

Contents lists available at ScienceDirect

Saudi Pharmaceutical Journal

journal homepage: www.sciencedirect.com



Original article

Bioactive epoxides and hydroperoxides derived from naturally monoterpene geranyl acetate



Suzan A. Khayyat a,*, Manal Y. Sameeh b

- ^a Chemistry Department, Faculty of Science, King Abdulaziz University, P.O Box 55075-21534, Jeddah, Saudi Arabia
- ^b Chemistry Department, Al-Leith College- Um Al-Qura University, Saudi Arabia

ARTICLE INFO

Article history: Received 7 August 2017 Accepted 12 November 2017 Available online 24 November 2017

Keywords:
Medicinal plants
Geranyl acetate
Hydroperoxide, epoxide
Photooxygenation, antifungal activity

ABSTRACT

Geranyl acetate **(1)** was oxidized thermally and photochemically using (mcpba, H_2O_2) respectively to obtain (E)-5-(3, 3-dimethyloxiran-2-yl)-3-methylpent-2-enyl acetate **(2)** and 3-(2-(3, 3-dimethyloxiran-2-yl) ethyl)-3-methyloxiran-2-yl) methyl acetate **(3)**. On the other hand, photooxygenation of **1** with tetraphenyl porphin (TPP) as a photo sensitizer gave corresponding acitic acid 2,6-bis-hy droperoxy-7-methyl-3-methylene-oct-7-enyl-ester **(4)**, acitic acid 7-hydroperoxy-3,7-dimethyl-octa-2, 5-dienyl ester **(5)** and Acitic acid 3-hydroperoxy-7-methyl-3,7-dimethyl-octa-1,6-dienyl ester **(6)**. Antifungal studies were carried out on geranyl acetate and its derivatives. Studies on the antifungal activity especially Microsporum gypsum, Trichophyton vercossum and Candida tropicalis showed that geranyl acetate, its epoxide and hydroperoxide derivatives have good antifungal action.

© 2017 The Authors. Production and hosting by Elsevier B.V. on behalf of King Saud University. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

1. Introduction

In the last years, there has been a rise in the usage of aromatic medicinal plants and their essential oils in technical research and industrial applications including nutritious therapeutic, and cosmetic uses (Kebede & Hayelom 2008; Kejlová et al., 2010; Nisaet et al., 2011; Ekor 2013; Ramadan 2015; Emmanuel et al., 2016). The therapeutic properties of certain medicinal plants generally related to their content of secondary metabolites, such as polyphenols, terpenes, phytosteroids, and alkaloids, which produced in considerable amounts and variable proportions (Emam et al., 2009). The essential oils of the plants are the principle of their fragrance. They are called also ethereal oils, or volatile oils because they vaporize rapidly when exposed to the air at ordinary temperatures. In general, the essential oils consist of many mixtures including different sorts of molecules. These chemical constituents divided into two broad classes: terpenes and phenylpropanoids. Nevertheless, most essential oils consist mainly of monoterpenes,

E-mail address: saekhayyat@kau.edu.sa (S.A. Khayyat). Peer review under responsibility of King Saud University.



Production and hosting by Elsevier

which are the main chemical constituents of the essential oils of these plants that found as mixtures of odoriferous components and can be obtained by steam distillation or solvent extraction from a large variety of aromatic plants. They are found in edible as well as in medicinal plants with a therapeutic properties (Dudai et al., 2005; Sousa et al., 2006; Quintans-Junior et al., 2008; Nerio et al., 2010; Reinaldo et al., 2011).

On the other hand, the chemistry of singlet molecular oxygen (${}^{1}O_{2}$), (which prepared through photooxidation reaction) has amazing consideration by chemists because of its environmental and biomedical importance beside its interesting mechanistic and synthetic aspects (Stratakis and Orfanopoulos, 2000; Khayyat 2011) Moreover, unsaturated terpenes are adept of trapping activated oxygen species *in vivo* to give intermediate epoxides which can alkylate DNAs, proteins, and other biological species (Richter, 2003; Elgendy and Khayyat, 2008; Khayyat and Saddiq, 2015). Geranyl acetate (1) is a monoterpene extant in the volatile oils of many plant species, such as Cypress, Origanum, and Eucalyptus oils (Aligiannis et al., 2001; Delaguis et al., 2002) (see Fig. 1).

Taking into account significant therapeutic value of monoterpene especially geranyl acetate and the important applications of its epoxides and hydroperoxide derivatives, in this study, we investigated some oxidation reaction of geranyl acetate and the biological activities of its products

^{*} Corresponding author.

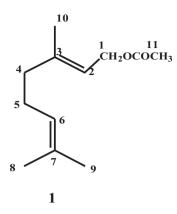


Fig. 1. Structure of geranyl acetate.

2. Materials and methods

2.1. Chemistry

¹H-NMR spectra were obtained in CDCl₃ solution with a Brucker AVANCE D.P.X. 600 MHz apparatus. A sodium lamp (Phillips G/5812 SON) was used for photo-irradiation reactions. Thin layer chromatography (TLC) and preparative layer chromatography (PLC): Polygram SIL G/W 254, Mecherey-Nagel.

2.2. General epoxidation procedures of geranyl acetate (1)

2.2.1. Method A: Photochemical epoxidation using hydrogen peroxide A solutions of H_2O_2 (2.5 mL, 50%) was added cautiously drop wise over 5 min to a stirred solution of 1 (5 mmol) in C_2H_5OH (25 mL) at 0 °C. The mixtures were irradiated using sodium lamp in an atmosphere of nitrogen. The reaction mixture evaporated and purified through column chromatography by petroleum ether 60–80 °C and ether (8:2) as elution (Elgendy and Khayyat, 2008) (Table 1).

2.2.2. Method B: Thermal epoxidation using m-chloroperbenzoic acid Solution of 1 (1.0 mmol) in CHCl₃ (25 mL) was cool at 0 °C. mcpba (5.0 mmol, 80%) was added portion wise to the reaction mixtures, then stirred at room temperature (TLC, peroxide test by KI, 10%). The reaction mixture was washed with a saturated aqueous solution of NaHCO₃ (3 × 10 mL), then with distilled water (3 × 10 mL). The organic layer was separated, dried, evaporated and purified by column chromatography using petroleum ether 60–80 °C and ether (8:2) gave the epoxide derivatives as viscous oils (Elgendy and Khayyat, 2008) (Table 1).

2.3. Photooxygenation reaction of geranyl acetate

One gram of 1 was take place in test tube, mixed with CHCl $_3$ and TPP as a sensitizer, then exposed to sodium lamp at $-20\,^{\circ}$ C. stream of dry oxygen gas was passed into the mixture of reaction throughout the irradiation. The solvent was evaporated at 20 °C. Then the crude product was purified by column chromatography on silica gel as adsorbent and a mixture of petroleum ether and ethyl acetate as elution to give the hydroperoxides products 4, 5 and 6,

Table 1Thermal and photo epoxidation of Geranyl acetate.

Comp. No.	Epoxidation	Solvent	Yield	Epoxid. Prod.
1	Thermally (mcpba)	CHCl ₃	0.80%	2: 60%
1	Photochemically (H ₂ O ₂)	C ₂ H ₅ OH	0.55%	3: 20% 3: 55%

which were successfully separated in pure form in the yields (Elgendy and Khayyat, 2008) (Table 2).

2.4. Spectroscopic data

2.4.1. Acetic acid 3,7-dimethyl-octa-2,6-dienyl ester (1)

Colorless oil, $C_{12}H_{20}O_2$ (M 196.29). IR (thin film): v^- : 1021, 1227, 1365, 1738.5 (COO), 2914.9 (CH str.) cm⁻¹. ¹H NMR (CDCl₃): δ :1.72(s, 3H, 10 CH₃), 1.8 (s, 6H, $^{8.9}$ CH₃), 2.0 (s, 3H, CH₃, H-11), 2.5 (Comp. pat., 4H, H-4, 5), 4.6 (d, 2H, J = 8 Hz, H-1), 5.06 (dd, 1H, J = 11 Hz, H-6), 5.33 (dd, 1H, J-11 Hz, H-2). 13 C NMR spectrum, δ ppm: 15.8 (10 C), 17.4 ($^{8.9}$ C), 20.3 (OCH₃), 25.4 (9 C), 25.9 (5 C), 39.1 (4 C), 60.7 (1 C), 118.0 (2 C), 123.3 (6 C), 131.1 (7 C), 141.4 (3 C), 170.3 (CO).

2.4.2. (E)-5-(3,3-dimethyloxiran-2-yl)-3-methylpent-2-enyl acetate (2)

Colorless oil, $C_{12}H_{20}O_3$ (M 212.29). ¹H NMR (CDCl₃): δ :1.27 (s, 3H, CH₃, H-8), 1.30(s,3H, CH₃, H-9), 1.73 (s, 3H, CH₃, H-10), 2.2 (Comp. pat., 4 Hz, H-4,5), 2.61 (dd, 1H, J = 8 Hz, H-6), 4.5 (d, 2H, J = 8 Hz, H-1), 5.30 (dd, 1H, J-11 Hz, H-2). ¹³C NMR spectrum, δ ppm: 16.1 (10 C), 20.7 (COCH₃), 24.2 (8 ,9C), 35.2 (4 C), 26.1(5 C), 42.2 (6 C), 61.2 (7 C), 118.6 (2 C), 140.5 (3 C), 170.2 (CO).

2.4.3. 3-(2-(3,3-dimethyloxiran-2-yl)ethyl)-3-methyloxiran-2-yl) methyl acetate (3)

Colorless liquid, $C_{12}H_{20}O_4$ (M 228.29). 1H NMR (CDCl₃): δ :1.22 (s, 3H, 9CH_3), 1.32 (s, 3H, 8CH_3), 1.34 (s, 3H, $^{10}CH_3$), 1.81 (Comp. pat. 4H, H-4,5), 2.10 (s, 3H, COCH₃), 2.61 (dd, 1H, J = 8 Hz, H-6), 2.71 (Comp. pat, 1H, H-2), 4.08 (Comp. pat., 1H, H-1), 4.30 (Comp. pat., 1H, H-1). ^{13}C NMR spectrum, δ ppm: 16.56 (^{10}C), 20.56 (CH₃-CO), 24.1 ($^{8.9}C$), 24.4 (^{5}C), 35.1 (^{4}C),), 58.1 (^{6}C), 58.9 (^{2}C), 59.98 (^{1}C), 63.01 (^{7}C), 76.9 (^{3}C), 170.5 (CO).

2.4.4. Acetic acid 2,6-bis-hydroperoxy-7-methyl-3-methylene-oct-7-envl-ester (4)

Colorless liquid, $C_{12}H_{20}O_6$ (M 260.28). ¹H NMR (CDCl₃): δ :1.72 (bs, 2H, H-5), 2.06 (bs, 2H, H-4), 2.19 (s, 3H, COCH₃), 4.26 (bs, 2H, H-1,6), 4.57 (bs, 2H, H-1,2), 4.97 (bs, 1H, H-8), 5.07 (bs, 1H, H-10), 5.33 (bs, 1H, H-8), 5.66 (bs, 1H, H-10), 8.95 (s, 1H, OOH), 9.46 (s, 1H, OOH). ¹³C NMR spectrum, δ ppm : 17.4(9 C), 20.7 (COCH₃), 24.2 (4 C), 26.6 (5 C), 61.2 (1 C), 81.5 (2 C), 88.4 (6 C), 114.0 (1 C), 118.2 (8 C), 142.0 (7 C), 143.8 (3 C), 172.4 (CO).

2.4.5. Acetic acid 7-hydroperoxy-3,7-dimethyl-octa-2,5- dienyl ester (5)

Colorless liquid, $C_{12}H_{20}O_4$ (M 228.14). 1H NMR ($CDCl_3$): δ :1.3 (s, 6H, $^{8.9}CH_3$), 1.7 (s, 3H, $^{10}CH_3$), 2.1 (s, 3H, 11CH₃), 2.86 (Comp. Pat., 2H, H-4), 4.5 (bs, 1H, H-1), 5.53 (dd, 1H, H-6), 6.1 (Comp. Pat., 1H, H-5), 7.3 (s, 1H, OOH). ^{13}C NMR spectrum, δ ppm : 17.3 (^{10}C), 24.2 ($^{8.9}C$), 45.0 (^{4}C), 60.4 (^{1}C), 82.0 (^{7}C), 125.5 (^{2}C), 128.5 (^{5}C), 136.2 (^{6}C), 142.5 (^{3}C), 174.9 (CO).

2.4.6. Acetic acid 3-hydroperoxy-7-methyl-3,7-dimethyl-octa-1,6-dienyl ester (6)

Colorless liquid, $C_{12}H_{20}O_4$ (M 228.14). 1H NMR (CDCl₃): δ :1.27 (s, 5H, H-4,10), 1.5–1.9 (Comp. pat., 8H, H-5,8,9), 2.21 (s, 3H,

Table 2 Photooxidation reaction of Geranyl acetate.

Comp No.	Start Wt. gm	Sensetizer	Solvent	Irradiation time	Yield	Photo products:
1	1	TPP	CHCl₃	12	0.68%	4: 20% 5: 33% 6: 15%

Download English Version:

https://daneshyari.com/en/article/8522560

Download Persian Version:

https://daneshyari.com/article/8522560

<u>Daneshyari.com</u>