



# Modelling water use efficiency in a dynamic environment: An example using *Arabidopsis thaliana*<sup>☆</sup>

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## ABSTRACT

Intrinsic water use efficiency ( $W_i$ ), the ratio of net  $\text{CO}_2$  assimilation ( $A$ ) over stomatal conductance to water vapour ( $g_s$ ), is a complex trait used to assess plant performance. Improving  $W_i$  could lead in theory to higher productivity or reduced water usage by the plant, but the physiological traits for improvement and their combined effects on  $W_i$  have not been clearly identified. Under fluctuating light intensity, the temporal response of  $g_s$  is an order of magnitude slower than  $A$ , which results in rapid variations in  $W_i$ . Compared to traditional approaches, our new model scales stoma behaviour at the leaf level to predict  $g_s$  and  $A$  during a diurnal period, reproducing natural fluctuations of light intensity, in order to dissect  $W_i$  into traits of interest. The results confirmed the importance of stomatal density and photosynthetic capacity on  $W_i$  but also revealed the importance of incomplete stomatal closure under dark conditions as well as stomatal sensitivity to light intensity. The observed continuous decrease of  $A$  and  $g_s$  over the diurnal period was successfully described by negative feedback of the accumulation of photosynthetic products. Investigation into the impact of leaf anatomy on temporal responses of  $A$ ,  $g_s$  and  $W_i$  revealed that a high density of stomata produces the most rapid response of  $g_s$  but may result in lower  $W_i$ .

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## 1. Introduction

In order to meet the projected demand for cereal production by 2050, crop yields must improve by 1.16–1.31% each year; however, current estimates are well below this required rate [1]. The primary determinant of crop yield is the cumulative rate of photosynthesis over the growing season and is determined by the ability of the plant to capture light and  $\text{CO}_2$ , use this energy to convert the  $\text{CO}_2$  to biomass, and how much of this biomass ends in usable yield. Improving photosynthetic efficiency is recognised as an important but unexploited avenue to increase yield potential in crop plants [2]. Increasing photosynthetic efficiency is accompanied by a higher  $\text{CO}_2$  demand, which can be limited by the resistance of  $\text{CO}_2$  diffusion into the leaf. Any attempt to decrease this resistance greatly increases the water loss by transpiration from the leaf.

Photosynthetic productivity is linked to water consumed by the plant and often measured as water use efficiency ( $WUE$ ).  $WUE$  can be defined at different scales of time and space and, at the leaf level, it is often assessed as the ratio of  $\text{CO}_2$  fixed per unit of  $\text{H}_2\text{O}$  transpired ( $E$ ). Intrinsic water use efficiency ( $W_i$ ) is defined when stomatal conductance to water vapour ( $g_s$ ) is used instead of  $E$ . The use of  $g_s$  to describe the stomatal control on the rate of  $E$  facilitates the comparisons between different leaves and environmental conditions. The photosynthetic capacity of the leaf determines the net  $\text{CO}_2$  assimilation ( $A$ ) as a function of the variation in the microclimate surrounding the leaf. Over the diurnal period,  $A$  is mainly determined by the irradiance absorbed by the leaf and the limitation of  $\text{CO}_2$  imposed by stomatal control. Under field conditions, environmental variables that affect both photosynthesis and stomatal behaviour are rarely constant. For example, light intensity (and spectral quality) alters in time scales of seconds to hours to which  $A$  and  $g_s$  must respond. The temporal response of  $A$  and  $g_s$  to a fluctuating environment are asynchronous, with  $g_s$  response often an order of magnitude slower than  $A$ , which results in rapid variations of  $W_i$ . Thus, it is important when describing the kinetic response of  $W_i$  to use an approach that considers responses by  $A$  and  $g_s$  simultaneously.

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## Nomenclature

|            |                                                                         |
|------------|-------------------------------------------------------------------------|
| $A$        | Net CO <sub>2</sub> assimilation                                        |
| $A_G$      | Gross CO <sub>2</sub> assimilation                                      |
| $R_d$      | Mitochondrial respiration                                               |
| $g_s$      | Stomatal conductance to water vapour                                    |
| $g_m$      | Mesophyll conductance to CO <sub>2</sub>                                |
| $g_b$      | Boundary layer conductance to water vapour                              |
| $g_t$      | Total conductance to CO <sub>2</sub>                                    |
| $W_i$      | Intrinsic water use efficiency                                          |
| $C_a$      | Atmospheric CO <sub>2</sub> concentration                               |
| $C_i$      | CO <sub>2</sub> concentration in the intercellular airspaces            |
| $C_c$      | CO <sub>2</sub> concentration at the site of carboxylation              |
| $a$        | Stomatal pore area                                                      |
| $a_s$      | Steady state target of stomatal pore area                               |
| $a_{min}$  | Minimum stomatal pore area                                              |
| $a_{max}$  | Maximum stomatal pore area                                              |
| $\alpha_L$ | Slope of the relationship                                               |
| $\theta_L$ | Curvature factor of the curve                                           |
| $k_i$      | Time constant for an increase in $a$                                    |
| $k_d$      | Time constant for a decrease in $a$                                     |
| $L$        | Percentage of efficiency                                                |
| $SD$       | Stomatal density                                                        |
| $D$        | Diffusivity of water in air                                             |
| $V$        | Molar volume of air                                                     |
| $l$        | Depth of stomatal pore                                                  |
| $P_a$      | The atmospheric pressure                                                |
| $S_a$      | Factor representing the influence of the rate of accumulation of sugars |
| $S_e$      | Factor representing the influence of the rate of export of sugars       |
| $VC_{max}$ | Maximum Rubisco activity                                                |
| $J_{max}$  | Maximum electron transport activity                                     |
| $\alpha$   | Proportion of light absorbed by PSII                                    |

Intrinsic water use efficiency ( $W_i$ ) is dependent on the anatomy (e.g. stomatal size and density) and the physiology (e.g. behaviour) of stomata as well as the leaf biochemistry (e.g. activity of the Calvin cycle), all of which interact to determine the kinetics of CO<sub>2</sub> and H<sub>2</sub>O gaseous exchange between the leaf and atmosphere. The dynamic nature of the interactions between the different components that determine  $W_i$  are not fully understood and need to be addressed if we are to successfully improve both  $A$  and  $W_i$  under dynamic field conditions.

It is possible to conceptualise the inherent complexity of gas exchange over a fluctuating light regime through modelling, which will improve our understanding of the  $W_i$  response by simulating a number of gas exchange scenarios (e.g. changes in light intensity and humidity) that would normally be difficult to assess in a reasonable amount of time using experimental approaches. Current models focus on predicting  $g_s$  in steady state [3] and cannot be used to infer the impact of stomatal behaviour on  $A$  or  $W_i$  under dynamic conditions. Although temporal responses of  $g_s$  have previously been described using a dynamic model [4,5], the relationship between stomatal response and leaf level gas exchange was not clearly described. We propose to use a model that will take into consideration the anatomy and physiology of stomata to more accurately represent the stomatal control of  $W_i$ .

To scale stomatal responses to leaf level  $g_s$ , the two most important stomatal characteristics are aperture and density [6–8]. A high stomatal density does not necessarily result in a higher  $g_s$  as stomata ultimately control their aperture depending on the guard cell responses to the external (e.g. light intensity) and internal (e.g. mesophyll demand for CO<sub>2</sub>) stimuli [9]. To link stomatal behaviour

to leaf level gas exchange responses, we propose a ‘big stoma’ approach that consists of simulating the response of one stoma that is representative of the heterogeneous response of many stomata and scaling the response to the leaf level. This approach was incorporated in an enhanced version of the multi compartments model described by Noe and Giersch [10] to predict  $A$  and  $W_i$ . Scaling up the dynamic of the stomatal response to the leaf level, with the improved model for CO<sub>2</sub> diffusion inside the leaf, will help to dissect  $W_i$  into traits of interest and predict potential gains in  $W_i$ .

The objective of this study was to develop a new model combining our most recent knowledge of kinetics in stomatal behaviour and photosynthesis to describe the temporal response of  $W_i$  over the course of a day with natural dynamic variations in irradiance. All the parameters of the model described here incorporate a trait of interest for  $W_i$  and were adjusted using Bayesian inference. The model was validated using a dataset with a different irradiance pattern to assess the predictive power of the model. A sensitivity analysis was finally performed to show the interaction among the parameters and display the potential gain in  $W_i$  in the case of one or two parameters changing. We used the output of the model to understand how temporal responses in  $g_s$  impacts  $A$  and  $W_i$ .

## 2. Material and methods

### 2.1. Dynamic modelling of photosynthesis and stomatal conductance

The model essentially consists of four differential equations describing the diffusion of CO<sub>2</sub> between different compartments represented by the atmosphere, the intercellular air spaces and the photosynthetic tissues (Fig. 1). The exchanges between these compartments are dependent on the stomatal aperture and the resistance of diffusion in the mesophyll cells. In addition, the model took into account the limitation of photosynthesis and stomatal aperture that appeared during a period of light.

$$\begin{cases} \frac{da}{dt} = \frac{a_s - a}{k_i} \text{ if } a < a_s \\ \frac{da}{dt} = \frac{a_s - a}{k_d} \text{ if } a \geq a_s \end{cases} \quad (1)$$

The first differential equation (Eq. (1)) described the temporal variations of the stomatal pore area ( $a$ ) with  $a_s$  the steady state target followed by  $a$  and two time constants,  $k_i$  and  $k_d$ , for an increase or a decrease of  $a$  respectively. Considering the spatial heterogeneity of the stomatal response, a top-down approach was used, signifying that the model simulated the response of one stoma representative of the sum of the individual stomatal responses and scaled it to leaf level instead of trying to integrate the response of each stoma.

The steady state target of  $a$  ( $a_s$ ) as a function of the light intensity (PPFD) was predicted using a non-rectangular hyperbola [4]:

$$a_s = [a_{min} + \frac{\alpha_L PPFD + (a_{max} - a_{min}) - \sqrt{\alpha_L PPFD + (a_{max} - a_{min})^2 - 4\theta_L \alpha_L PPFD (a_{max} - a_{min})}}{2\theta_L}] \cdot L \quad (2)$$

with  $a_{min}$  and  $a_{max}$  the minimum and maximum stomatal pore area,  $\alpha_L$  the slope of the relationship,  $\theta_L$  the curvature factor of the curve and  $L$  the percentage of efficiency (see below).

$$\frac{dC_i}{dt} = [g_t (C_a - C_i) - g_m (C_i - C_c)] \frac{RT_l}{d_a P_a} \quad (3)$$

Eq. (3) described the variation of the CO<sub>2</sub> concentration in the intercellular airspaces ( $C_i$ ) with  $C_a$  the atmospheric CO<sub>2</sub> concentration and  $C_c$  the CO<sub>2</sub> concentration at the sites of carboxylation.

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