 7.22 \times 133 10^{-3} mol/l, 1.81×10^{-3} mol/l, 9.02×10^{-4} mol/l, 7.22×10^{-5} mol/l; 134 octanal: 3.17×10^{-2} mol/l, 1.90×10^{-2} mol/l, 1.27×10^{-2} mol/l, 135 6.34×10^{-3} mol/l, 1.59×10^{-3} mol/l, 7.92×10^{-4} mol/l, 6.34×10^{-5} mol/l; nonanal: 2.79×10^{-2} mol/l, 1.67×10^{-2} mol/l, 1.12×10^{-2} mol/l, 1.12×10^{-2 136 137 10^{-2} mol/l, 5.58×10^{-3} mol/l, 1.40×10^{-3} mol/l, 6.98×10^{-4} mol/l, 138

 5.58×10^{-5} mol/l) were prepared by diluting 50 µl of the respective 139 liquid aldehydes with ethanol and making subsequent dilutions. 140

Standard aldehyde gaseous mixtures were produced by injecting 10 141 µl of the stock solution mixtures into 3 L Tedlar bags, which were prefilled with 3 l of high purity nitrogen via septa. Before use, the Tedlar 143 bags were flushed with high purity nitrogen five times and the residual 144 gas was evacuated. Compared to standard Tedlar bag handling 145 procedures [36], we did not heat the Tedlar bag filled with N₂ at 85 °C 146 for 8 h in the bag cleaning process. Because standards and real samples 147 were stored in different Tedlar bags with different sizes in our experi-148 ment, and no residual contamination interference was found in our 149 study or other literatures [37], thus, we simplified the clean procedures 150 of Tedlar bag. The gas volumes in the Tedlar bags were measured using 151 a mass flow controller. We found that 20 min at room temperature 152 (approximately 20 °C) was sufficient for the standards to evaporate 153 completely. 154

Indoor air is an unsuitable sample matrix because the background155levels of aldehydes are relatively high and fluctuating. Nevertheless, it156has been reported that the composition of the matrix did not affect alde-157hyde determinations [37]. Therefore, the calibrations were performed158using high purity nitrogen.159

2.2. Human subjects

Three subject groups were studied: 24 HC, 17 breast benign tumor161(BBT) patients, and 22 BC patients (five at stage I, seven at stage II, 162162nine at stage III, and one at stage IV). All of the subjects were Chinese163females who had no history of smoking.164

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The patients were recruited from the Galactophore Department of 165 China West Hospital of Sichuan University. The patients were selected 166 randomly. Subjects were eligible to participate if they were \geq 18 years 167 old, and if clinical breast examination or imageological examination exhibited breast neoplasm. All patients' EB samples were collected before 169 surgery, radiotherapy or chemotherapy. The patient diagnoses were all 170 confirmed by the postoperative pathological diagnoses. The HC were 171 healthy volunteers with no medical history of malignancy or clinically 172 significant breast disease. Subjects were eligible to participate if they 173 were aged \geq 18 years and had no history of previously diagnosed cancer 174 at any site. Informed consent was obtained from each subject at the time 175 of enrolment. The study was approved by the Ethics Committee of 176 Sichuan University and conducted in accordance with the Declaration 177 of Helsinki. 178

2.3. Breath collection apparatus and sample storage device

The end-tidal breath samples were collected from the human sub- 180 jects using a Bio-VOC® sampler (Markes International Ltd.) which is a 181 preproduction device based on a syringe made from an inert plastic 182 and fitted with a one way valve piston [38]. The subject blows through 183 the sampler syringe and the last portion of breath is retained in the sam- 184 pler, and subsequently the sample is transferred by pushing the piston 185 with a 94% transfer efficiency and no significant loss of analyte or con-186 tamination [38]. The Bio-VOC® sampler collects a representative sam- 187 ple of end-tidal air (approximately 150 ml), without contaminating or 188 diluting with breath from the bronchial tubes or mouth. It can even be 189 used to collect breath samples from a small child and elderly or bedrid- 190 den patients, without causing discomfort. Compared to breath sampling 191 equipment with standard CO2 trigger valves [39], the Bio-VOC® sampler 192 is more simple, miniature, and do not need any electronic power supply, 193 therefore, it is more flexible. Moreover, its usage is simpler and desired 194 end-tailed breath samples can be obtained without any professional 195 operators. 196

The breath sampling for the subjects was done in a separate room 197 after participants had rested for 10 min. The sampling room had good 198 ventilation and no obvious chemical pollution source. The subjects 199 were asked to exhale a single slow breath through a disposable 200

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