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## Research paper

PEGylated  $\gamma$ -tocotrienol isomer of vitamin E: Synthesis, characterization, *in vitro* cytotoxicity, and oral bioavailabilityAhmed Abu-Fayyad<sup>a</sup>, Fathy Behery<sup>a</sup>, Asma Sallam<sup>a</sup>, Saeed Alqahtani<sup>a</sup>, Hassan Ebrahim<sup>a</sup>,  
Khalid A. El Sayed<sup>a</sup>, Amal Kaddoumi<sup>a</sup>, Paul W. Sylvester<sup>a</sup>, Jennifer L. Carroll<sup>c,d</sup>,  
James A. Cardelli<sup>c,e</sup>, Sami Nazzal<sup>a,b,\*</sup><sup>a</sup> College of Health and Pharmaceutical Sciences, School of Pharmacy, University of Louisiana at Monroe, Monroe, LA, USA<sup>b</sup> College of Pharmacy, Taipei Medical University, Taipei, Taiwan<sup>c</sup> Department of Microbiology and Immunology, Louisiana State University Health Sciences Center, Shreveport, LA, USA<sup>d</sup> Department of Biochemistry and Molecular Biology, Louisiana State University Health Sciences Center, Shreveport, LA, USA<sup>e</sup> Feist-Weiller Cancer Center, Louisiana State University Health Sciences Center, Shreveport, LA, USA

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## ABSTRACT

Vitamin E refers to a family of eight isomers divided into two subgroups, tocopherols and the therapeutically active tocotrienols ( $T_3$ ). The PEGylated  $\alpha$ -tocopherol isomer of vitamin E (vitamin E TPGS) has been extensively investigated for its solubilizing capacity as a nonionic surfactant in various drug delivery systems. Limited information, however, is available about the PEG conjugates of the tocotrienol isomers of vitamin E. In this study two PEGylated  $\gamma$ - $T_3$  variants with mPEG molecular weights of 350 ( $\gamma$ - $T_3$ PGS 350) and 1000 ( $\gamma$ - $T_3$ PGS 1000) were synthesized by a two-step reaction procedure and characterized by  $^1\text{H}$  NMR, HPLC, and mass spectroscopy. The physical properties of their self-assemblies in water were characterized by zeta, CMC, and size analysis. Similar physical properties were found between the PEGylated  $T_3$  and vitamin E TPGS. PEGylated  $T_3$  were also found to retain the *in vitro* cytotoxic activity of the free  $T_3$  against the MCF-7 and the triple-negative MDA-MB-231 breast cancer cells. PEGylated  $\gamma$ - $T_3$  also increased the oral bioavailability of  $\gamma$ - $T_3$  by threefolds when compared to the bioavailability of  $\gamma$ - $T_3$  formulated into a self-emulsified drug delivery system. No significant differences in biological activity were found between the PEG 350 and 1000 conjugates. Results from this study suggest that PEGylation of  $\gamma$ - $T_3$  represents a viable platform for the oral and parenteral delivery of  $\gamma$ - $T_3$  for potential use in the prevention of breast cancer.

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## 1. Introduction

Vitamin E from palm oil refers to a family of eight related, lipid soluble, compounds that are divided into two subfamilies known as tocopherols (T) and tocotrienols ( $T_3$ ). Each subgroup has  $\alpha$ ,  $\beta$ ,  $\gamma$ , and  $\delta$  isomers that differ in the number of methyl substitutions on the chromane moiety and the degree of saturation in their phytyl side chain [1].

Vitamin E was initially discovered as an essential factor for reproduction [2]. The antioxidant properties of this compound were later identified [3]. The poor water solubility of vitamin E,

however, limited its clinical use [4]. To overcome this limitation, a PEG conjugate of the  $\alpha$ -tocopherol isomer of vitamin E, known as d-alpha tocopheryl polyethylene glycol 1000 succinate or simply vitamin E TPGS was synthesized [5]. This water soluble derivative of vitamin E (1 g/10 mL [6]), was successfully used to treat children with low birthweight, and patients with cholestasis [7]. The tocotrienol isomers of vitamin E were only discovered during the 1960s [8–10]. Remarkably, it was only during the 1990s that the anticancer activities of tocotrienols were identified [11], which established a distinction in the health and therapeutic benefits between the tocotrienol and tocopherol isomers of vitamin E [12].

The  $\gamma$  tocotrienol isomer of vitamin E ( $\gamma$ - $T_3$ ) was found to have the most promising anticancer activity [11]. Several studies have reported that  $\gamma$ - $T_3$  isomer is the most abundant and active as anti-cancer agent [1,13–22]. Unfortunately, it has also been reported that it is difficult to obtain therapeutic levels of  $\gamma$ - $T_3$  in the blood and target tissues by simple oral administration due to its poor

\* Corresponding author at: Department of Basic Pharmaceutical Sciences, School of Pharmacy, College of Health and Pharmaceutical Sciences, University of Louisiana at Monroe, 1800 Bienville Drive, Monroe, LA 71201, USA. Tel.: +1 318 342 1726; fax: +1 318 342 1737.

E-mail address: [nazzal@ulm.edu](mailto:nazzal@ulm.edu) (S. Nazzal).

aqueous solubility and limited absorption and oral bioavailability [4,23–26]. To overcome these limitations, our laboratory was engaged over the past decade in developing lipid-based delivery systems for the oral and parenteral administration of  $\gamma$ -T<sub>3</sub> for use in breast cancer prevention and therapy [24–34]. Preliminary results demonstrated that incorporating  $\gamma$ -T<sub>3</sub> into lipid-based delivery systems enhances its oral bioavailability and potentiates its anticancer activity [24,26,32,34,35]. The enhancement in the oral bioavailability of  $\gamma$ -T<sub>3</sub>, nonetheless, was limited and was found to be as low as 5.6% at a dose of 1 mg/kg of body weight [25]. In addition to its low aqueous solubility, recent *in situ* permeability studies have revealed that the intestinal permeability of  $\gamma$ -T<sub>3</sub> is dose dependent when tested at 1, 2.5, 10, 25 and 50 mg/kg dose intervals [4,24–26]. The intestinal absorption of  $\gamma$ -T<sub>3</sub> is a saturable process that was found to be mediated by the Niemann-Pick C1-like 1 (NPC1L1) transporter [24–26]. Moreover, it was found that surfactants (i.e. Cremophor EL (BASF, Mount Olive, NJ) and Labrasol (Gattefossé, Paramus, NJ)) used in lipid-based formulations for the solubilization and oral delivery of  $\gamma$ -T<sub>3</sub> caused a 85% reduction in its cellular uptake due to the surfactant-induced inhibition of the NPC1L1 transporter protein resulting in nonlinear absorption kinetics [25,26]. In addition to passive diffusion and NPC1L1 mediated transport of  $\gamma$ -T<sub>3</sub>, vitamin E intestinal absorption was also found to be, at least in part, mediated by the scavenger receptor class B type I (SR-B1) protein [4,23,24].

In light of the aforementioned issues, we aimed to improve the oral bioavailability of  $\gamma$ -T<sub>3</sub> by synthesizing water soluble derivatives. We hypothesized that chemically conjugating  $\gamma$ -T<sub>3</sub> to a polyethylene glycol (PEG) moiety via a succinate linker would enhance the solubility, oral bioavailability, and therapeutic activity of  $\gamma$ -T<sub>3</sub>. PEGylation, the process of covalent attachment of PEG, has already been successfully used to enhance the activity of therapeutic proteins and small organic molecules, such as naloxegol (MOVANTI<sup>TM</sup>), a PEGylated derivative of naloxone, which was recently approved by the FDA [36,37]. While the oral bioavailability and pharmaceutical applications of vitamin E TPGS are well documented [5], the bioavailability and therapeutic activity of the PEG conjugates of the tocotrienol isomers have not been previously reported in the literature. Therefore, the objectives of the current study were to (a) synthesize and characterize two PEGylated  $\gamma$ -T<sub>3</sub> variants with m-PEG molecular weights of approximately 350 ( $\gamma$ -T<sub>3</sub>PGS 350) and 1000 ( $\gamma$ -T<sub>3</sub>PGS 1000); (b) examine the biological activity of the PEGylated  $\gamma$ -T<sub>3</sub> by evaluating their *in vitro* cytotoxic activity against two human breast cancer cell lines (MCF-7 and MDA-MB-231) and (c) determine the oral bioavailability of the PEGylated  $\gamma$ -T<sub>3</sub> in rats. To the best of our knowledge this marks the first report on the biological evaluation of PEGylated tocotrienol isomers of vitamin E and their self-assembly in aqueous media.

## 2. Materials and methods

### 2.1. Materials

Methoxy polyethylene glycols (mPEG 1000 and 350) were provided by INEOS Oxide (Antwerp, Belgium). Tocotrol<sup>TM</sup> L50P, a tocotrienol rich fraction of palm oil was purchased from Fuji Health Science, Inc. (Burlington, NJ). Triethylamine and succinic anhydride were from Alfa Aesar (Ward Hill, MA). Toluene and Chloroform-d (CDCl<sub>3</sub>) were from Acros (Bridgewater, NJ). p-Toluenesulfonic acid monohydrate (p-TsOH), hexane, sodium sulfate (Na<sub>2</sub>SO<sub>4</sub>) and sodium bicarbonate (NaHCO<sub>3</sub>) were from Avantor (Center Valley, PA). Ethyl acetate was from Pharmco-AAPER (Brookfield, CT). Acetonitrile and dichloromethane were from EMD Millipore (Temecula, CA). Silica gel was from Natland (Morrisville, NC). Pancreatin from porcine pancreas (Lipase not less than 8 USP

unites/mg, protease not Less than 100 USP unites/mg, and amylase not less than 100 USP unites/mg) was from Sigma Aldrich (St. Louis, MO). All chemicals and solvents were of reagent grade or higher and were used as supplied without further modification.

### 2.2. Extraction of $\gamma$ -Tocotrienol from Tocotrol<sup>TM</sup> L50P

$\gamma$ -T<sub>3</sub> was extracted from a 500 g batch of Tocotrol<sup>TM</sup> L50P, a palm oil fraction that contains approximately 43% tocotrienol isomers of which 40% are  $\gamma$ -T<sub>3</sub>. The T<sub>3</sub> rich oil was chromatographed on open column containing 1.5 kg silica gel. Initially the column was flushed with approximately 70 L n-hexane to remove non-vitamin E lipid fractions. The column was then eluted with a gradient solvent system composed of n-hexane and (0–12%) ethyl acetate. Approximately 10–15 L of solvent was used at each ethyl acetate increment. Fractions were collected at different time intervals and analyzed for content by thin layer chromatography (TLC), high performance liquid chromatography (HPLC), mass spectroscopy (MS), and proton nuclear magnetic resonance (<sup>1</sup>H NMR). Fractions rich in  $\gamma$ -T<sub>3</sub> were concentrated using a Heidolph Laborota 4000 efficient rotary evaporator (Elk Grove Village, IL) to give a yellow to orange viscous oil (~25 g  $\gamma$ -T<sub>3</sub>). The content and purity of the  $\gamma$ -T<sub>3</sub> in the product was confirmed by HPLC as previously described [38].

### 2.3. Synthesis of $\gamma$ -T<sub>3</sub> succinate

The procedure for the synthesis of  $\gamma$ -T<sub>3</sub> succinate was adapted from Lipshutz et al. [39]. Briefly, triethylamine (0.09 mL) was added to a solution of  $\gamma$ -T<sub>3</sub> (1.125 g) and succinic anhydride (0.4 g) in toluene (5.2 mL) and stirred at 85 °C. After 9 h water was added to the reaction mixture, which was then extracted with dichloromethane twice. The combined organic layers were washed with 1 N HCl (3 × 13 mL) and water (2 × 7.5 mL), then dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, and finally concentrated *in vacuo* to yield a yellow liquid. The viscous liquid was further purified by flash column chromatography on silica gel eluted with a 10% ethyl acetate/hexane to 35% ethyl acetate/hexanes gradient to afford  $\gamma$ -T<sub>3</sub> succinate (1 g) as pale, yellow viscous syrup.

### 2.4. Synthesis of $\gamma$ -T<sub>3</sub>PGS 350 and $\gamma$ -T<sub>3</sub>PGS 1000

Two  $\gamma$ -T<sub>3</sub> PEG variants with m-PEG molecular weights of approximately 350 and 1000 were synthesized via a straightforward two-step route outlined by the scheme in Fig. 1. m-PEG 1000 was selected to be comparable to the commercially available vitamin E TPGS, whereas the shorter m-PEG 350 was chosen to observe the effect of PEG molecular weight on the oral bioavailability and anticancer activity of  $\gamma$ -T<sub>3</sub> PEG. It was reported that the molecular weight of the PEG moiety can affect the availability and absorption of the conjugate molecules when taken orally [37]. A mixture containing  $\gamma$ -T<sub>3</sub> succinate (0.41 g), PEG 350 (0.28 g) or PEG 1000 (0.82 g) and p-TsOH (0.02 g) in toluene (3 mL) was refluxed for 8 h using a Dean–Stark trap. After cooling to room temperature the mixture was poured into a saturated aqueous NaHCO<sub>3</sub> solution and extracted with dichloromethane twice. The combined organic layers were washed with saturated NaHCO<sub>3</sub> (3 × 7 mL) and brine (2 × 4.5 mL), then dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, and finally concentrated *in vacuo* to afford  $\gamma$ -T<sub>3</sub>PGS 350 and  $\gamma$ -T<sub>3</sub>PGS 1000 conjugates (0.6 g) as brownish yellow syrups.

### 2.5. <sup>1</sup>H-NMR analysis of $\gamma$ -T<sub>3</sub>PGS 350 and $\gamma$ -T<sub>3</sub>PGS 1000

NMR studies were carried out to confirm the PEGylation of  $\gamma$ -T<sub>3</sub>. High-resolution <sup>1</sup>H NMR spectra of samples prepared in CDCl<sub>3</sub>

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