



## Short communication

Immune function against bacteria of chitin deacetylase 1 (*EcCDA1*) from *Exopalaemon carinicauda*Yuying Sun<sup>a</sup>, Jiquan Zhang<sup>a,b,c,\*</sup>, Jianhai Xiang<sup>b,c</sup><sup>a</sup> College of Life Sciences, Hebei University, Baoding, Hebei 071002, China<sup>b</sup> Laboratory for Marine Biology and Biotechnology, Qingdao National Laboratory for Marine Science and Technology, Qingdao 266000, China<sup>c</sup> Key Laboratory of Experimental Marine Biology, Institute of Oceanology, Chinese Academy of Sciences, Qingdao 266071, China

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## ABSTRACT

Chitin deacetylase (CDA, EC 3.5.1.41), belonging to a family of extracellular chitin-modifying enzymes, can catalyze the deacetylation of chitin. In this study, the full-length cDNA sequence encoding chitin deacetylase 1 (*EcCDA1*) was obtained from *Exopalaemon carinicauda*. The complete nucleotide sequence of *EcCDA1* contained a 1611 bp open reading frame (ORF) encoding *EcCDA1* precursor of 536 amino acids. The domain architecture of the deduced *EcCDA1* protein contained a signal peptide, a chitin-binding peritrophin-A domain (ChtBD2), a low-density lipoprotein receptor class A domain (LDLa) and a Polysacc\_deac\_1 domain. *EcCDA1* mRNA was predominantly expressed in the gills. The expression of *EcCDA1* in the prawns challenged with *Vibrio parahaemolyticus* and *Aeromonas hydrophila* changed in a time-dependent manner. The expression of *EcCDA1* in the prawns challenged with *V. parahaemolyticus* was up-regulated at 12 h ( $p < 0.05$ ), and significantly up-regulated at 24 h and 48 h ( $p < 0.01$ ), and then returned to the control levels at 96 h post-challenge ( $p > 0.05$ ). At the same time, the expression in *Aeromonas*-challenged group was significantly up-regulated at 12, 24 and 48 h ( $p < 0.01$ ) and returned to the control levels at 120 h post-challenge ( $p > 0.05$ ). Then, *EcCDA1* was recombinantly expressed in *Pichia pastoris* and the purified recombinant *EcCDA1* could not inhibit the growth of *V. parahaemolyticus* or *A. hydrophila*, which indicated that the CDA1 may play its biological activity in immune defense by deacetylation from chitin.

## 1. Introduction

Chitin, one of the most important biopolymers in nature, is mainly produced by fungi, arthropods and nematodes. In arthropods, their cuticles can form an exoskeleton to keep pace with body growth due to the presence of chitin and sclerotized proteins [1]. In addition, their growth and morphogenesis are strictly dependent on the capability to remodel chitin-containing structures [1]. Chitin-related enzymes play fundamental roles in chitin metabolism and they can be divided into three main categories, based on their functions to synthesize chitin (chitin synthases), to enzymatically alter chitin by deacetylation (chitin deacetylase, CDA) and to degrade chitin by hydrolytic process (chitinases and N-acetylglucosaminidases) [2]. CDAs (EC 3.5.1.41) are secreted proteins belonging to a family of extracellular chitin-modifying enzymes and they can hydrolyze the acetamido group in the N-acetylglucosamine units of chitin and chitosan [3]. CDA was first discovered from extracts of *Mucor rouxii* and it could convert nascent chitin into chitosan [4,5].

At present, a lot of CDA genes have been obtained in the species of

Arthropod, especially in insects [3,6–10]. In insects, CDAs can convert chitin into chitosan, the N-deacetylated form of chitin, which influenced the mechanical and permeability properties of structures such as the cuticle and peritrophic matrices [11]. At present, a family of genes encoding chitin deacetylase (CDA)-like proteins in insects had been identified in the annotated genome sequences and the number of CDA genes was five to nine depending on the species [7]. All of the insect CDAs could be clustered into five major groups [6]. In *Helicoverpa armigera*, it is reported that the downregulation of a midgut-specific CDA-like protein as a possible mechanism to reduce susceptibility to baculovirus by decreasing peritrophic membrane (PM) permeability [12]. However, there was only one CDA gene reported in crustaceans, that is, CDA cDNA (named *PmCDA1*) cloned from the gills of black tiger shrimp, *Penaeus monodon* [13]. *PmCDA1* was reported to be distinctly highly expressed in the gills of shrimp and the authors thought that gills in shrimps served as the predominant site for the formation of hemocyte nodules during injection of foreign particles and accumulation of viable bacteria during infection, suggesting its significant role in shrimp defense [13]. As we know, there is no model animal in Crustacean to be

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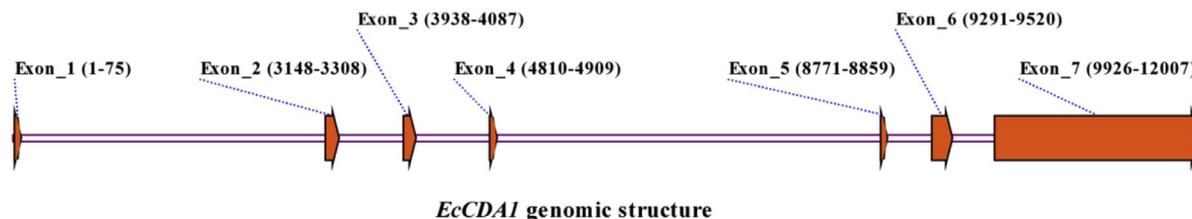
**Table 1**  
Primers mentioned in the paper.

Primers	Sequences (5'-3')	Sequence information
RT-EcCDA1F	AGAAGAAGAGGGTTCGTCGAAGC	Real-time PCR
RT-EcCDA1R	GACTCATCGGCACAGTCAAAT	Real-time PCR
18S-F	TATACGCTAGTGGAGCTGGAA	Real-time PCR
18S-R	GGGGAGGTAGTGACGAAAAAT	Real-time PCR
9k-EcCDA1F	GCGAAATTCATCATCACCATCACCCAGAAATAGTGAAACGCCAGCGGCCAC	Construct the expression vector, introducing a restriction enzyme site for <i>EcoR</i> I and a 6 × His-tag
9k-EcCDA1R	GCGCGGCCGCTTAGAAGTAACCCCTCTCCAAAGGATC	Construct the expression vector, introducing a restriction enzyme site for <i>Not</i> I
5'AOX1	GACTGGTTCCAATTGACAAGC	Confirm the insert target gene
3'AOX1	GCAATGGCATTCTGACATCC	Confirm the insert target gene

Note: F and R stand for forward primers and reverse ones, respectively.

1 GTGAACGGTTGGTTCGGAGAGTCAACATGACAAGTATACAGATTGCGGCGATAGCCCTTGCCCTCTTGGCATCGCATCCGGAGAAATAGTGAAACGCCAGGGCCACCCTAAGCGAAC 120  
M T S I R V A A I A L A L L G I A S G E I V K R Q A A T V S E  
121 CCACTGAAGATGAGGCCGATGCCTTCACCAAGAGTTGTGTCGAGACAAGGGCGCAGGCGAATGGTTCAGGCTCGATCTGAATGATTGTCGTGATGTCATCCAGTGCACAGGGCAGGGC 240  
P T E D E A D A F T K E L C R D K G A G E W F R L D L N D C R D V I Q C T E A G  
241 TTCAGGCGCTGAGATGTCCTCACGGTCTGGCTTTCAACCTGGAATTGCAGACCTGTGACTGGAAGGACAACGTCAGAAGTGCACAGGAGAGAAGAGGGTCTGCAAGCCCTGCG 360  
L Q A L R C P H G L A F N L E L Q T T C D W K D N V K N C N Q K E K R V V K P L  
361 TCACACCCGTCGAGCCTCTTTGTCAGGAGAACCTGTGCGCTGCGCGATGGAACGTGCATTGTCGAAACCCCTGTTTTCGCGATGGCAAATTTGACTGTGCCGATGAGTCCGACGAAAAACA 480  
L N T V E P L C Q E N L L A C G D G T C I D R T L F C D G K F D C A D E S D E N  
481 CCTGTGATATCAAGAGTATCCCAACAGCGCACCCATTGCAACCCGGATGAGTGCAGCCTCCCGGCTGTTACTGTCTTAATAACGCCAACGAGGTCGCCGACACATCAACCTACCA 600  
T C D I K S D P N S A P I C N P D E C R L P D C Y C F N N A N E V P D A N M (N) P T  
601 ATGTACCCCAAATGATCACCATAACATTGACGATGCTGTCAACATTGAAAACATCGACCTTTACAATATCATTTTCGATAGCCGCTTCAACCCTAACAGTGTCCATCAAGTGCAGCT 720  
N V P Q M I T I T F D D A V N I E N I D L Y N I I F D S R F N P N Q C S I K S T  
721 TCTTGTCTCCACAAATACAAAATCTACTCCGCTGTGACGAGATTGTCATCGCCTTGCTCAGCAAAATGGCATCCACTCAATCAGCCACAGCAAAATGAAACATTTGCGCAAGAGCTT 840  
F F V S H K Y T (N) Y S A V Q D L H R L G H E I A I H S I S H S N N E T F W T K A  
841 CCCCAGTGAATGGGACGTGAGATGGCAGGTGGTGTGATTGTGAAAGATTGGCAACATCACCGACTCTTCTGTGATTGGTGTGAGATCTCCCTACCTAGTGTGGTGGAAACA 960  
S P D E W E R E M A G G R V I V E R F A (N) I T D S S V I G V R S P Y L R V G G N  
961 ACCAATTCGGCATGAGGACGAGATGCCTTCTCTACGACTCCACCATGACTGCCCACTGCAGAACCCCACTTGGCCTTACACCCCTTTATTACCGCATGCCACAGCTTGGCCAG 1080  
N Q F G M M E Q N A F L Y D S T M T A T G Q A N C P P L W P Y T L Y Y R M P H A C H  
1081 GCAACCTCCGAATGGCCACCCTTCTTCTCGCGTCTGGGAAATGGTCATGAACGAGATGGACCGTCTGTGAGGAACCAACCATTGAAGAAGATTGGCTGGATGTGATGGTGGATT 1200  
G N L Q N C P T R S F A V W E M V M N E M D R R E E P T I E E D L P G C A M V D  
1201 CTTGCTTCCCAACAGCCAGCTGATCAGTTCTATAAAATCTCTGTAACAACTTTGACCGTCATACCTACCAACCCGTCGCCCAATGGGTCTACTTCCATTTCTGCTTCTCTCA 1320  
S C F S N K P T A D Q F Y K F L V N N F D R H Y L T N R A P M G L Y F H S A F L  
1321 AAAATGATCCAGAAATATGGACGCTTCTCTTCTGCTGGATGAGACCTTGGCAAAACCCGATGTTACTTCTGTCACCATGACTCAAGTCAATCAATGGATGCAGGACCCACAGC 1440  
K N D P E I L D A F L F W L D E T L A N N P D V Y F V T M T Q V I Q W M Q D P Q  
1441 CCGTCAGCAACCTCAAGAACTACGAGCCCTGGAAGGAGAAGTGAACGTCAGTGGACCTCCTTCTGCTACGGTGGCACCACCACTGTAACCTCAACTGATGAACCTCCCGGTGAGACCC 1560  
P V S N L K N Y E P W K E K C N V A G P P F C Y G G T N C E L N T D E L P G E T  
1561 TTCGCTGGCCACCTGCATGCGATGCCAACAGGTATCCCTGGTTATGGATCCTTTGGGAGAGGGTACTTCTAAGAGCACTGCTGCTGCATTAATGCTGTGCCCTGTCCCTGGTT 1680  
L R L A T C M R C P N R Y P W L L D P L G E G Y F \*  
1681 GAGTCAGCCCTCCTCCTCCCGTGGGACAGTTGTCTCATCTGTAAATATATTGTTTATAAATTTGTACATGATCGTGTCCAGGCATGTGATGCCCTATTCTTAACAGTGGTT 1800  
GGCTGGTCAGGGTTCGGCGGCAACACCACCATCATCTGTCGCTAAGGCCAAGAAATCAGTTTGTCTACAATGAACCTGTGCAACGGTCTCCAGAGTGTGCTGCTGTGCTGATG 1920  
1921 CTGCTGCCTTCTGTCAGCCTCATGACAGTCCGAGCTCATATTGCAAACTATTCCAAATCCCTTGTAAACCCCTTAAATCACTGTGCGGGATGAGCTGGGGCTGCTCTTCCAAA 2040  
2041 CTTGCGGATCCCAAACTTCCCTTCAACTCTATCTCATCTATGGGAAATACCGCGGGTCCCAATTAATATTATTAGACCTGGAGGGCCCTCATCACCCTTTTCTTAATAC 2160  
2161 CTCACCCCTTAAAAATACAAAAACATCACCAAAACAGGTGGGGCCATGTGACAGTGCCTACCACCTGAGCACAATTTTGGATTAACTAGCTGATGATCGTTTAGA 2280  
2281 ATTTTAGCGTCACTTGTTTTACCTTCCATATTATGTTTAAATGTTACCCAAATAAAAAACATTATTCTCAATATTCTCATTATCCTTAGAATAATATACCTTACATAAAGAAA 2400  
2401 AATATAGTCCCTGTGTAAGAAAATATCTTTTCAATACCTACCTCAACTATTACACAGAACTATACCAACGAACTCTCAATCTTGAATATAAAGGCAAAAAACAGACG 2520  
2521 TACAGCATGAGAACCCAGTACAGTATTATACATAATGATGAATATTATTAACAATAGTACAACTTCAAAACTATTAGATAAGATCGATACACAGCTTACATAA 2640  
2641 TAGTACAAAACTGACCAAGAAGATATGAATCCAGAAATGCAATGACAGAAAAAACTCCAGCAACTCTTACATGAAAAACCTTGAACAATTTTGGACACATCCATAAT 2760  
2761 TAACTATCAAGAAATCTACTGGTTCCATAACCCATTCCTTGAAGATATGATGTTCTAGACCTAACTATAAATTTCTAATATGCATGCTAATACATTAGAAGCACGGGTCTATATACG 2880  
2881 CTTAAG 2887

(A)



(B)

**Fig. 1.** (A) The nucleotide sequence and deduced amino acid sequence of EcCDA1. The predicted signal peptide is underlined in red. Chitin-binding peritrophin-A domain (ChtBD2) is underlined in black, low-density lipoprotein receptor class A domain (LDLa) is underlined in pink, and the Polysacc\_deac\_1 domain is underlined in blue. The N-glycosylation sites are circled in red. (B) The genomic structure of EcCDA1. (For interpretation of the references to colour in this figure legend, the reader is referred to the Web version of this article.)

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