

## Short communication

Proinflammatory cytokine interferon- $\gamma$  increases the expression of BANCR, a long non-coding RNA, in retinal pigment epithelial cellsR. Krishnan Kutty<sup>a,\*</sup>, William Samuel<sup>a</sup>, Todd Duncan<sup>a</sup>, Olga Postnikova<sup>a</sup>, Cynthia Jaworski<sup>a</sup>, Chandrasekharam N. Nagineni<sup>b</sup>, T. Michael Redmond<sup>a</sup><sup>a</sup> Laboratory of Retinal Cell and Molecular Biology, National Eye Institute, National Institutes of Health, Bethesda, MD 20892, United States<sup>b</sup> Radiation Biology Branch, Center for Cancer Research, National Cancer Institute, National Institutes of Health, Bethesda, MD 20892, United States

## ARTICLE INFO

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## ABSTRACT

The inflammatory response may contribute to retinal pigment epithelial (RPE) dysfunction associated with the pathogenesis of age-related macular degeneration (AMD). We investigated whether the inflammatory response affects the expression of long coding RNAs (lncRNAs) in human RPE-derived ARPE-19 cells. This class of regulatory RNA molecules recently came to prominence due to their involvement in many pathophysiological processes. A proinflammatory cytokine mixture consisting of IFN- $\gamma$ , IL-1 $\beta$  and TNF- $\alpha$  altered the expression several lncRNAs including BANCR in these cells. The cytokine responsible for increasing BANCR expression in ARPE-19 cells was found to be IFN- $\gamma$ . BANCR expression induced by IFN- $\gamma$  was suppressed when STAT1 phosphorylation was blocked by JAK inhibitor 1. Thus, proinflammatory cytokines could modulate the expression of lncRNAs in RPE cells and IFN- $\gamma$  could upregulate the expression of BANCR by activating JAK-STAT1 signaling pathway.

## 1. Introduction

Recent studies have shown that long non-coding RNAs (lncRNAs), non-protein coding transcripts with a minimum length of 200 nucleotides, may mediate many pathophysiological processes including inflammatory response [1,2]. BRAF-activated non-coding RNA (BANCR, LINC00586), a 693-nucleotide long lncRNA transcribed from human chromosome 9, has been shown to regulate proliferation and migration of melanoma and other cancer cells [3–5]. This lncRNA is also reported to regulate epithelial-mesenchymal transition (EMT) [4,5]. Furthermore, BANCR is highly expressed in retinoblastoma tissues and cells, and silencing its expression resulted in a decrease in retinoblastoma cell proliferation, migration and invasion [6].

Retinal pigment epithelium (RPE), a polarized monolayer of pigmented cells located adjacent to photoreceptor cells, is indispensable for visual function since it provides nutrients to the photoreceptor cells, regenerates the visual chromophore 11-*cis*-retinal and phagocytoses rod outer segment discs generated by circadian shedding [7]. Dysfunction of RPE resulting from abnormal inflammatory response may play a role in the pathology of age-related macular degeneration (AMD) [8,9]. Proinflammatory cytokines secreted by infiltrating macrophages and lymphocytes may trigger retinal pigment epithelial (RPE) cell

dysfunction. We have reported earlier that human RPE derived ARPE-19 cells respond to proinflammatory cytokines IFN- $\gamma$ , TNF- $\alpha$  and IL-1 $\beta$  by increasing the expression of cytokines and chemokines [10]. The proinflammatory cytokines also decreased the expression of RPE characteristic genes and induced epithelial-mesenchymal transition (EMT)-like changes in these cells [11]. It is not yet known whether lncRNAs play a role in mediating the deleterious effect of IFN- $\gamma$ , TNF- $\alpha$  and IL-1 $\beta$  on RPE cells. Therefore, we investigated whether these proinflammatory cytokines can affect the expression of lncRNAs like BANCR in the ARPE-19 cells. We observed that the inflammatory response altered the expression of several lncRNAs in these cells and that the activation of JAK-STAT1 signaling pathway by IFN- $\gamma$  increased the expression of BANCR.

## 2. Materials and methods

## 2.1. Cell culture

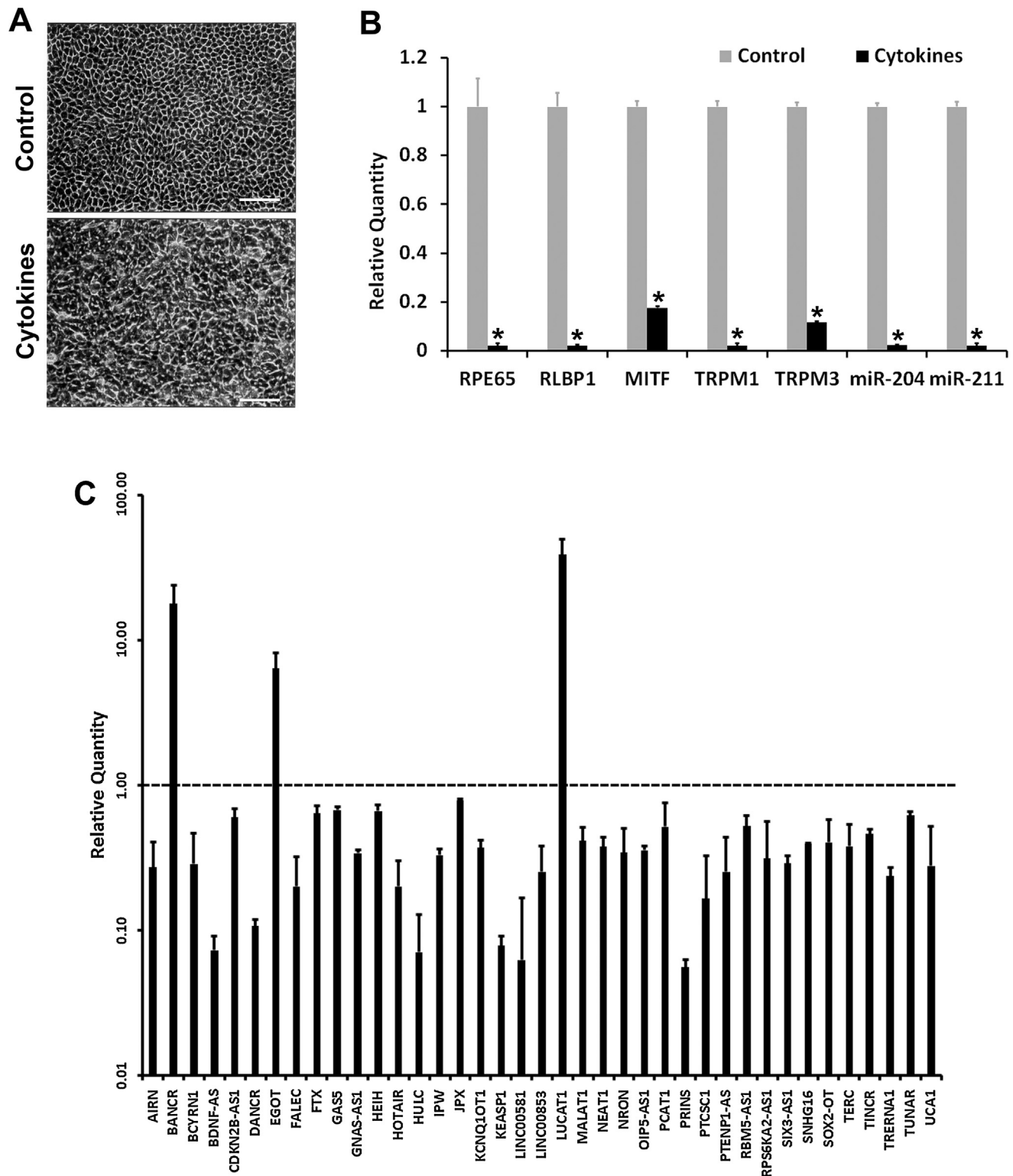
Human ARPE-19 cells were cultured until they exhibited RPE characteristics as recently described from our laboratory [11–13]. Briefly, the cells were grown in Dulbecco's modified Eagle's medium containing 4.5 g/L glucose, 4 mM L-glutamine, 1 mM sodium pyruvate,

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**Fig. 1.** Proinflammatory cytokines alter the expression of lncRNAs in RPE cells. Differentiated ARPE-19 cells were treated for 4 days with a proinflammatory cytokine mixture consisting of IFN- $\gamma$  (100 u/ml), TNF- $\alpha$  (10 ng/ml) and IL-1 $\beta$  (10 ng/ml). (A), Phase contrast microscopic images of control and treated cells shows that proinflammatory cytokines caused a disruption in the epithelial morphology of ARPE-19 cells. Photomicrographs were taken at a magnification of 100 $\times$ ; scale bar = 100  $\mu$ m. (B) Real-time PCR analysis shows that proinflammatory cytokines decrease the expression of RPE characteristic genes and miRNAs in ARPE-19 cells. \* =  $p < .05$  when compared to control,  $n = 3$ . (C) Real-time PCR analysis shows that proinflammatory cytokines alter the expression of lncRNAs in ARPE-19 cells. Relative expression level of lncRNAs affected by the treatment are shown ( $p < .05$  when compared to control;  $n = 3$ ). Broken line indicates that the relative expression level of an lncRNA in the control (untreated cells) is 1.

100 U/ml penicillin, 100  $\mu$ g/ml streptomycin and 1% fetal bovine serum at 37  $^{\circ}$ C in a humidified environment of 5% CO<sub>2</sub>. The culture medium was replaced twice every week. The cells kept under these conditions for 4 months exhibited RPE characteristics such as

pigmentation, epithelial morphology and expressed visual cycle genes. The cells were treated with the proinflammatory cytokine mixture consisting of IFN- $\gamma$  (100 u/ml), IL-1 $\beta$  (10 ng/ml) and TNF- $\alpha$  (10 ng/ml) in the presence of serum for 4 days unless indicated otherwise. Human

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