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ACCEPTED MANUSCRIPT

Control of *Aspergillus flavus* growth and aflatoxin production in transgenic maize kernels expressing a tachyplesin-derived synthetic peptide, AGM182

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Highlights:

- Designed the synthetic peptide AGM182 modeled after the naturally occurring tachyplesin 1.
- AGM182 was five-times more effective in controlling Aspergillus flavus compared to tachyplesin 1.
- Transgenic maize plants expressing the synthetic peptide AGM182 were produced and advanced to third generation by selfing.
- Kernel Screening Assay showed significant reduction in fungal growth (72%) and spread inside transgenic kernels.
- Concomitant, significant reduction in aflatoxin levels (76-98%) was also achieved in transgenic kernels.

Abstract

Aspergillus flavus is an opportunistic, saprophytic fungus that infects maize and other fatty acidrich food and feed crops and produces toxic and carcinogenic secondary metabolites known as aflatoxins. Contamination of maize with aflatoxin poses a serious threat to human health in addition to reducing the crop value leading to a substantial economic loss. Here we report designing a tachyplesin1-derived synthetic peptide AGM182 and testing its antifungal activity both *in vitro* and *in planta*. *In vitro* studies showed a five-fold increase in antifungal activity of AGM182 (vs. tachyplesin1) against *A. flavus*. Transgenic maize plants expressing AGM182 under maize *Ubiquitin*-1 promoter were produced through *Agrobacterium*-mediated transformation. PCR products confirmed integration of the AGM182 gene, while RT-PCR of maize RNA confirmed the presence of AGM182 transcripts. Maize kernel screening assay using a highly aflatoxigenic *A. flavus* strain (AF70) showed up to 72% reduction in fungal growth in the transgenic AGM182 seeds compared to isogenic negative control seeds. Reduced fungal growth in the AGM182 transgenic seeds resulted in a significant reduction in aflatoxin levels (76-98%). The results presented here show the power of computational and synthetic biology to rationally design and synthesize an antimicrobial peptide against *A. flavus* that is effective in

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