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2 Review

Influence of membrane lipid composition on flavonoid-membrane 64 interactions: Implications on their biological activity

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ABSTRACT

The membrane interactions and localization of flavonoids play a vital role in altering membrane-mediated cell signaling cascades as well as influence the pharmacological activities such as anti-tumour, anti-microbial and anti-oxidant properties of flavonoids. Various techniques have been used to investigate the membrane interaction of flavonoids. These include partition coefficient, fluorescence anisotropy, differential scanning calorimetry, NMR spectroscopy, electrophysiological methods and molecular dynamics simulations. Each technique will provide specific information about either alteration of membrane fluidity or localization of flavonoids within the lipid bilayer. Apart from the diverse techniques employed, the concentrations of flavonoids and lipid membrane composition employed in various studies reported in literature also are different and together these variables contribute to diverse findings that sometimes contradict each other. This review highlights different techniques employed to investigate the membrane interaction of flavonoids with special emphasis on erythrocyte model membrane systems and their significance in understanding the nature and extent of flavonoid-membrane interactions. We also attempt to correlate the membrane localization and alteration in membrane fluidity with the biological activities of flavonoids such as anti-oxidant, anti-cancer and anti-microbial properties.

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64 1. Introduction

65 Flavonoids are a group of polyphenols ubiquitously distributed 66 in vegetables, fruits, seeds, roots and stem either in the aglycone 67 form or as glycone derivatives [1]. All flavonoids possess a typical structure comprising three cyclic carbon rings denoted as A, B 68 69 and C. The substitutions in the 'C' ring are used to distinguish 70 different flavonoid subgroups [1,2]. Flavonoids possess numerous 71 pharmacological activities such as anti-cancer, anti-diabetic, 72 anti-atherosclerotic, anti-oxidant, anti-inflammatory properties, 73 etc [1,2]. They are also consumed as food supplements. The phar-74 macological activities of flavonoids have been primarily attributed 75 to their ability to alter the membrane-mediated signaling path-76 ways resulting in modification of cell membrane permeability 77 and also to their binding with specific protein targets [3]. Fig. 1 78 shows the structure of flavonoids and their major subgroups.

79 It has now been recognized that plant-derived drugs have differ-80 ent molecular targets in the cells that include the cell membrane, 81 intracellular compartments and DNA [4-6]. The gateway for any 82 molecule to enter into the cell is the plasma membrane and hence 83 membrane interactions of a molecule will have a major influence on 84 their mechanisms of action [3,7–12]. Consequently, the biological 85 activities of a molecule will be a manifestation of its mode of inter-86 action with the membrane components as well as its localization 87 and residence time in the cell membrane. As the lipid bilaver 88 construct is a universal component of all membranes, interactions 89 of molecules with the membrane lipid components can be termed 90 as 'non-specific' interactions. Such interactions can also alter the 91 functions of the membrane-associated proteins [13]. The 92 membrane permeation of a molecule is chiefly associated with 93 the nature and extent of its interaction with cellular membranes 94 [8,14]. In the case of flavonoids, the mode of membrane interaction 95 has been mainly attributed to their lipophilicity and planar struc-96 ture [15]. The number and position of hydroxyl groups influences 97 the relative hydrophobicity of different flavonoids [3,16,17]. The 98 presence of a 2,3 double bond in the B ring and absence of glycosidic 99 substituent in the flavonoid structure tends to render it more planar 100 [1]. Thus, it is evident that a strong correlation exists between the 101 structure and biological activity of a flavonoid.

Another major parameter that can influence the biological activity of flavonoids is the fluidity of the membrane. Plasma membrane offers a diffusion barrier that protects the cell cytoplasmic contents from adverse interactions of molecules. However, it transforms into a highly permeable structure when subjected to oxidative stress leading to cell dysfunction and death [13,18]. Oxidative

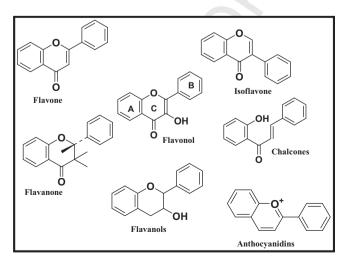


Fig. 1. Structure of flavonoids and their major subgroups predominantly present in the plant kingdom.

stress-induced membrane damage due to lipid peroxidation has 108 been implicated in various free radical-mediated diseases such as 109 neuronal degeneration, atherosclerosis, cancer, rheumatoid arthri-110 tis, etc., all of which are associated with altered membrane fluidity 111 [19]. Apart from free radicals, different exogenous compounds are 112 also known to cause similar changes in the packing density of the 113 membrane lipids on interaction with the plasma membrane [20]. 114 Flavonoids have been reported to localize either in the hydropho-115 bic core of the lipid bilayer or at the membrane interface leading 116 to corresponding alterations in the membrane fluidity or rigidity 117 [3,7,8]. The alteration of membrane fluidity by flavonoids can 118 significantly influence the membrane-mediated cell signaling 119 pathways. Thus, the structure of the flavonoids as well as their 120 ability to alter the membrane fluidity are both important factors 121 that influence the nature and magnitude of their biological activity. 122 This review attempts to elaborate the mode of membrane interac-123 tion of flavonoids and their biological importance with respect to 124 lipid composition and types of membranes. This will pave a way 125 for understanding the pharmacological importance of membrane-126 mediated signaling cascades for developing novel therapeutic 127 agents. 128

A scan of literature reveals that though the importance of flavonoid-membrane interactions has been duly recognized, widely contrasting reports exist on their mode of interactions. The reasons for such conflicting reports may arise due to a combination of many factors. One of the reasons for the observed variations in the results could be the widely varying concentration of the flavonoids employed in these studies [8]. The concentration of the flavonoid could determine the magnitude of molecular stress at the membrane surface and in the interior of the bilayer that would be reflected in the observed effects of the interaction. Yet another aspect that could contribute to the conflicting reports on the effects of flavonoid-membrane interactions is the type of membrane model used for the study. The most common membrane models employed for such studies are planar lipid bilayers and liposome vesicles [21–25]. The lipid composition employed for forming the membrane models will have a pronounced effect on the nature as well as extent of flavonoid-membrane interactions apart from influencing their localization within the lipid bilayer. The most common lipids that have been employed for forming membrane mimics are dipalmitoyl phosphatidyl choline (DPPC), distearoyl phosphatidyl choline (DSPC), palmitoyl oleoyl phosphatidyl choline (POPC) and cholesterol. Fig. 2 depicts the lipid membranes formed using these lipids.

A distinct feature in these membrane bilayers is the difference in their lipid-packing factor. Membranes formed from DPPC or DSPC are more compactly organized with fewer packing defects due to the strong hydrophobic associative forces that exist between the saturated fatty acyl chains of DPPC or DSPC. In contrast, membranes formed using phospholipids containing unsaturated acyl chains such as POPC possess larger number of packing defects due to the *cis* double bond in the unsaturated acyl chain that prevents close association of the neighboring acyl chains [26]. Such membranes are more fluid than those formed from DPPC or DSPC. Introduction of cholesterol in the membrane tends to rigidify it because of the presence of stiffer phenanthrene rings [26]. However, increasing the cholesterol content beyond a certain limit has been found to impart greater fluidity to the membrane bilayer. This has been attributed to the mismatch in the chain length of the phospholipid acyl chains and the cholesterol structure [27]. As a result, more voids are created in the bilayer, thereby transforming it into a highly permeable structure.

The cell membrane consists of different types of lipids along with proteins. The distribution and composition of the lipids depends upon the cell type. Among the different membrane lipids, phosphatidylethanolamine (PE), phosphatidylcholine (PC) and

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