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## ORIGINAL ARTICLE

# Salicylic acid affects the expression of *VvCBF4* gene in grapes subjected to low temperature

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**Abstract** The present study investigates the effects of exogenous salicylic acid (SA) on the expression of *Vitis vinifera C-repeat binding factor 4 (VvCBF4)* gene under low-temperature conditions in an Iranian *Vitis vinifera* L. ‘Sultanina’. The experiment was conducted as a factorial experiment based on a completely randomized design with four replications. 100  $\mu\text{mol/L}$  SA (0, 1, 6 and 12 h before applying cold stress) in temperatures of  $1 \pm 0.5^\circ\text{C}$  (for 1, 3, 6 and 12 h) and  $22^\circ\text{C}$  (as control) were applied. The highest expression was observed in plants treated 6 h before sampling. By increasing the duration of low temperature, the expression of *VvCBF4* increased. Increasing the duration of cold stress to 6 h in  $1^\circ\text{C}$  increased the expression of *VvCBF4* to 24.3 fold. Exogenous application of SA and cold stress treatments increased the expression of *VvCBF4*. In conclusion, exogenous application of SA in cold stress, increased the expression of *VvCBF4* depending on treating time before cold stress. The highest *VvCBF4* expression was observed in plants treated 6 h before sampling and increasing the time decreased the expression. By increasing the expression of *VvCBF4* the tolerance of plant to cold stress increased.

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## 1. Introduction

Low temperature is one of the most important environmental stresses that limits the productivity and distribution of plants [22]. Various plant species can increase their freezing tolerance in response to low non-freezing temperatures; this phenomenon is defined as cold acclimation. Some molecular and physiological changes are involved in cold acclimation [27].

Although, the molecular basis of this acquired chilling acclimation is poorly understood, but the effect of some

transcription factors involved in response to low temperature is well established [17,23]. *C-Repeat Binding Factors (CBFs)* are transcription factors that have a vital role in gene regulation during cold acclimation in plant species [3,2,7]. Constitutive expression of either *CBF1* or *CBF3* transcriptional activators in transgenic Arabidopsis induced the expression of cold-regulated genes and also enhanced the freezing tolerance in non-acclimated plants [1,8,13]. In *CBF3*-expressing plants the proline and total soluble sugars had raised, also in cold-acclimated plants with overexpressed *CBF3*, freezing tolerance have been increased [8]. Furthermore, the ectopic expression of *CBFs* from other plant species can increase the freezing tolerance of transgenic Arabidopsis [26].

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Salicylic acid (SA) is an endogenous simple phenolic acid with hormonal function and omnipresent distribution among plants [24]. SA is involved in the regulation of major variety of metabolic and physiological processes in plants. Numerous studies have reported the valuable effects of SA treatments on cold tolerance of plants species such as tomato [4,19], wheat [20], banana [11], maize [18] and so on.

Grape is one of the most cultivated fruit crops that its importance is well known. Nevertheless, the cold stress always affects the growth, development, and productivity of this plant and limits its geographical distribution [6]. All in all, the present study was designed to investigate the effects of exogenous SA on expression of *VvCBF4* gene under low temperature conditions in an Iranian grape (Sultanina cultivar).

## 2. Materials and methods

### 2.1. Plant materials and study design

The experiment was conducted in a controlled-environment on two year old greenhouse-grown plants (*Vitis vinifera* L. 'Sultanina') under day and night temperature of 28–25 °C and 20–18 °C, respectively and maintained under a 16:8 light/dark cycle. The application of SA was through spraying of 100 µmol/L SA (0, 1, 6 and 12 h before applying cold stress) in temperatures of 1 ± 0.5 °C (for 1, 3, 6 and 12 h) and 22 °C (as control) in a factorial experiment based on completely randomized design in four replications.

### 2.2. RNA extraction and DNA synthesis

Total RNAs were extracted and purified from the leaves of grapes following the method described by Tattersall et al. [21]. Only the extractions having an  $A_{260}/A_{280}$  ratio of 1.8–2.0 and an  $A_{260}/A_{230}$  ratio >2.0 were applied for further analysis. The integrity of extracted RNAs was verified using 2% agarose gel electrophoresis followed by ethidium bromide staining. Oligo-dT, were used for first strand cDNA synthesis. The reaction mixture (Table 1) was prepared in a microtube on ice and was made up to 20 µl using RNase-free water.

### 2.3. Primer design and RT-qPCR analysis

The RNA sequences of *VvCBF4* gene were taken from NCBI ([www.ncbi.nlm.nih.gov](http://www.ncbi.nlm.nih.gov)) and its forward and reverse primers was designed by Oligo 7 (Table 2).

RT-qPCR analysis applied by an ABI StepOne Detection System (Applied Biosystems, USA), using the SYBR Green

**Table 1** Required amounts of reactives for cDNA synthesis.

Reactive	Volume
Vivantis RT Enzyme Mix I	0.5 µl
Buffer RT Enzyme	2 µl
Oligo dT Primer (50 µ M)	0.5 µl
Random 6 mers (100 µ M)	0.5 µl
dNTP	1 µl
DDW	11.5 µl
Total RNA (500 ng)	5 µl
Total	20 µl

**Table 2** Used primers for RT-PCR reaction.

Primer name	Sequence
<i>VvCBF4</i> F	5-ACCCTCACCGCTCGTATG-3
<i>VvCBF4</i> R	5-CCGCGTCTCCGAAACTT-3

**Table 3** The composition of reaction mixture for RT-PCR reaction.

Volume	Reactive
RT reaction solution (cDNA)	2 µl
Primer F	0.4 µl
Primer R	0.4 µl
Power SYBR Green PCR Master Mix	10 µl
DDW	7.2 µl
Total	20 µl

PCR Master Mix (TaKaRa, Toyoto, Japan). The reaction mixture (Table 3) was made up to 20 µl total volume per sample. An initial denaturation step at 95 °C for 10 s, followed by 40 cycles of 95 °C for 5 s and 60 °C for 60 s were performed (Fig. 1). Following amplification, a melting curve analysis was performed to guarantee the absence of primer dimers and other nonspecific products. Relative quantification was executed by the comparative CT ( $2^{-\Delta\Delta C_t}$ ) method [15]. To quantify the transcript level, a standard curve (copy number as a function of Ct) was created by a 10× mass dilution series of each cDNA fragment. The exact copy number was presented by extrapolation of the Ct value for each cDNA on the standard curve and determined as copy number ng<sup>-1</sup> of cDNA.

## 3. Results and discussion

Low temperature is one of the most important environmental stresses that hampers the manifestation of full genetic potential in plants. In recent studies, transcriptome analyses of cold response have shown that some transcription factors such as *CBFs* are involved in response to low temperature [26]. Also the effects of SA in ameliorating environmental stresses have been numerously reported [10].

Putting this all together, in the present investigation, we have decided to address the effects of exogenous SA on the expression of the *VvCBF4* gene under low temperature conditions in *Vitis vinifera* L. 'Sultanina'.

The exogenous SA, activated the *VvCBF4* gene in control plants (22 °C). The highest expression was observed in the plants treated 6 h before sampling. By increasing the time of SA treatment before sampling, a significant decrease in the expression of *VvCBF4* was observed (Fig. 2). Cold stress (1 °C) for 1 h along with SA also increased the expression of *VvCBF4*, however, had some decrease in comparison with control plants. The highest *VvCBF4* expression belonged to 1 h treating in 1 °C and application of SA 6 h before sampling (Fig. 3). It has been demonstrated that pretreatment with 0.1 mM SA would induce the chilling tolerance in potato plants [16]. Also in banana seedlings, 0.5 mM SA has been reported to induce the chilling tolerance both when sprayed

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