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Protein Sequences Recapitulate Genetic Code Evolution 1

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ABSTRACT

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Several hypotheses predict ranks of amino acid assignments to genetic code's codons. Analyses here show that 18 average positions of amino acid species in proteins correspond to assignment ranks, in particular as predicted 19 by Juke's neutral mutation hypothesis for codon assignments. In all tested protein groups, including co- and 20 post-translationally folding proteins, 'recent' amino acids are on average closer to gene 5' extremities than 21 'ancient' ones. Analyses of pairwise residue contact energies matrices suggest that early amino acids 22 stereochemically selected late ones that stabilize residue interactions within protein cores, presumably produc- 23 ing 5'-late-to-3'-early amino acid protein sequence gradients. The gradient might reduce protein misfolding, also 24 after mutations, extending principles of neutral mutations to protein folding. Presumably, in self-perpetuating 25 and self-correcting systems like the genetic code, initial conditions produce similarities between evolution of 26 the process (the genetic code) and 'ontogeny' of resulting structures (here proteins), producing apparent teleon- 27 omy between process and product. 28

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1. Introduction 42

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The structure of biological molecules includes imprints of ancient 43 evolution at life's dawn. For example, comparisons between protein 44 and RNA structures suggest affinities between viruses and hypothetical 45 bacterial-like cellular ancestors (as described for protein structural fam-46 ilies, [61], Nasir et al. 2017; and for RNA secondary structures, [101]). 05 The ribosome's structure testifies to even more ancient events: ribo-48 49 somal protein amino acids interact preferentially with ribosomal 50 RNA trinucleotides that correspond to that amino acid's assigned anticodon(s) according to the standard genetic code [42]. This striking fossil-51 ization of the process that determined some codon-amino acid 52 53 assignments in the ribosome's structure confirms that at least some 54 codon-amino acid assignments result from stereochemical affinities between RNA and amino acids [118-120]. 55

1.1. Steps in the Evolution of the Genetic Code and the Translational 56 **Apparatus** 57

58 Johnson and Wang [42] suggest that several processes structured 59 the genetic code, meaning determined codon-amino acid assignments.

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Indeed, structurally simple amino acids tend to associate with rRNA 60 nucleotide triplets corresponding to their genetic code codon assign- 61 ments, while complex amino acids associate with their anticodons (ste- 62 reochemical complexity according to Dufton [19]). This indicates a 63 primary phase of direct codon-amino acid contact, and secondarily evo- 64 lution of mRNA, anticodon and from there the proto-tRNA [97]. 65

Several hypotheses predict the order of inclusion of amino acids in 66 the genetic code. These orders tend to be consensual among hypotheses, 67 and usually consider that structurally simple amino acids were included 68 early, and complex one's late [36,56,113,114]. Considering 40 hypothe- 69 ses about the inclusion order of amino acids in the genetic code 70 reviewed by Trifonov [114], the strength of association between 71 amino acids and their anticodons in rRNA (data from [42], therein figure 72 1) increases with their order of inclusion in the genetic code. This corre-73 lation is strongest with the inclusion order predicted by the tRNA-Urgen 74 hypothesis ([20,21], here Fig. 1). 75

1.2. Imprints of the Genetic Code Evolution in Modern Protein Sequences 76

Above observations about the ribosome's structure suggest that 77 imprints of the genetic code's evolution might remain also in protein 78 structures. Here I test the hypothesis that the inclusion order of amino 79 acids in the genetic code correlates with average positions of amino 80 acids in proteins. 81

This working hypothesis is derived from principles of the biogenetic 82 law or Meckel-Serres law, formulated by Haeckel as 'ontogeny recapit- 83 ulates phylogeny' [50]. As in that evo-devo hypothesis, the history of a 84

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Fig. 1. Strength of association of amino acids with ribosomal RNA triplets corresponding to their anticodons in the ribosome's structure, based on contacts between proteins and rRNAs in crystallized ribosomes [42], as a function of the order of inclusion of amino acids in the genetic code according to the tRNA Urgen hypothesis which has only 12 ranks (all 'late' amino acids get rank 12, [20,21,114]). Association strengths are ratios between observed numbers of amino acid contacts with anticodon triplets and expected random contacts, after data in Fig. 1 of Johnson and Wang [42]. Amino acids are classified according to three levels of structural complexity [19]: low (hollow circles), intermediate (gray circles) and high (filled triangles). The latter group would include cysteine, for which the ribosome's structure does not include contacts between residues and rRNA.

85 process might be imprinted in the structures produced by that process [44]. The reason to expect this apparent teleonomy frequently observed 86 in biological processes is that self-organizing and self-perpetuating pro-87 88 cesses such as the genetic code are by definition self-correcting [49]. 89 Structures resulting from early historical initial conditions are frequently conserved or recovered by resulting processes and structures. 90 Hence historical/evolutionary processes would be conserved as 91 imprints in modern structures because self-corrections towards the 92 93 least error-prone structures conserve or recover the same initial struc-94 tures/constraints. Accordingly, protein structures should also reflect 95 the evolution of the genetic code.

96 1.3. Evolution for Coding Versatility

The genetic code evolved to include more complex amino acids, 97 98 which are also more diverse in physicochemical terms than randomly 99 selected potential amino acids [31,40,67]. Directional evolution of genetically coded amino acids towards diversification and greater com-100 101 plexity corresponds to the most recently integrated amino acids in the 102 genetic code, selenocysteine and pyrrolysine [122], complex amino 103 acids with peculiar properties (i.e. selenocysteine includes a selenium atom (doesn't occur in other natural amino acids) where cysteine has 104 a sulfur atom (occurs only in one other natural amino acid)). 105

This suggests constraints towards increasing the genetic code's ver-106 satility for diverse types of specialized proteins. The evolutionary need 107 108 to develop proteins with new functions would have driven inclusion of complex and physicochemically outstanding amino acids. Presum-109 ably, RNA secondary structure-based punctuation signals initiated 110 translation before the genetic code assigned start codons [22,70]. The 111 presumably late assignment of methionine, a structurally complex and 112 'special' amino acid, to initiation codon(s) would suggest that 'late' 113 amino acids would tend to be coded close to gene 5' extremities, and 114 115 ancient amino acids closer to their 3' extremities.

The working hypothesis expects that the genetic code evolved to 116 include complex amino acids to stabilize protein structures, beyond 117 increasing the diversity of potentially coded proteins. Predictions are 118 tested versus lack of bias in average locations of amino acid species in 119 genes/proteins. 120

121

2. Materials and Methods

Analyses focus on eight groups of proteins, seven from the 122 Escherichia coli proteome (downloaded from GenBank entry 123 NC_002695). The two groups consist of all tRNA synthetases of 124 Escherichia coli (as used previously, [92]), subdivided in tRNA synthe- 125 tase class I and class II (10 amino acid species per class, 10 proteins 126 for class I and 13 for class II (including both subunits alpha and beta 127 for tRNA synthetases Phe and Gly)). Class II tRNA synthetases are 128 completed by the tRNA synthetase for pyrrolysine found in some 129 archaea [69,109]. The tRNA synthetases are chosen because these con- 130 served proteins essential to translation occur in all organisms [66,74], 131 including some viruses (Megavirales, [1,2,71,75]), and because within 132 each class they are related among each other, facilitating comparative 133 analyses [30,60,65] The two tRNA synthetase classes differ in their 134 structures: class I are usually monomeric proteins with a Rossman 135 fold catalytic domain. Class II tRNA synthetases are usually di- or 136 multimeric with an anti-parallel betasheet fold flanked by alpha 137 helices 138

Other protein groups from *E. coli*'s proteome are: 67 ribosomal 139 proteins, 36 polymerases, 119 membrane-linked proteins. Using predic- 140 tions on *E. coli* protein folding modes [15], a group of 63 proteins folding 141 cotranslationally is compared with another group of 101 proteins fold- 142 ing post-translationally. These were chosen from a longer protein list 143 because predicted folding mode in these proteins does not vary with 144 specific conditions as computationally tested by Ciryam et al. [15]. 145 Identities and sequences of the 408 analyzed *E. coli* proteins are avail- 146 able in the supplementary data. The *E. coli* proteome is translated from 147

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