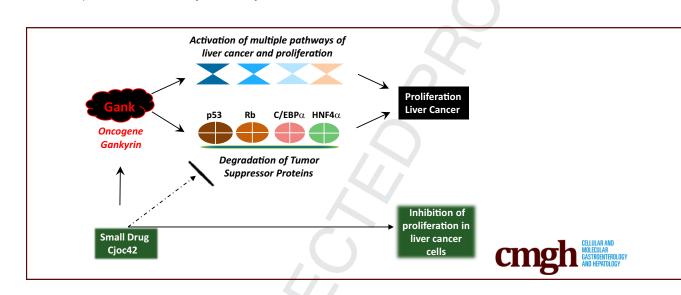
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cmgh ORIGINAL RESEARCH

Gankyrin Promotes Tumor Suppressor Protein Degradation to Drive Hepatocyte Proliferation

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SUMMARY

The mechanisms by which gankyrin promotes hepatic proliferation are not known. This study shows that gankyrin promotes proteosomal degradation of tumor-suppressor proteins. Gankyrin deletion restored tumor-suppressor protein expression and delayed regenerative hepatocyte proliferation in vivo. Furthermore, proteasome inhibition limited growth of human- and mouse-derived liver cancer cell lines in vitro.

BACKGROUND & AIMS: Uncontrolled liver proliferation is a key characteristic of liver cancer; however, the mechanisms by which this occurs are not well understood. Elucidation of these mechanisms is necessary for the development of better therapy. The oncogene Gankyrin (Gank) is overexpressed in both hepatocellular carcinoma and hepatoblastoma. The aim of this work was to determine the role of Gank in liver proliferation and elucidate the mechanism by which Gank promotes liver proliferation.

METHODS: We generated Gank liver-specific knock-out (GLKO) mice and examined liver biology and proliferation after surgical resection and liver injury.

RESULTS: Global profiling of gene expression in GLKO mice
showed significant changes in pathways involved in liver cancer and proliferation. Investigations of liver proliferation after

partial hepatectomy and CCl4 treatment showed that GLKO mice have dramatically inhibited proliferation of hepatocytes at early stages after surgery and injury. In control LoxP mice, liver proliferation was characterized by Gank-mediated reduction of tumor-suppressor proteins (TSPs). The failure of GLKO hepa-tocytes to proliferate is associated with a lack of down-regulation of these proteins. Surprisingly, we found that hepatic progenitor cells of GLKO mice start proliferation at later stages and restore the original size of the liver at 14 days after partial hepatectomy. To examine the proliferative activities of Gank in cancer cells, we used a small molecule, cjoc42, to inhibit interactions of Gank with the 26S proteasome. These studies showed that Gank triggers degradation of TSPs and that cjoc42-mediated inhibition of Gank increases levels of TSPs and inhibits proliferation of cancer cells.

CONCLUSIONS: These studies show that Gank promotes hepatocyte proliferation by elimination of TSPs. This work provides background for the development of Gank-mediated therapy for the treatment of liver cancer. RNA sequencing data can be accessed in the NCBI Gene Expression Omnibus: GSE104395. (*Cell Mol Gastroenterol Hepatol 2018*; =: =-=; https://doi.org/10.1016/j.jcmgh.2018.05.007)

Keywords: Liver; Proliferation; Cancer; Tumor Suppressor 116 Proteins; Progenitor Cells.

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117 Q10 • he development of liver cancer is associated with multiple alterations in cellular function and gene 118 Q11 119 ^{Q12} expression.¹ One of the main hallmarks of liver cancer is 120 uncontrolled proliferation, which is owing in part to damage 121 of pathways essential to cell-cycle control. In addition, 122 regulation in the coordinated expression of oncogenes and 123 Q13 tumor-suppressor proteins (TSPs) is vital to tumor prolif-124 eration. One of the key oncogenes and promoters of liver 125 proliferation is a small subunit of 26S proteasome, Gankyrin 126 (Gank). This non-adenosine triphosphatase subunit of the 127 ubiquitin proteasome system (UPS) is a notorious oncogene 128 expressed in several cancer types, including hepatocellular 129 carcinoma (HCC), in which it was first discovered.²⁻⁴ In 130 agreement with these observations, Gank has been identi-131 fied as the driver oncogene in the early development of liver 132 cancer through chemical models as well as age-dependent 133 hepatic tumorigenesis.^{2–7}

134 Gank promotes the development of HCC through several 135 mechanisms, including the neutralization of TSPs. TSPs are 136 the main proteins that support the quiescent status of the 137 liver, and it has been shown that the activities of more than 138 20 different TSPs are lost in HCC because of mutations or hypermethylation of their promoters.⁸ In addition, the 139 140 elimination of TSPs by Gank is essential to carcinogenesis.⁴ 141 Specifically, Gank leads to the neutralization of essential 142 4 TSPs such as p53, through stabilization of MDM2 ligase and 143 Q15 subsequent enhanced ubiquitination, and Rb, by direct interaction, both of which trigger UPS-mediated degrada-144 tion.^{2,3,9} Studies of liver cancer have identified 2 additional 145 146 targets of Gank: CCAAT/enhancer binding protein α (C/EBP α) and hepatocyte nuclear factor 4α (HNF 4α).^{6,7,9} 147 148 C/EBP α belongs to the C/EBP family of proteins, bZIP 149 proteins, which contain basic region and leucine zipper re-150 gions.¹⁰ C/EBP α has been shown to be a strong inhibitor of proliferation and a strong TSP.^{4,6,10,11} In fact, several recent 151 152 reports with activation of the C/EBP α gene in animal 153 models of liver carcinogenesis showed that its activation 154 leads to inhibition of liver proliferation and carcinogenesis as well as normalization of liver function.^{12–14} HNF4 α is also 155 a strong TSP and expression of this protein correlates with 156 157 the epithelial-mesenchymal transition involved in metastatic tumor formation.¹⁵ It also has been shown that deletion 158 159 of HNF4 α promotes hepatocyte proliferation and diethylnitrosamine (DEN)-induced liver cancer.¹⁶ In addition to these 160 known TSPs, recent studies have identified RNA CUG triplet 161 162 repeat binding protein 1 (CUGBP1) as a tumor suppressor, 163 whose activity depends on phosphorylation/dephosphorylation at serine 302.¹⁷ Generation of CUGBP1-S302A KI mice 164 165 showed that this TSP protects the liver from the develop-166 ment of cancer and that during liver carcinogenesis, Gank 167 eliminates this isoform of CUGBP1.¹⁷ In agreement with this, 168 livers of CUGBP1 knock-out mice show a molecular signa-169 ture of hepatoblastoma and express increased levels of stem 170 cell markers and reduced levels of markers of hepatocytes.⁹ Increasing evidence has shown how Gank is responsible

171 172 for the activation of additional pathways critical to liver 173 cancer. For example, in addition to its effect on TSPs, Gank 174 also stabilizes the stem cell marker Oct4 through elimina-175 Q16 tion of WWP2, the ubiquitin ligase that normally marks

Oct4 for degradation.¹⁸ To promote uncontrolled prolifera-176 tion, Gank also binds to D-type kinase, cdk4, and replaces 177 p16^{INK4a} from cdk4, leading to the activation of cdk4 and 178 cell-cycle progression.² In addition, Gank increases levels of 179 oncogene Nrf2 by the elimination of Keap1 ligase, which 180 triggers degradation of Nrf2.¹⁹ 181

Regulation of activities of Gank in the liver is quite 182 complicated. In the quiescent liver, farnesoid X receptor 183 (FXR) partially represses Gank, however, with DEN-184 185 mediated carcinogenesis, there is a reduction of FXR, activation of Gank, and subsequent activation of the cascade of 186 Gank-dependent pathways including loss of TSPs.⁷ Our 187 recent article showed that activation of FXR by GW4064 188 inhibits the development of liver cancer and that the 189 FXR-Gank axis is involved in the development of pediatric 190 liver cancer.9 In agreement with these findings, a recent 191 report showed that DEN-mediated liver cancer is reduced 192 significantly in mice with liver-specific deletion of Gank.²⁰ 193

In this study, we examined the proliferative activities of 194 Gank in recently generated liver-specific Gank liver-specific 195 knock-out (LKO) mice. By using 2 models of liver prolifer-196 ation/regeneration, partial hepatectomy (PH), and CCl₄ 197 treatments, we obtained evidence showing that Gank pro-198 motes liver proliferation via direct interaction and elimi-199 nation of at least 5 TSPs. We also found that inhibition of 200 Gank by a small drug, cjoc42, inhibits proliferation of liver 201 cancer by blocking the Gank-TSPs axis, suggesting that 202 203 cjoc42 might be considered a novel therapy approach.

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Methods

Animals

207 Experiments with animals were approved by the Insti-208 tutional Animal Care and Use Committee at Cincinnati 209 Children's Hospital (protocol IACUC2014-0042). A Gank 210 LKO (GLKO) mouse model was created using the Cre-Lox 211 system. Mice expressing the Cre recombinase protein 212 driven by the albumin promoter were crossed with mice 213 that had LoxP sequences flanking exons 2-4 of the Gank 214 gene. The resulting offspring had the Gank gene excised only $\frac{q_{17}}{215}$ in cells expressing albumin. 216

Histoloav

219 Liver tissue was taken from the left lobe and fixed in 4% formaldehyde. Mice were injected intraperitoneally with $\frac{018220}{1000}$ 221

222 Abbreviations used in this paper: BrdU, bromodeoxyuridine; cDNA, complementary DNA; C/EBP, CCAAT/enhancer binding protein; Co-IP, 223 co-immunoprecipitation; CUGBP1, CUG triplet repeat binding protein 224 1; DEN, diethylnitrosamine; FXR, farnesoid X receptor; Gank, Gankyrin; 225 HCC, hepatocellular carcinoma; GLKO, Gankyrin liver-specific knock-226 out; HNF4α, hepatocyte nuclear factor 4α; LKO, liver-specific knockout; mRNA, messenger RNA; Opn, osteopontin; PCNA, proliferating 227 cell nuclear antigen; PH, partial hepatectomy; Rb, ; RT-228 PCR, reverse-transcriptase polymerase chain reaction; TSP, tumor-229 suppressor protein; 2D, 2-dimensional; UPS, ubiquitin proteasome system; WT, wild-type. 230 © 2018 The Authors. Published by Elsevier Inc. on behalf of the AGA 231

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