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Nuclear Medicine and Biology

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¹⁸F-labeled estradiol derivative for targeting estrogen receptor-expressing breast cancer[☆], ☆ ☆



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ARTICLE INFO

Article history: Received 31 October 2017 Received in revised form 25 December 2017 Accepted 22 January 2018 Available online xxxx

Keywords: Estrogen receptor Breast cancer PET imaging Biodistribution ¹⁸F-radiopharmceutical

ABSTRACT

Introduction: A novel radiotracer 1 (2 (2 (2 [18F]fluoroethoxy)ethoxy)ethyl) 1H 1,2,3 triazole estradiol ([18F] FETE) was successfully synthesized, characterized and evaluated in mice for estrogen receptor (ER)-positive breast cancer targeting with positron emission tomography (PET) imaging.

Methods: The tosylate precursor **3** was radiolabeled with 18 F and then reacted with 17 α ethinyl estradiol to produce the final [18 F]FETE. The physicochemical properties of [18 F]FETE were tested in vitro, including determination of the octanol/water partition coefficient, stability and cellular uptake in MCF-7 (ER-positive) and MDA-MB-231 (ER-negative) cells. An ex vivo biodistribution study was performed in normal Sprague Dawley rats, and in vivo microPET imaging was performed on MCF-7 and MDA-MB-231 tumor-bearing mice. The results of biodistribution and PET imaging of [18 F]FETE were compared with that of known 16 α [18 F] fuoro 17 β estradiol ([18 F]FES). Radiation dose estimates for [18 F]FETE were also analyzed.

Results: [18 F]FETE was obtained in high radiochemical yield ($^{46.59} \pm 8.06\%$) with high radiochemical purity ($^{>99\%}$) after HPLC purification and high molar activity ($^{15.45} \pm 3.15$ GBq/µmol). [18 F]FETE is a moderate lipophilic compound with good in vitro stability and the total synthesis time was 55 to 65 min. In biodistribution studies, [18 F]FETE showed high uptake in the ER-abundant uterine tissue of normal immature SD rats ($^{8.55} \pm 1.21$ and $^{6.83} \pm 1.70\%$ ID/g at 1 h after intravenous and intraperitoneal injection, respectively), and could be blocked with estradiol effectively (the uterus uptake was decreased to $^{0.63} \pm 0.35\%$ ID/g at 1 h after iv injection. MicroPET imaging of tumor-bearing mice with [18 F]FETE at 1 h after iv injection revealed considerable uptake in ER-positive MCF-7 tumors ($^{4.63} \pm 0.73\%$ ID/g) that could be inhibited ($^{1.47} \pm 0.29\%$ ID/g) and low uptake in ER-negative MDA-MB-231 tumors ($^{1.97} \pm 0.36\%$ ID/g). [18 F]FES has relatively low uptake in ER-positive tumor ($^{0.24} \pm 0.19\%$ ID/g) when compared with [18 F]FETE. The adult female effective radiation dose of [18 F]FETE in mice was estimated as $^{0.0022}$ mSv/MBq.

Conclusions: A novel 17α ethinyl estradiol-based ER probe [18 F]FETE was developed with high molar activity and good in vitro stability. Based on the results of bio-evaluation in normal immature rats and tumor-bearing mice, it might be a promising candidate for specific PET imaging of ER-positive breast cancer.

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1. Introduction

Breast cancer is a major health problem for women throughout the world. It has become the second cause of cancer deaths in females [1]. Diagnosis in the early stages is the key to the treatment of breast cancer [2]. According to previous work by Lippman et al. [3] approximately 70–80% of all breast carcinoma are estrogen receptor (ER) positive. Estrogen receptor-positive (ER+) breast tumors are more likely to respond to hormone therapy than ER-negative (ER-) tumors [4]. Knowing the receptor status can lead to more effective individualized treatment.

[†] Author Contributions: The manuscript was written through contributions of all authors. All authors have given approval to the final version of the manuscript.

^{☆☆} Contract/grant sponsor: This study was financially supported by the National Key Basic Research Program of China (2014CB744503), National Natural Science Foundation of China (81471707, 81360222, 21271030).

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With the development of positron emission tomography (PET) and single-photon emission computed tomography (SPECT), many estrogen derivatives and other non-hormonal medicines have been labeled with ¹⁸F or other radioisotopes, including ¹¹C, ⁷⁷Br, ^{99m}Tc, ¹²³I, and ¹²⁵I, making it possible to quantitatively image in a way that can monitor changes in tumor receptor levels [5].

Of all the ER-imaging agents, 16α [18 F]fluoro 17β estradiol ([18 F] FES) is the most successful. [18 F]FES has good affinity for the ER [6] and is currently in clinical trials as a PET-imaging agent for breast cancer [7]. However, [18 F]FES is metabolized mainly in the liver [6], and rapid blood clearance leads to a decrease in specific binding.

In recent years, with the purpose of improving the metabolic stability and enhancing the receptor-binding affinity of [$^{18}\text{F}]\text{FES}$ for ER imaging, a variety of FES derivatives have been synthesized and evaluated [8–11]. According to previous work, most sites on estradiol, including C-4, C-7, C-11, and C-17, can be substituted and still be tolerated by the ER [8]. Studies of 4 fluoro and 11 β methoxy substitutions (4FM [$^{18}\text{F}]\text{FES}$) achieved successful PET imaging of a breast cancer mouse model with better sensitivity than [$^{18}\text{F}]\text{FES}$ [12]. However, 4FM[$^{18}\text{F}]$ FES failed to improve on the metabolism of [$^{18}\text{F}]\text{FES}$ in vivo [12]. PET imaging studies of >20 FES derivatives have been published [8]. However, rapid blood clearance, resulting in low tumor uptake, is still a problem.

Therefore, we designed [18 F]FETE, which contains 17α ethinyl estradiol and polyethylene glycol (PEG) moieties. 17α Ethinyl estradiol, with an ethinyl group at C-17, has high affinity for the ER. Its relative binding affinity (RBA) is 190% when the binding affinity of estradiol is set at 100% [13 ,14]. PEG is a useful addition to lower the lipophilicity of the whole molecule. Theoretically, the addition of PEG could decrease liver metabolism and increase the probability of tumor uptake [10]. In this study, we synthesized and characterized this estradiol-based probe, labeled it with 18 F and evaluated its ER-targeting ability in vitro and in vivo.

2. Materials and methods

2.1. Chemistry and radiochemistry

All reagents and solvents were purchased from commercial sources. Reversed-phase high-pressure liquid chromatography (HPLC) was performed on a system with an UltiMate 3000 pump (DIONEX) and a B-FC-1000 flow counter (Bioscan). The C-18 reversed-phase semi-preparative HPLC column (250 \ast 10 mm, 5- μ m particle size [Hypersil GOLD-C18; Thermo Scientific]) was eluted at a flow rate of 4 mL/min.

The syntheses of the [18 F]FETE precursor and its nonradioactive reference compound ([19 F]FETE) are available in the supplemental data. 1 H NMR spectra were recorded on a Bruker (400 MHz) spectrometer. Mass spectra were obtained with a Bruker Esquire 3000 plus ESI-MS. Chemical shifts are reported in δ (ppm) values.

Fluorine-18 was obtained from the PET Center of the First Affiliated Hospital of Xiamen University. The labeling procedure for [18F]FETE is

shown in Scheme 1. [¹⁸F]Fluoride was isolated from enriched water by trapping on an anion exchange resin (20 mg) in its carbonate form and then eluting with a CH₃CN-H₂O solution (1 mL, 9:1) of potassium carbonate (3 mg) and Kryptofix [2] (14 mg). This solution was dried by azeotropic distillation with acetonitrile (0.5 mL) three times under a stream of nitrogen at 110 °C, and compound **3** (in 0.2 mL of acetonitrile) was then added. The reaction mixture was shaken at 90 °C for 20 min. After cooling the reaction mixture to room temperature, deionized water (4 mL) was added, and the mixture was passed through a Sep-Pak Plus C18 cartridge, which was then washed with deionized water (4 mL), followed by elution of the [¹⁸F]AE intermediate ([¹⁸F]-4b in Scheme 1) from the cartridge with 1 mL of tetrahydrofuran (THF). The purity was confirmed by HPLC analysis.

Next, 17α ethinyl estradiol (2 mg), DIPEA (2 mg) and CuI (1 mg) were added to [18 F]AE, and the reaction mixture was vibrated at 60 °C for 20 min. Then, the mixture was passed through a syringe filter (0.2 µm). The final product of [18 F]FETE was purified by semi-preparative HPLC with a constant gradient (0–20 min) of 35% ACN in pure water (4 mL/min).

[¹⁸F]FES was prepared according to the published literature for comparison [9]. The detailed synthetic method is presented in the supplementary information.

2.2. Octanol/water partition coefficient

The octanol-to-water partition coefficient of [18 F]FETE was measured according to a procedure published previously [15]. Briefly, 1 mL of 1 octanol and 1 mL of saline were added to a centrifuge tube containing 370 kBq of [18 F]FETE. The radiotracer was fully mixed with vigorous vortexing for 1 min, and the tube was centrifuged at 5000 rpm for 5 min. Identical volumes of both the organic and inorganic layers were removed and measured in triplicate using a 2480 Wizard [2] Automatic 2 -counter (PerkinElmer). Then, the partition coefficient, which is expressed as log P. was calculated.

2.3. In vitro stability of [18F]FETE

The in vitro stability was measured according to a previously published method [16]. Briefly, the radiolabeled compound was incubated in physiological saline at room temperature for 1, 2, and 4 h. To determine its serum stability, the radiotracer was incubated in mice serum at 37 °C for 4 h. To remove plasma proteins from the serum, 1 mL of acetonitrile was added, and the sample was centrifuged. Finally, the radiochemical purities of the radiotracer were analyzed by radio-HPLC using the procedure described above.

2.4. In vivo metabolic stability of [18F]FETE

The metabolic stability studies of [¹⁸F]FETE were performed according to published procedures [17,18]. Balb/c mice were intravenously injected with 3.7 MBq of [¹⁸F]FETE. The mice were sacrificed at 1 min,

HOOOON TSCI Et₃N TSO OON TS NAN3 TSO OON N3

1 TSO OON N3

Aa: KF/Kryptofix ACN 90
$$^{\circ}$$
C ACN 90 $^{\circ}$ C ACN 90 $^{\circ}$ C

17 $^{\circ}$ -Ethynylestradiol Cul, DIPEA, THF

5a: R=F 5b: R= 18 F| 18 F|FETE

4a: R=F 4b: R= 18 F| 18 F|AE

Scheme 1. Synthetic routes for [18/19F]FETE.

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