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## Review

## A short review on the implications of base excision repair pathway for neurons: Relevance to neurodegenerative diseases

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## ABSTRACT

Oxidative DNA damage results from the attack by reactive oxygen and nitrogen species (ROS/RNS) on human genome. This includes base modifications such as oxidized bases, abasic (AP) sites, and single-strand breaks (SSBs), all of which are repaired by the base excision repair (BER) pathway, one among the six known repair pathways. BER-pathway in mammalian cells involves several evolutionarily conserved proteins and is also linked to genome replication and transcription. The BER-pathway enzymes, namely, DNA glycosylases (DGs) and the end-processing proteins such as abasic endonuclease (APE1), form complexes with downstream repair enzymes via protein–protein and DNA–protein interactions. An emerging concept for BER proteins is their involvement in non-canonical functions associated to RNA metabolism, which is opening new interesting perspectives. Various mechanisms that are underlined in maintaining neuronal cell genome integrity are identified, but are inconclusive in providing protection against oxidative damage in neurodegenerative disorders, main emphasis is given towards the role played by the proteins of BER-pathway that is discussed. In addition, mechanisms of action of BER-pathway in nuclear vs. mitochondria as well as the non-canonical functions are discussed in connection to human neurodegenerative diseases.

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Abbreviations: AD, Alzheimer's disease; AP site, apurinic/aprimidinic site; APE1, apurinic/aprimidinic endonuclease 1; A $\beta$ , amyloid beta; BER, base excision repair; CaMKII, Ca<sup>2+</sup>/Calmodulin-Dependent Protein Kinase II; CDK, cyclin-dependent kinase; CREB, cAMP response element-binding; IP, immunoprecipitate; NFT, neurofibrillary tangles; PARP-1, poly [ADP-ribose] polymerase 1; Ref-1, redox effector factor-1; SSB, single-strand break repair; SSBs, single-strand breaks; TF, transcription factor; MGMT, O<sup>6</sup>-methylguanine DNA methyltransferase; NER, nucleotide excision repair; NHEJ, nonhomologous end joining; XP, Xeroderma pigmentosum; CS, Cockayne syndrome; TTD, trichothiodystrophy; CNS, central nervous system; GEN1, gap endonuclease; XRCC1, X-ray repair cross-complementing 1; PCNA, proliferating cell nuclear antigen; RPA, replication protein A; DG, DNA glycosylase; PNKP, polynucleotide kinase 3'-phosphatase; ROS/RNS, reactive oxygen species/reactive nitrogen species; UV-A, ultraviolet ray-A; ND, neurodegenerative disease; mtDNA, mitochondrial DNA; nuDNA, nuclear DNA; FEN, flap endonuclease; TDG, thymine-DNA glycosylase; OGG1, 8-oxoguanine glycosylase; NEIL1, endonuclease VIII-like 1; UDG1, uracil-DNA glycosylase; SN-BER, short patch-base excision repair; LP-BER, long patch-base excision repair; HhH, helix-hairpin-helix; MUTYH, mutY homolog *E. coli*; MPG, 3-methyladenine-DNA glycosylase; BRCT, BRCA1 C-terminus; BBB, blood-brain barrier; MMS, methyl methanesulfonate.

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## 71 1. Introduction

72 DNA damage is balanced with repair in a homeostatic process, and  
73 when damage exceeds the repair, final outcome may be cell cycle arrest,  
74 apoptosis or genome mutation. DNA damage occurs due to various  
75 external and internal causes. In neuronal cells, most of the DNA damage  
76 is repaired by the base excision repair (BER) pathway, as neuronal cells  
77 are partially differentiated cells and replication derived repair is not  
78 possible in these cells. It is very important to study the mechanisms  
79 and enzymes involved in BER-pathway for neuronal survival. In neuro-  
80 nal cells, the role of different proteins in BER-pathway in both nucleus  
81 and mitochondria is not fully elucidated, yet.

## 82 2. Roles of base excision repair in oxidative DNA damage repair

83 Mammalian cells are constantly exposed to stress from external and  
84 internal agents. Oxidative stress is a common feature of all stresses, and  
85 to maintain cellular integrity, mammalian cells have evolved different  
86 repair mechanisms. Various chemical events may lead to DNA damage  
87 including hydrolysis and exposure to reactive oxygen substances  
88 (Nilsen et al., 2000) and other reactive metabolites (Bergamini et al.,  
89 2004). In normal dividing cells, the DNA damage is sensed through dif-  
90 ferent cell cycle checkpoints, and if DNA damage occurs before cell divi-  
91 sion then cell division stops in order to repair the damage (Houtgraaf  
92 et al., 2006). DNA repair and its mechanism in neurons are one of the  
93 less studied area. DNA damage repair mechanisms in neuronal cells  
94 are different, as neurons are in postmitotic stage and not able to divide  
95 (Fishel et al., 2007; Nospikel and Hanawalt, 2002). The repair mecha-  
96 nisms in neuronal cells involve different pathways including: direct rever-  
97 sal ( $O^6$ -methylguanine DNA methyltransferase; MGMT), mismatch  
98 repair, double strand break (DSB) repair via homologous recombination  
99 (HR) and non-homologous end joining (NHEJ). The mismatch occurs  
100 due to wrong incorporation of bases and deamination of bases due to  
101 oxidative damage, but these repair mechanisms are not highly efficient  
102 in neurons as they are terminally divided cells and cell division is not  
103 possible in these cells (Kruman, 2004). Nucleotide excision repair  
104 (NER) was initially the main focus of research as most of the neuronal  
105 damage occurs due to the defect in NER (Hitomi et al., 2007). Xeroderma  
106 pigmentosum (XP), Cockayne syndrome (CS) and trichothiodystrophy  
107 (TTD) are the family of sunlight sensitive disease and caused by ineffi-  
108 ciency in the components of the NER pathway (Laposa and Cleaver,  
109 2001). BER is the primary nuclear and mitochondrial DNA repair path-  
110 way for small base modifications such as alkylation, deamination and  
111 oxidation, and is thought to play a critical role during development  
112 and maintenance of the central nervous system, CNS (Chen et al., 2000).

113 BER is the main repair pathway in postmitotic cells, in which simple  
114 base modifications are more likely to occur than major damages to DNA  
115 (Rao, 2007). BER is a repair pathway predominant for the processing of  
116 small base lesions, derived from oxidation and alkylating damage and  
117 genotoxic chemicals (Hegde et al., 2008). The overall estimate of  
118  $10^4$  base damages/mammalian cells/day underlines the importance of  
119 BER (Lindahl, 1993). The BER pathway mainly requires four types of  
120 enzymes, DNA glycosylase, AP endonuclease, DNA polymerase and  
121 DNA ligases (Dantzer et al., 2000; Hegde et al., 2008, 2010). In addition,

proteins like XRCC1, PCNA and RPA, are also required (Mitra et al.,  
2001). The basic BER reaction comprises three steps: (1) base lesion  
recognition and excision by a DNA glycosylase, followed by cleavage of  
the resulting apurinic/aprimidinic (AP) site in a concerted reaction by  
the DG itself (for bifunctional DG) or by APE1 (for monofunctional DG)  
(Hegde et al., 2012; Kulkarni et al., 2008; Mitra et al., 2001; Pena-Diaz  
et al., 2012); (2) cleaning of 3' blocked termini at the strand break  
by APE1 and/or polynucleotide kinase 3' phosphatase (PNKP) and  
5' blocking phosphodeoxyribose by DNA polymerase- $\beta$  (pol- $\beta$ ) and  
gap filling by a DNA polymerase; and (3) nick sealing by DNA ligase to  
complete the repair [Fig. 1] (Hegde et al., 2008, 2012). Notably, the en-  
zymes involved in BER-pathway are highly conserved phylogenetically.

## 3. Sources of endogenous and exogenous DNA damage to neuronal cells

Damage to DNA can be induced by several chemical reactive  
species and physical agents or may occur spontaneously through intrin-  
sic instability of chemical bonds in DNA (Table 1). Even under normal  
physiologic conditions, DNA is continuously being damaged (Altieri F  
Fau-Grillo et al., 2008). These attacks can be divided into two broad  
categories: exogenous and endogenous (Altieri F Fau-Grillo et al.,  
2008). Exogenous and environmental sources of oxidation relate to  
specific exposures of the organism to ionizing radiations like X,  $\gamma$  and  
cosmic rays. Apart from that, radon decay, oxidizing chemicals and  
UV-A solar light are also involved in these types of DNA damage  
(Branzei and Foiani, 2008). Brain constitutes around 2–3% of total  
body mass, but it utilizes 20% of body basal oxygen supply (Marlatt  
et al., 2004). Intracellular (endogenous) sources of oxidative stress are  
primarily produced by  $O_2$  metabolism (electron transport chain),  
immune responses and inflammation (Lee and Wei, 2007). The result  
is production of reactive oxygen/nitrogen species (ROS/RNS) which  
react with the DNA and produce various lesions and adducts (Raffoul  
et al., 2012). DNA damage can be induced also in neighboring or distant  
cells via an inflammatory-based mechanism, and the first barrier  
defense is accomplished through different enzymatic antioxidants,  
which act as scavengers of free radicals (Lobo et al., 2010).

The occurrence of neurodegenerative disease is a slow, progressive  
and irreversible degeneration of neurons and synapse of selected  
areas of nervous system (Fitzner and Simons, 2010). Neurodegenerative  
diseases (NDs) are caused by multifactorial etiologic causes which  
include genetic, environmental or endogenous insults (Singh et al.,  
2013). NDs are classified according to genetic factors and the major  
depositing compounds, so NDs are also known as protein misfolding  
diseases or proteinopathies (Jellinger, 2010). In Alzheimer's and  
Parkinson's diseases (AD and PD), a large body of evidence shows dam-  
aged energy metabolism and around 50% reduction in mtRNA content  
and is likely to reduce oxidative phosphorylation (Jellinger, 2010). In  
PD, iron is deposited in the substantia nigra (SN), and increased iron de-  
posits in the SN may have genetic and non-genetic causes (Gerlach et al.,  
2006). In PD brain, increased iron is often accompanied by decreased  
ferritin synthesis, resulting in free iron overload (Dexter et al., 1990).  
The Cu(II) in submicromolar and Fe(II/III) in micromolar concentrations  
specifically inhibit the NEILs and not OGG1 (Hegde et al., 2010). Both

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