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Q1 Vaginal drug distribution modeling

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ABSTRACT

This review presents and applies fundamental mass transport theory describing the diffusion and convection driven mass transport of drugs to the vaginal environment. It considers sources of variability in the predictions of the models. It illustrates use of model predictions of microbicide drug concentration distribution (pharmacokinetics) to gain insights about drug effectiveness in preventing HIV infection (pharmacodynamics). The modeling compares vaginal drug distributions after different gel dosage regimens, and it evaluates consequences of changes in gel viscosity due to aging. It compares vaginal mucosal concentration distributions of drugs delivered by gels vs. intravaginal rings. Finally, the modeling approach is used to compare vaginal drug distributions across species with differing vaginal dimensions. Deterministic models of drug mass transport into and throughout the vaginal environment can provide critical insights about the mechanisms and determinants of such transport. This knowledge, and the methodology that obtains it, can be applied and translated to multiple applications, involving the scientific underpinnings of vaginal drug distribution and the performance evaluation and design of products, and their dosage regimens, that achieve it.

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1. Introduction

The human vagina is a fibromuscular tract that connects the vulva to the cervix and uterus and thence the organs of the upper reproductive tract [1]. It is about 10–15 cm long, and its lumen is largely collapsed, resulting in a relatively flat cross section (width about 2–3 cm), except at the innermost region, the fornix. Its net surface area is about 80–110 cm². Its mucosa consists of two layers: a stratified squamous epithelium, that varies in thickness during the menstrual cycle from about 200–400 μm; and a lamina propria (connective tissue, also termed the stroma) that is about 2.5–3 mm thick. The stroma is about 10% vasculature by volume (there is no vasculature in the epithelium), and it contains host cells that HIV virions can infect (e.g. CD4⁺ T cells and macrophages). Below the stroma are muscular and outer connective tissue layers. The vagina does not secrete fluid per se. Mucus from the cervix leaks down through the external cervical os into the floor of the fornix, primarily at midcycle. It may then be distributed in a thin layer over the epithelial surfaces of the vaginal canal, although details of this are not well understood. Sexual stimulation increases the pressure in the vasculature in the stroma, resulting in transudation of fluid through the epithelium to its surface (“lubrication”) [2]. We have only approximate knowledge of the quantitative amount and

distribution of ambient fluid in the vagina [3]. It is maximal under conditions of maximum estrogen presence in the vaginal environment (e.g. at the midportion of the menstrual cycle) and minimal in conditions of low estrogen and high progesterone (viz. in the luteal phase of the cycle, and post menopause; here the epithelium is thinnest). The maximum volume of ambient vaginal fluid is believed to about around 1–2 ml (Fig. 1).

Delivery of drugs via the vagina has been implemented for many years, for many purposes [4]. Commercial and in-development vaginal products now introduce a variety of drugs intended for systemic delivery (e.g. contraceptive hormones and prostaglandins [5]) and topical delivery (e.g. spermicides, agents against urinary tract infections and candida infections, anti-bacterial vaginosis medications, labor inducing agents, etc. [6–10]). At present there is much activity directed at development of products that deliver drugs—termed *microbicides*—which act topically in the vaginal environment to inhibit infection by sexually transmitted HIV and/or other pathogens, e.g. human papilloma virus (HPV) and herpes simplex virus (HSV) [11–13]. Complementary approaches have focused on delivering antimicrobial antibodies. This application of drug delivery was pioneered by Saltzman and colleagues in the mid 1990s [14,15]. Drug delivery via the vagina is currently implemented or being developed via administration of a range of dosage forms to the vaginal canal: monolithic solid materials (e.g. intravaginal rings, IVRs) and soft, semi-solid materials (e.g. gels, creams, suppositories, films, non-woven porous textile materials, dissolving tablets, and fiber-woven meshes) [16–18]. A number of review articles have already

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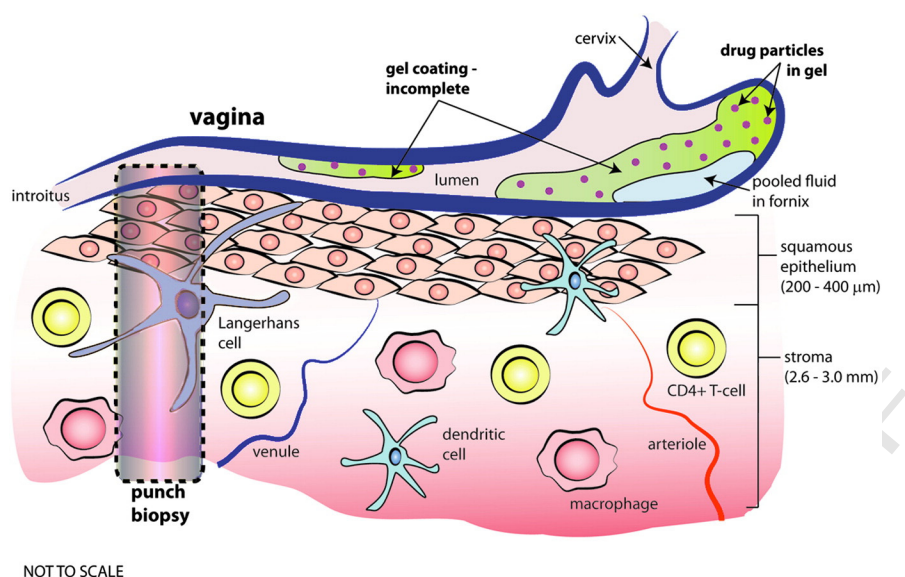


Fig. 1. Drawing of human vaginal canal and mucosal tissue (not to scale). The canal contains gel that partially coats the mucosal surface (anti-pathogenic microbicide molecules are depicted as red dots), and there is semen in the luminal space not occupied by gel. On the left is a column that represents the tissue collected by a punch biopsy, typically used to collect tissue for measuring drug concentration in pharmacokinetic studies.

85 been written about vaginal drug delivery [6–10]. For the most part,
 86 these have focused upon the compositions and properties of the
 87 various delivery systems, with reference to the delivered drugs and
 88 vaginal physiology and anatomy. There has been relatively little
 89 attention to the use and potential impact of modeling of the drug
 90 delivery performance per se of the various products. To be sure, the
 91 benefits of modeling vary, depending upon the site and mechanism
 92 of action of the delivered drug. For example, therapeutic delivery of
 93 antifungal compounds by creams, gels or suppositories against
 94 candida infections essentially involves coating, if not filling, the
 95 vagina canal with drug that will subsequently mix with ambient
 96 vaginal fluids and complete the transvaginal distribution process
 97 over time. Maximizing retention of drug on the mucosal surfaces,
 98 e.g. via mucoadhesive agents, is important here. However, the time
 99 constraint on minimizing the interval before onset of therapeutic
 100 action is not acute. In contrast, prophylactic delivery of topically
 101 acting anti-HIV microbicide drugs (to vaginal fluids and/or mucosal
 102 tissues) is acutely time and space dependent. In a sense, it is a race
 103 against the arrival of infectious virions from semen to sites, distri-
 104 buted throughout the length of the mucosa, that are vulnerable to
 105 infection. The goal of drug delivery for prophylaxis is to rapidly
 106 achieve prophylactic concentrations of drugs at target sites, and to
 107 achieve and maintain a high level of prophylactic action as long as
 108 possible. Here, the value of modeling can be substantial, viz. it can
 109 predict the time interval after product application at which pro-
 110 phylactic concentrations of drug(s) are achieved and sustained, at
 111 the level of such prophylaxis, at target sites. More fundamentally, it
 112 can help guide the rational design and evaluation of candidate
 113 products. This was appreciated by Saltzman and colleagues in their
 114 studies of delivery of anti-HSV IgG antibodies by EVA disks [15].
 115 They created a pharmacokinetic compartmental model of antibody
 116 concentration (volume averaged) in the vagina by diffusion out
 117 from a circular disk, and they applied the model to experimental
 118 data in the mouse. In follow up, Saltzman presented two additional
 119 model analyses: bolus delivery in the presence of vaginal fluid
 120 flow; and diffusion controlled drug release from an intravaginal
 121 ring (see section below on IVR modeling) [19,20].

122 Since those pioneering analyses (over a decade ago), there has
 123 been relatively little modeling of microbicide drug delivery to the
 124 vaginal (and rectal) environments. This has fundamentally limited
 125 our ability to understand the determinants of that performance and

to incorporate such knowledge into objective product design and
 dosing schemas. The review article here is intended to help fill this
 gap. Our focus is upon drug delivery systems that introduce active
 pharmaceutical ingredients (APIs) that act topically within the
 vaginal environment (fluids and tissues), with particular emphasis
 upon microbicides. Quantitative illustrations of applications of
 modeling are presented for a leading microbicide drug tenofovir,
 although the general methodological approach extends more broad-
 ly to the spectrum of microbicide drugs being evaluated. Recently,
 some aspects of such modeling were reviewed [18].

In this article, we present a summary of the nature of determi-
 nistic modeling of vaginal drug delivery and distribution, achieved
 by multiple dosage forms. We then describe several specific applica-
 tions of modeling that can have value to pharmacological and
 biological understanding of vaginal product functionality: (1) translating
 modeling of drug distribution per se (pharmacokinetics) to inform-
 ation about drug functionality (pharmacodynamics); (2) comparing
 effects of different gel dosage regimens; (3) interpreting conse-
 quences of gel aging during storage; (4) comparing modeling
 results for different dosage forms, viz. gels vs. intravaginal rings;
 and (5) planning and interpreting studies across multiple species,
 comparing humans to smaller animals.

The approach reviewed here is complementary to, but different in
 many ways from traditional empirical PBPK modeling, which has
 been applied in the context of vaginal drug delivery [21–24]. Both
 approaches implement conservation of drug mass principles, but
 deterministic modeling embodies additional biophysical principles
 that govern *how* drugs migrate within and between compartments.
 Our descriptions of how the modeling works necessarily involve
 technical language and use of equations to characterize cause and
 effect in the examples presented. We hope the information and
 illustrations of modeling here will be helpful to the broader field of
 vaginal drug delivery, and be useful in connecting scientific under-
 standing and research to product development and dosing specifica-
 tion, e.g. in the current microbicide pipeline.

2. Building deterministic models

Distribution of a drug throughout the vaginal environment is a
 mass transport process in which several types of active forces (e.g.
 squeezing by the walls of the canal, pressure gradients imposed on

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