

RESEARCH LETTER

Mutant KRAS Exosomes Alter the Metabolic State of Recipient Colonic Epithelial Cells



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In colorectal cancer (CRC) cells, mutant Kirsten rat sarcoma (KRAS) cell-autonomously imparts Warburg-like¹ metabolic changes through induction of Glucose transporter 1 (GLUT-1) (SLC2A1).^{2,3} We previously reported that mutant KRAS has marked effects on the constituents of CRC exosomes,

including proteins and enzymes involved in metabolism and glycolysis.^{4,5} The present studies were designed to test whether mutant KRAS exosomes can alter the metabolic state cell-nonautonomously in recipient colonic epithelial cells.

We isolated exosomes purified from Daniel L. Dexter derived 1 (DLD-1) cells, which contain 1 wild-type (WT) and 1 mutant KRAS allele, and those from DLD-1 isogenic cell variants genetically engineered to express only the WT KRAS allele (DKs-8) or only the mutant KRAS allele (DKO-1).⁶ Adding DKO-1 exosomes to DKs-8 cells significantly reduced glucose concentrations in the medium, suggesting increased

cellular glucose uptake in these WT KRAS cells (Figure 1A). After 48-hour exposure to DKO-1 exosomes, a significantly higher percentage of recipient DKs-8 cells were in S and G2/M phases of the cell cycle (Supplementary Figure 1A and B), and cell number was increased at 120 hours (Supplementary Figure 1C). We next used the endogenous fluorescent characteristics of Nicotinamide adenine dinucleotide reduced (NADH) and Flavin adenine dinucleotide (FAD) to determine the relative balance between glycolysis and oxidative phosphorylation, as previously reported.⁷⁻⁹ Addition of mutant KRAS exosomes selectively and significantly increased

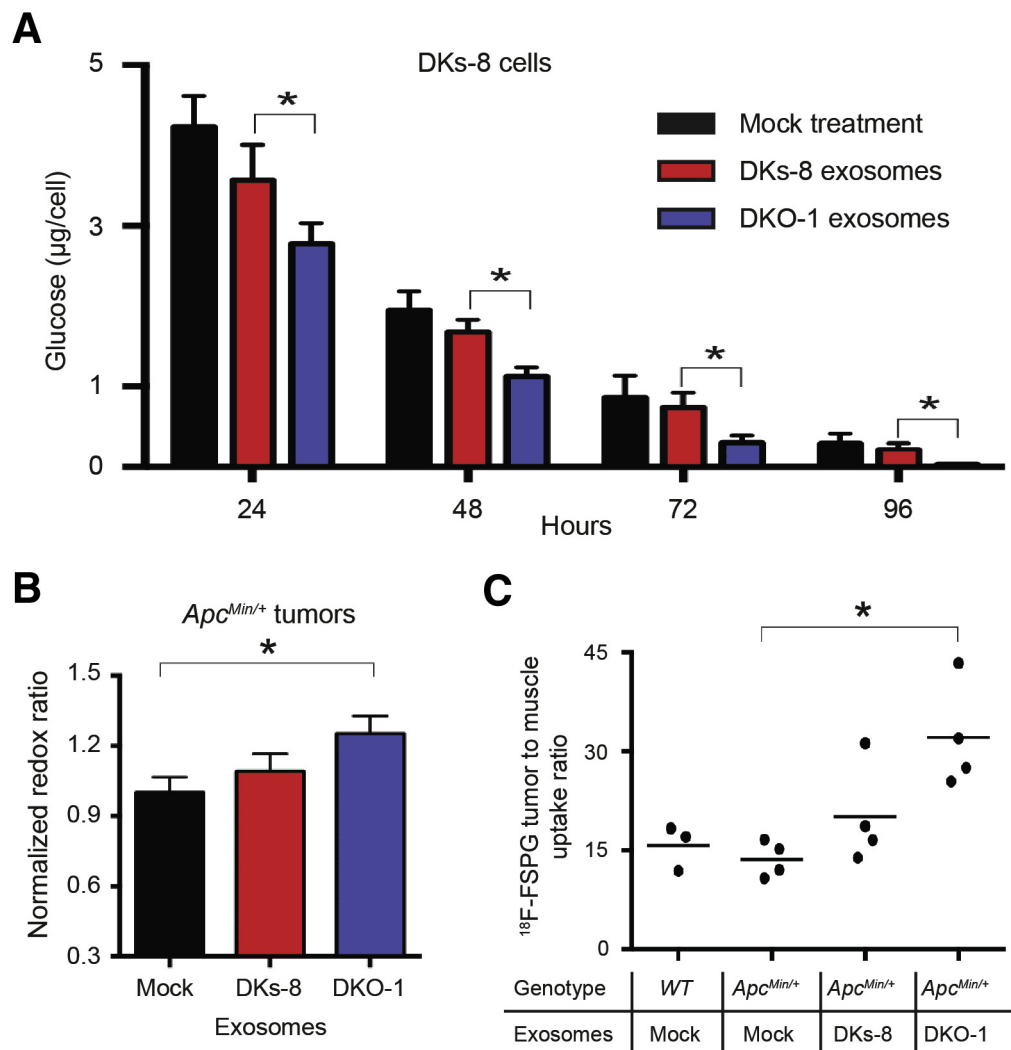


Figure 1. Mutant KRAS exosomes alter metabolism in vitro and in vivo. (A) Glucose consumption of exosome-treated DKs-8 cells (N = 3 in triplicate). Data are plotted as means ± SD. (B) Normalized redox ratio of exosome-treated *Apc^{Min/+}* colonic tumors. (C) ¹⁸F-FSPG tumor-to-muscle uptake ratio in WT or *Apc^{Min/+}* mice injected with exosomes analyzed by 1-way analysis of variance followed by a post hoc Tukey test. *P < .05.

the redox ratio in recipient normal mouse colonic cells cultured in Matrigel or on plastic (Supplementary Figure 2A–D) and WT KRAS DKs-8 cells (Supplementary Figure 2E).

To test whether mutant KRAS CRC exosomes function *in vivo*, we used the Adenomatous polyposis coli multiple intestinal neoplasia (*Apc^{Min/+}*) mouse model in which adenomas develop throughout the gastrointestinal tract. *Apc^{Min/+}* mice received intraperitoneal injections of DKs-8 or DKO-1 exosomes over 4 successive days. The redox ratio of tumors treated with DKO-1 exosomes was increased significantly (Figure 1B and Supplementary

Figure 3A), suggesting that treatment with these mutant KRAS exosomes increases aerobic glycolysis in recipient tumor cells. We also performed (S)-4-(3-[¹⁸F]-fluoropropyl)-L-glutamic acid (¹⁸F-FSPG) positron emission tomography imaging 2 hours after the last injection of exosomes. ¹⁸F-FSPG is a novel positron emission tomography tracer that follows the import of cystine by the glutamate/cystine antiporter *SLC7A11*, which is overexpressed in CRC.¹⁰ We found that *Apc^{Min/+}* mice injected with DKO-1 exosomes had a significant increase in ¹⁸F-FSPG uptake in the tumor region (Figure 1C and Supplementary Figure 3B and C),

suggesting that mutant KRAS CRC exosomes can alter tumor cell metabolism *in vivo*.

Levels of GLUT-1 were increased in DKO-1 exosomes, as well as in cell lysates (Figure 2A). To test whether these mutant KRAS exosomes contained functional GLUT-1, we measured ¹⁸F-fluorodeoxyglucose incorporation. After 1 hour, ¹⁸F-fluorodeoxyglucose uptake was significantly higher in DKO-1 exosomes (Figure 2B). The purified exosomes showed the characteristic cup-shaped morphology and size reported for exosomes (40–100 nm) (Supplementary Figure 4A and B). Further purification of exosomes on an

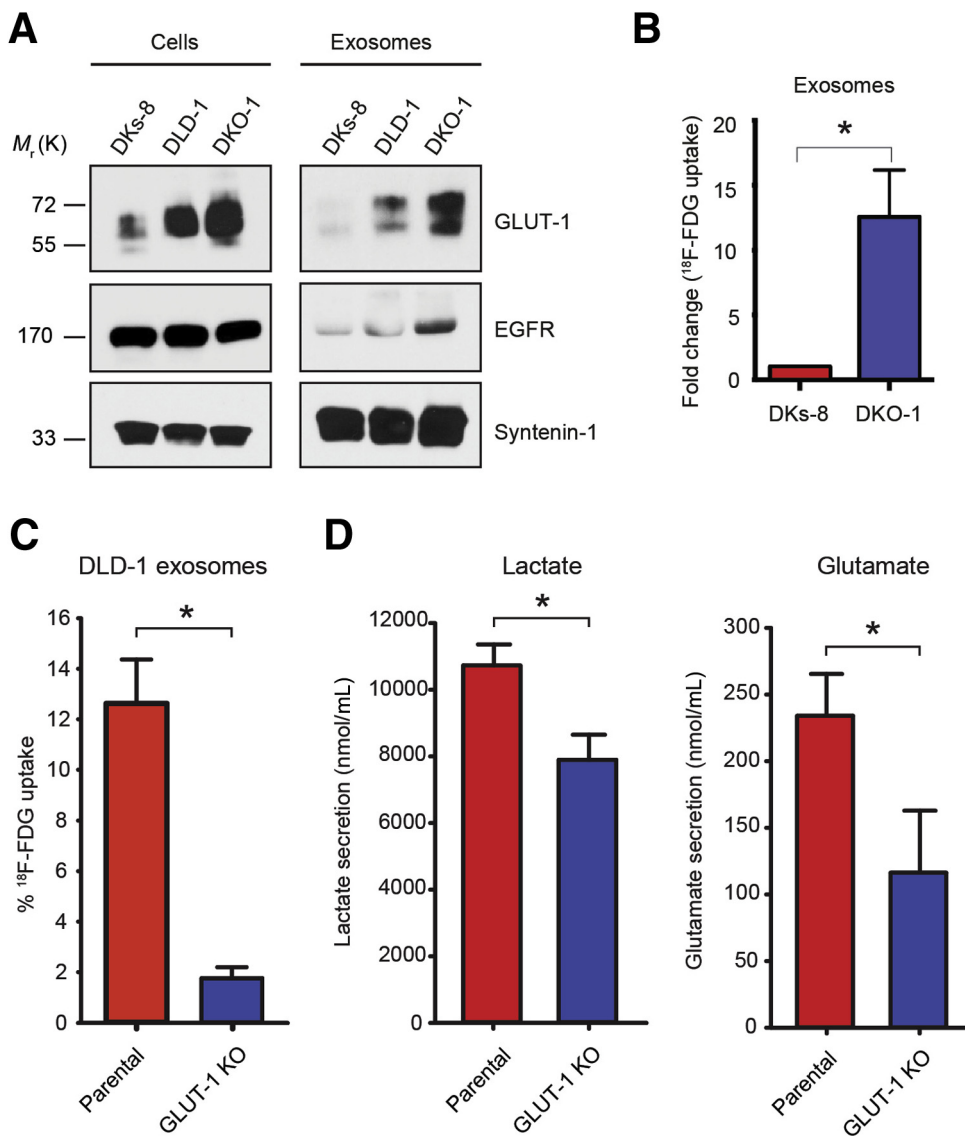


Figure 2. Exosomal GLUT-1 partially drives metabolic changes in recipient cells. (A) Immunoblot analysis of cells and exosomes. After normalization to syntenin-1, levels of GLUT-1 were increased 2.5- and 3.1-fold in cell lysates and 3.1- and 5.2-fold in exosomes of DLD-1 and DKO-1 cells, respectively. (B) Percent ¹⁸F-fluorodeoxyglucose (¹⁸FDG) uptake in exosomes isolated from DKs-8 and DKO-1 cells (N = 3 in triplicate). (C) Fold change of ¹⁸FDG uptake in exosomes isolated from parental and GLUT-1 KO DLD-1 cells. (D) nuclear magnetic resonance (NMR) determination of glutamate and lactate secretion in recipient DLD-1 GLUT-1 KO cells 43 hours after treatment with exosomes from parental DLD-1 or GLUT-1 KO cells (N = 2 in triplicate). Data are plotted as the means ± SD. *P < .05.

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