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Method paper

# Transcriptomics reveals tissue/organ-specific differences in gene expression in the starfish *Patiria pectinifera*

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#### ABSTRACT

Starfish (Phylum Echinodermata) are of interest from an evolutionary perspective because as deuterostomian invertebrates they occupy an "intermediate" phylogenetic position with respect to chordates (e.g. vertebrates) and protostomian invertebrates (e.g. Drosophila). Furthermore, starfish are model organisms for research on fertilization, embryonic development, innate immunity and tissue regeneration. However, large-scale molecular data for starfish tissues/organs are limited. To provide a comprehensive genetic resource for the starfish Patiria pectinifera, we report de novo transcriptome assemblies and global gene expression analysis for six P. pectinifera tissues/organs - body wall (BW), coelomic epithelium (CE), tube feet (TF), stomach (SM), pyloric caeca (PC) and gonad (GN). A total of 408 million high-quality reads obtained from six cDNA libraries were assembled de novo using Trinity, resulting in a total of 549,598 contigs with a mean length of 835 nucleotides (nt), an N50 of 1473 nt, and GC ratio of 42.5%. A total of 126,136 contigs (22.9%) were obtained as predicted open reading frames (ORFs) by TransDecoder, of which 102,187 were annotated with NCBI non-redundant (NR) hits, and 51,075 and 10,963 were annotated with Gene Ontology (GO) and Kyoto Encyclopaedia of Genes and Genomes (KEGG) using the Blast2GO program, respectively. Gene expression analysis revealed that tissues/organs are grouped into three clusters: BW/CE/TF, SM/PC, and GN, which likely reflect functional relationships. 2408, 8560, 2687, 1727, 3321, and 2667 specifically expressed genes were identified for BW, GN, PC, CE, SM and TF, respectively, using the ROKU method. This study provides a valuable transcriptome resource and novel molecular insights into the functional biology of different tissues/organs in starfish as a model organism.

#### 1. Introduction

Starfish are deuterostomian invertebrates belonging to the phylum Echinodermata that are recognized as fascinating animals with many features of special interest (Arnone et al., 2016). These include their status as a canonical example of a keystone species in ecology (Paine, 1966) and as model organisms for research on neuroendocrinology (Semmens et al., 2016), innate immunity (Franco et al., 2011), temporary adhesion (Hennebert et al., 2014), and tissue regeneration (Thorndyke et al., 2001).

Recently, transcriptome sequencing of emerging marine model

organisms has proven to be an efficient method for relatively low cost gene discovery and analysis of differential gene expression (Martin and Wang, 2011). For these reasons, several research groups have utilized high throughput sequencing technology for molecular level characterization of various biological processes in several starfish species, including Asterias anurensis (Richardson and Sherman, 2015), Asterias rubens (Semmens et al., 2013; Semmens et al., 2016), Acanthaster planci (Stewart et al., 2015) and Coscinasterias muricata (Gabre et al., 2015). However, these transcriptomic studies were limited to analysis of whole larvae or just one adult starfish tissue/organ. Starfish have many different tissues and organs that are responsible for a variety of biological

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Abbreviations: BW, body wall; CE, coelomic epithelium; TF, tube feet; SM, stomach; PC, pyloric caeca; GN, gonad; BUSCO, Benchmarking Universal Single-Copy Orthologs; KOG, EuKaryotic Orthologous Groups; ORFs, open reading frames; NR, non-redundant; GO, gene ontology; KEGG, Kyoto Encyclopaedia of Genes and Genomes; DEGs, differentially expressed genes; BP, biological process; CC, cellular component; MF, molecular function; FPKM, fragments per kilobase per million mapped reads; RSEM, RNA-Seq by expectation maximization; AIC, Akaike's information criterion; FDR, false discovery rate; PCA, principal component analysis

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processes. For example: 1. the mechanical state of the body-wall determines body stiffness and posture in starfish (Motokawa, 2011), 2. the coelomic epithelium is involved in wound healing, regeneration, and hematopoiesis (Gabre et al., 2015), 3. the stomach (pyloric and cardiac stomach) and pyloric caeca enable intake, digestion, absorption and storage of nutrients (Ferguson, 1964), 4. a multitude of tube feet enable locomotion, utilising secretion of adhesive materials for adhesion (Hennebert et al., 2011), and 5. gonads are, of course, essential for reproduction (Stewart et al., 2015). It is not known, however, if the physiological roles of different tissues/organs in starfish are reflected in their gene expression profiles. Therefore, it is of interest to comprehensively analyze differentially expressed genes (DEGs) in starfish tissues/organs.

The starfish species *Patiria pectinifera* is widely distributed in the northern Pacific Ocean and has been used as a model organism for studying different aspects of starfish physiology (Davydov et al., 1990; Kim et al., 2016; Mita et al., 2009) and is also of interest from economic and environmental perspectives (Jo et al., 2013; Popov et al., 2016). In this study, we characterise the *P. pectinifera* transcriptome by RNA-seq employing paired-end Illumina HiSeq<sup>TM</sup> 2500 sequencing technology and subsequent *de novo* assembly to generate a comprehensive set of reference contigs for gene discovery and for analysis of DEGs among six different tissues/organs. Our study provides a genetic resource for future comparisons with other echinoderm transcriptomes and for functional analysis of gene expression.

#### 2. Data description

#### 2.1. RNA isolation and illumina sequencing

Live specimens of the starfish species *Patiria pectinifera* (approximate diameter 8 cm) were collected at low tide from the coast of Cheongsapo of Busan, Korea (Table 1). Approval by the local institution/ethics committee was not required for this work because

Table 1

MixS descriptors

MixS descriptors and statistics for the sequencing and *de novo* assembly of the transcriptome of the starfish Patiria pectinifera.

experimental work on starfish is not subject to regulation and P. pectinifera is not an endangered or protected species. Six tissues/organs, gonad (GN), pyloric caeca (PC), coelomic epithelium (CE), stomach (SM), tube feet (TF), and body-wall (BW, excluding CE) were dissected from six individual specimens of P. pectinifera and then total RNA was extracted using RNeasy Total RNA Isolation kit (Qiagen, USA) according to the manufacturer's instructions. The concentration, quality, and integrity of RNA preparations were determined using a NanoDrop-2000 spectrophotometer (Thermo, USA) and a Bioanalyzer 2100 (Agilent Technologies, USA). Then the RNA preparations were disrupted into short fragments. Double-stranded cDNA was synthesized with sequencing adapters using Illumina TruSeq™ RNA Library Prep Kit v2 (San Diego, CA, USA) following the manufacturer's instructions. Finally, six RNA-seq libraries were subjected to paired-end sequencing with a read length of  $2 \times 101$  nucleotides on an Illumina HiSeq 2500 platform. Illumina HiSeq 2500 produced a total of 417,972,264 reads representing a total of 42,215,198,664 nucleotides from six tissues/organs, with the maximum number of reads (75,840,222) generated from the SM library and the minimum number of reads (63,360,640) generated from CE library (Table 1). The raw reads were deposited in the Sequencing Read Archive (SRA) of NCBI with accession numbers SRR5229423, SRR5229424, SRR5229425, SRR5229426, SRR5229427, SRR5229428 for TF, SM, CE, PC, GN and BW, respectively.

#### 2.2. De novo assembly and functional annotation

After sequencing was completed, Illumina TruSeq adapter sequences, low-quality bases, and reads under minimum length were trimmed from the reads using CutAdapt v1.10 with -q 20, -m 30 parameters (Martin, 2011). Then, reads were filtered into clean reads. From these reads, contamination removal was performed by Bowtie2 v2.2.9 against the bacterial and ocean metagenome databases downloaded from NCBI (ftp://ftp.ncbi.nlm.nih.gov/genomes/Bacteria, ftp:// ftp.ncbi.nlm.nih.gov/genomes/Bacteria\_DRAFT, ftp://ftp.ncbi.nlm.nih.

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Item		Descr	ription		Item		Description	
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Sequencing (Illumina HiSeq2500; paired-end, $2 \times 101$ ) stats								
		Tissues						
Process		Body-wall	Coelomic	Gonad	Pyloric caeca	Stomach	Tube feet	Total
			epithelium					
Raw read	Number	68,676,738	63,360,640	69,973,024	68,743,218	75,840,222	71,378,422	417,972,264
	Total size	6.936.350.538	6,399,424,640	7.067.275.424	6.943.065.018	7.659.862.422	7,209,220,622	42.215.198.664
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Adapter trimming	Number	67 107 554 (97 7)	61 920 652	68 288 434 (97 6)	67 210 592	74 127 038 (97 7)	69 813 734 (97 8)	408 468 004 (97 7
ridupter trimining	<sup>a</sup> (%)	07,107,001 (77.77)	(97 7)	00,200,101 (57.0)	(97.8)	/ 1,12/,000 ()/./)	0,010,701 (77.0)	100, 100,001 (97.7
Contamination	Number	67 068 478 (97 7)	61 888 058	68 209 202 (97 5)	67 180 392	74 104 076 (97 7)	69 717 310 (97 7)	408 167 516 (97 7
removal	<sup>b</sup> (%)	07,000,470 (77.7)	(07.7)	00,209,202 (97.3)	(07.7)	/4,104,0/0 ()/./)	0,717,510 (77.7)	400,107,510 (57.7
<sup>a</sup> (0/2) (number of c	(70) dontor trimm	ad road /number of r	(97.7)		(97.7)			
<sup>b</sup> (0()) (number of a	adapter trimin	ieu reau/number of r	aw read) × 100		0			
(%), (number	of containing	ation removed read /	number of adapte.	$r trimined read) \times 10^{\circ}$	0			
Assembly (De novo assembly; Trinity 2.0.6) stats								
Process	Number of	Total size (bp)	Mean size (bp)	Minimum size (bp)	Maximum size	N50 (bp)	GC, %	Data accessibility
	contigs				(bp)			DDBJ/EMBL/
					· • • ·			GenBank
De novo	549.598	459.064.184	835	224	35.675	1473	42.5	GFOO000000000
	2.12,320	,			,			

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