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Peptides

journal homepage: www.elsevier.com/locate/peptidesThree new antimicrobial peptides from the scorpion *Pandinus imperator*[☆]Q1 Xian-Chun Zeng^{a,*}, Lingli Zhou^a, Wanxia Shi^a, Xuesong Luo^a, Lei Zhang^a, Yao Nie^a, Jinwei Wang^a,
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ABSTRACT

Three novel cysteine-free venom peptides, which were referred to as Pantinin-1, Pantinin-2 and Pantinin-3, respectively, have been identified from the scorpion *Pandinus imperator* by cDNA cloning strategy. The precursor of each peptide consists of a signal peptide, a mature peptide with no disulfide bridges, and an acidic propeptide with a typical processing signal. Each of the three peptides is an α -helical, cationic and amphipathic molecule with 13 or 14 amino acid residues. Their amino acid sequences are homologous to those of some 13-mer antimicrobial peptides isolated from scorpions. Antimicrobial assay showed that all the three peptides possess relatively strong activities against Gram-positive bacteria and a fungus, but have very weak antimicrobial activities against Gram-negative bacteria. Toxicity assay showed that the three peptides exhibit very low or mild hemolytic activities against human red blood cells. It is interesting to see that Pantinin-3 is able to potently inhibit the growth of vancomycin-resistant *Enterococcus* (VRE) S13, a pathogen that can cause a number of human infections; this suggests that Pantinin-3 has great potential to be applied in the treatment of VRE infections. Our findings gain new insights into the structure/function relationships of the small linear cationic antimicrobial peptides from scorpions, and provide new templates for designing of antimicrobial agents targeting antibiotic-resistant pathogenic bacteria.

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1. Introduction

Antimicrobial peptides (AMPs) are considered to be important components of innate immunity that fight against invading pathogenic organisms in various unfavorable conditions [8,22,24]. They often show a broad-spectrum, non-specific activity against a wide range of microorganisms, including viruses, Gram-positive and Gram-negative bacteria, protozoa and fungi, and may also exert hemolytic effect on erythrocytes [32]. Because AMPs have a mechanism of action that differs from that of currently available therapeutic antibiotics, antibacterial peptides have become an alternative type of antibiotics for treatment of bacterial infectious diseases [10,23,28].

AMPs are commonly characterized as cationic and amphipathic peptides (13–44 amino acids long), which can be classified into three main groups according to their structures: (i) linear peptides containing amphipathic α -helical structure without cysteines; (ii) peptides including one or more disulfide bridges, forming β -sheet or both α -helix and β -sheet; (iii) peptides with irregular amino acids [13,16,19]. These peptides were derived from prokaryotes, amphibians, mammals and arthropods [1,12,17,33].

Until now, at least thirty AMPs have also been identified from scorpion venoms, including Hadrurin from the scorpion *Hadrurus aztecus* [29], Parabutopirin from *Parabuthus schlechteri* [26], BmKn1, BmKn2, BmKb1 and BmKbpb from *Mesobuthus martensii* Karsch [15,35,36], IsCT and IsCT2 from *Opisthacanthus madagascariensis* [6,7], StCT1 and StCT2 from *Scorpiops tibetanus* [3,31], CII-dlp from *Centruroides limpidus limpidus* [27], Pandinin 1, Pandinin 2 from *Pandinus imperator* [4], Opistopirin 1 and Opistopirin 2 from *Opisththalmus carinatus* [18], Vejovine from *Vaejovis mexicanus* [11], VmCT1 and VmCT2 from *Vaejovis mexicanus smithi* [25], VsCT1 and VsCT2 from *Vaejovis subcristatus* [25], Imcropirin from *Isometrus maculatus* [38], Mucropirin from *Lychas mucronatus* [5], Ctriporin from *Chaerilus tricostatus* [9], Hp1090 and Hp1035 from *Heterometrus petersii* [30], HsAp, HsAp2, HsAp3 and HsAp4 from *Heterometrus spinifer* [20], UyCT1, UyCT2 and UyCT5 from

[☆] The cDNA sequences of Pantinin-1, Pantinin-2 and Pantinin-3 reported in this paper have been deposited to the GenBank database under the accession numbers of KC538864, KC538865 and KC538866, respectively.

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58 *Urodacus yaschenko* [14]. Among these scorpion AMPs, Ctriporin,
59 StCT2 and Imcroporin have been shown to have great potentials to
60 treat multidrug-resistant bacterial infections.

61 Here, we identified three new antimicrobial peptides, which
62 were referred to as Pantinin-1, Pantinin-2, Pantinin-3, respectively,
63 from the scorpion *P. imperator* by a cDNA library screening strategy.
64 These peptides contain 13 or 14 amino acid residues, respec-
65 tively. They are cationic and amphipathic consisting of α -helical
66 structures. We found that they possess very weak antimicro-
67 bial activities against Gram-negative bacteria, but have relatively
68 strong activities against Gram-positive bacteria and a fungus.
69 Their antimicrobial activities and toxicity in human red blood
70 cells were shown to be highly associated with the hydrophobic-
71 ity of their hydrophilic regions of the molecules. It is noteworthy
72 that Pantinin-3 has great potential to treat the infections caused
73 by vancomycin-resistant *Enterococcus*. Our studies provide new
74 insights into the structure/function relationship of the small linear
75 antimicrobial peptides from scorpions.

76 2. Materials and methods

77 2.1. Animals

78 Scorpions of *P. imperator* were purchased from the Guangdong
79 province, southern China. The animals were kept in an escape-
80 proof tank, which was provided with some cricket foods and a wet
81 sponge.

82 2.2. Construction of a cDNA library from the venom glands of the 83 scorpion

84 Ten specimens of the scorpion were stimulated using electric-
85 ity. This allows the toxin-producing cells of the venom glands to
86 enter a secretory phase. Twenty four hours later, the telsons of the
87 scorpions were cut off and immediately placed into a mortar con-
88 taining liquid nitrogen. The telson tissues were thoroughly ground
89 into fine powders with a pestle. Total RNA was extracted from
90 the homogenized tissues with the TRIZOL reagent. Poly(A) mRNA
91 was purified using a TaKaRa mRNA purification kit. A cDNA library
92 was constructed using the SuperScriptTM cDNA library construc-
93 tion kit according to the manufacturer's instructions. The purified
94 cDNA was ligated into the pMD[®]-19 T plasmid. To generate a cDNA
95 library, the ligated cDNAs were transformed into the *Escherichia coli*
96 DH5 α competent cells.

97 2.3. Screening of the cDNA library

98 The cDNA library was screened using a "size-selective sequenc-
99 ing strategy" as described previously [34]. The insert sizes of the
100 clones from the cDNA library were determined by PCR using the
101 universal primers M13F and M13R. The cDNA clones that are
102 350–650 base pairs long were sequenced by Nanjing GenScript
103 Biotechnology Corporation.

104 2.4. Bioinformatics analysis

105 Searches for open reading frames (ORFs) of nucleotide
106 sequences were performed using a Translate tool ([http://web.
107 expasy.org/translate/](http://web.expasy.org/translate/)). Protein amino acid sequence homology
108 search was performed using the BLASTP server ([http://blast.ncbi.
109 nlm.nih.gov/Blast.cgi](http://blast.ncbi.nlm.nih.gov/Blast.cgi)). Signal peptide was predicted with the
110 SignalP 4.0 program (<http://www.cbs.dtu.dk/services/SignalP/>).
111 Multiple sequence alignment was performed using the ClustalX
112 program in the BioEdit software. Secondary structure was predicted
113 using the NPS@ sever (www.bioinf.manchester.ac.uk/dbbrowser/

bioactivity/NPS2.html). The helical wheel diagram of an α -
114 helical peptide was generated using the HeliQuest server
115 (<http://heliquest.ipmc.cnrs.fr/cgi-bin/ComputParams.py>). Bio-
116 chemical properties were analyzed with the ProtParam tool
117 (<http://web.expasy.org/protparam/>).
118

119 2.5. Peptide synthesis

120 Peptides were synthesized by GL Biochem Ltd. (Shanghai,
121 China), using the solid phase method on the CS Bio automated mul-
122 tiple peptide synthesizer, and modified by C-terminal amidation.
123 Synthesized peptides were purified using the reverse-phase high
124 performance liquid chromatograph (RP-HPLC), and verified by the
125 mass spectrometry and amino acid composition analysis.

126 2.6. Antimicrobial assay

127 The antimicrobial activity of the synthesized peptides was
128 tested against some representative strains of Gram-positive, Gram-
129 negative bacteria and a fungus. Bacteria were grown in the BactoTM
130 Tryptic Soy Broth (TSB) medium at 37 °C or 30 °C until the OD₆₀₀
131 value of the culture reached around 0.4. The bacterial culture was
132 then diluted with the fresh TSB medium into a final OD₆₀₀ of 0.002.
133 Twenty microliters of each peptide solution prepared by serial dilu-
134 tion was added to a 96-well plate containing 180 microliters of the
135 diluted bacterial culture. The plate was incubated at 37 °C or 30 °C
136 with vigorous shaking. Bacterial proliferation was determined by
137 measuring the absorbance at 620 nm, and the minimal inhibitory
138 concentration (MIC) of the peptide, expressed as the lowest con-
139 centration that causes 100% inhibition of bacterial growth, was thus
140 achieved.

141 2.7. Hemolytic assay for the human red blood cells

142 Hemolytic activity of the peptides was determined by measur-
143 ing the release of hemoglobin from the human red blood cells as
144 described previously [35].

145 3. Results

146 3.1. Precursors of three novel cysteine-free peptides from the 147 scorpion *P. imperator*

148 Three novel full-length cDNAs, which are 371, 392 and 359 base
149 pairs long, respectively, were obtained from the cDNA library made
150 from the venom glands of the scorpion *P. imperator*. The cDNAs
151 code for the precursors of three novel cysteine-free peptides that
152 were referred to as Pantinin-1, Pantinin-2, and Pantinin-3, respec-
153 tively. Each of the three transcripts consists of a 5'-untranslated
154 region (5'-UTR), an open reading frame (ORF), and a 3'-untranslated
155 region (3'-UTR) with single or double putative polyadenylation
156 signal (AATAAA or AACAAA) and a typical eukaryotic polyA tail
157 (Fig. 1A–C). The ORFs of the transcripts of Pantinin-1, Pantinin-2
158 and Pantinin-3 are 210, 207 and 207 nucleotides long, respectively,
159 and display high homology (61–81%) with each other. However, the
160 nucleotide sequences of their 5'-UTRs, which are 113, 65 and 44
161 nucleotides long, respectively, show little similarity to each other.
162 The 3'-UTR sequences of the three transcripts, which are 48, 120
163 and 108 nucleotides long, respectively, also show little homology
164 with each other. It has been established that 5'-UTR and 3'-UTR play
165 key roles in the control of amount and timing of gene expression.
166 Therefore, the high level of sequence divergence in the 5'-UTRs and
167 3'-UTRs of the three transcripts would lead to highly different abun-
168 dances of the three peptides in the scorpion venom. Our findings
169 also suggest that the genes of the three peptides were originated
170 from a common ancestor by gene duplication, and subsequently

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