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Research paper

Human-porcine reassortant rotavirus generated by multiple reassortment events in a Sri Lankan child with diarrhea



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ABSTRACT

A human-porcine reassortant rotavirus, strain R1207, was identified from 74 group A rotaviruses detected in 197 (37.6%) stool samples collected from patients who attended a tertiary care hospital in Ragama, Sri Lanka. This is the first report of a human-porcine reassortant rotavirus in Sri Lanka. The patient was a 12-month-old boy who had been hospitalized with fever and acute diarrhea with a duration of 6 days. *The family had pigs at home before the birth of this boy. However, the neighbors still practice pig farming.* The genotype constellation of R1207 was G4-P [6]-I1-R1-C1-M1-A1-N1-T1-E1-H1. This is based on the assignment of all the eleven gene segments a full genome-based genotyping system. R1207 showed a 4-2-3-2 genomic electrophoretic migration pattern, which is characteristic of group A rotaviruses. Our analyses revealed that five (NSP2, NSP4, VP1, VP2, and VP7) of the 11 genes were closely related to the respective genes of porcine strains. Although the remaining six genes (NSP1, NSP3, NSP5, VP3, VP4, and VP6) were related to human strains, with the exception of the gene sequence of NSP1, all of these human strains were human-porcine reassortants. With a genogroup 1 genetic backbone, this strain was possibly formed via multiple genetic reassortments. We do not know whether this strain is circulating in pigs, as no data are available on porcine rotaviruses in Sri Lanka. Surveillance should be strengthened to determine the epidemiology of this genotype of rotavirus in Sri Lanka and to assess whether the infection was limited or sustained by ongoing human-to-human transmission.

1. Introduction

Rotavirus is one of the important causes of gastroenteritis in children and is responsible for over 200,000 deaths annually in children under five years of age (Tate et al. 2016), over 90% of these deaths occur in developing countries. The virus is divided into ten groups A-J (Banyai et al. 2017), with group A being most common in human. Rotavirus has a double-stranded RNA genome which are divided into 11 segments, those encode six structural (VP1, VP2, VP3, VP4, VP6, and VP7) and six non-structural proteins (NSP 1–6) (Matthijnssens et al. 2008b). Based on the nucleotide sequences of the genes for two capsid proteins, VP7 and VP4, a binary classification system, G and P genotypes, is used to classify different strains of rotaviruses. Till now 35 G

genotypes and 50 P genotypes have been reported (Agbemabiese et al. 2017). Among numerous possible G and P genotype combinations, only a limited number of genotype combinations are found commonly in human infections, they are G1P[8], G2P[4], G3P[8], G4P[8], and G9P [8].

Some of the rotavirus genotypes that have emerged and spread in humans worldwide during the last decade are thought to have originated by zoonotic transmission from different animals and gene reassortment (Martella et al. 2010). Although rotaviruses infect particular host species preferentially (for which they have been defined as the homologous strains), heterologous rotavirus infections may occur (Martella et al. 2010; Midgley et al. 2012). Evidence of interspecies transmission and genetic reassortment between human and animal ro-

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Table. 1

G and P consensus, type-specific and gene-specific primers used for RT-PCR amplification.

| Primer name | 5'- sequence – 3' | Position | Product size (bp) |
|-----------------------|-----------------------------------|----------------|-------------------|
| VP7 | | | |
| Beg9 | GGCTTTAAAAGAGAGAATTTCCGTCTGG | 1–28 | 1062 |
| End9 | GGTCACATCATACAATTCTAATCTAAG | 1062-1036 | |
| G-typing | | | |
| VP7-R | AAC TTG CCA CCA TTT TTT CC | 914–932 | |
| aBT1 | CAAGTACTCAAATCAATGATGG | 314–335 | 618 |
| aCT2 | CAATGATATTAACACATTTTCTGTG | 411-435 | 521 |
| aDT4 | CGTTTCTGGTGAGGAGTTG | 480-498 | 452 |
| G3 | ACG AAC TCA ACA CGA GAG G | 250-269 | 682 |
| G9 | CTT GAT GTG ACT AYA AAT AC | 757–776 | 179 |
| VP4 | | | |
| Con3 | TGGCTTCGCCATTTTATAGACA | 11-32 | 876 |
| Con2 | ATTTCGGACCATTTATAACC | 868-887 | |
| P-typing | | | |
| HumCom5 | CTC TCG ATG GTC CAT ATC AAC C | 200–221 | |
| P[8]P1A | TGT ACG TCT ATT ATA AAA TTC ATT T | 456-480 | 280 |
| P[4]P1B | ATA TAT TGC CTA TTT GTT TGA C | 347–368 | 168 |
| P[6]P2 | GTA TTA CAG TTT CTA CTT CAG A | 592-613 | 413 |
| P[9]P3 | CGT CGC TCC TTG ATA CCA GT | 533-552 | 352 |
| Gene-specific primers | | | |
| VP1 F | GGCTATTAAAGCTRTACAATGG | 1–22 | 3299 |
| VP1 R | CACATCTAAGCACTCTAATCTTG | 3277-3299 | |
| VP2 F | GGCTATTAAAGGCTCAATGG | 1–20 | 2696 |
| VP2 R | TTGGCGTTTACARTTCGTTCA | 2676-2696 | |
| VP3 F | TGCGTTTTACCTCTGATGGTG | 24–44 | 2566 |
| VP3 R | TCACATCATGACYAGTGTGTTAAG | 2566-2589 | |
| VP4 F | GGCTATAAAATGGCTTCGCT | 1–20 | 2362 |
| VP4 R | GGGGGTCACATCCTC | 2348-2359 (+3) | |
| VP6 F | GGCTTTWAAACGAAGTCTTC | 1–20 | 1356 |
| VP6 R | GGTCACATCCTCTCACT | 1340-1356 | |
| VP7 F | GGCTTTAAAAGMGAGAATTTCC | 1–22 | 1062 |
| VP7 R | GGGGGTCACATCATACAATTCT | 1041-1059 (+3) | |
| NSP1 F | GGCTTTTTTTATGAAAAGTCTTGTG | 1–25 | 1547 |
| NSP1 R | CTAGGCGCTACTCTAGT | 1531–1547 | |
| NSP2 F | GGCTTTTAAAGCGTCTCAGTC | 1–21 | 1058 |
| NSP2 R | GGTCACATAAGCGCTTTCTATTC | 1036-1058 | |
| NSP3 F | GGCTTTTAATGCTTTTCAGTGGTTG | 1–25 | 1050 |
| NSP3 R | GGTCACATAACGCCCCTATAG | 1030-1050 | |
| NSP4 F | CTTTTAAAAGTTCTGTTCCGAGAG | 3–26 | 739 |
| NSP4 R | AAGACCATTCCTTCCATTAAC | 721–741 | |
| NSP5 F | GGCTTTTAAAGCGCTACAGT | 1–20 | 663 |
| NSP5 R | GGTCACAAAACGGGAGTGGGGA | 642–663 | |

Reference: Gouvea et al. JCM 1990.Vol.28.No2.276-282

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taviruses has been accumulating continuously in the literature. Several studies have shown that G3 and G6 are reassortant of human-canine and human-bovine strains, respectively (Martella et al. 2010). Even the widely circulating G9 and G12 strains are reassortants of human-porcine strains (Matthijnssens et al. 2010). Reassortant rotaviruses bearing a mixed genome constellation from human and animal rotaviruses seem to be more successful compared with completely heterologous rotaviruses (Martella et al. 2010; Nakagomi and Nakagomi 2002). A whole genome based rotavirus classification system denotes the genome of rotavirus as Gx-Px-Ix-Rx-Cx-Mx-Ax-Nx-Tx-Ex-Hx where x represents the genotype number (Matthijnssens et al. 2008a). In this classification system human rotavirus strains are grouped into the Wa-like (G1/G3/ G4-P[8]-I1-R1-C1-M1-A1-N1-T1-E1-H1), DS-1-like (G2-P[4]-I2-R2-C2-M2-A2-N2-T2-E2-H2), and AU-1-like (G3-P[9]-I3-R3-C3-M3-A3-N3-T3-E3-H3)(Matthijnssens et al. 2008a; Matthijnssens et al. 2008b). This classification system revealed that human Wa-like and porcine rotavirus strains share a common ancestor. Porcine rotavirus strains possess

the following genomic constellation (G3/4/5/9-P[6]/[7]/[13]/[19]/ [23]-I5-R1-C1-M1-A8-N1-T1/7-E1-H1)(Agbemabiese et al. 2017; Matthijnssens et al. 2008a; Monini et al. 2014; Silva et al. 2016; Theuns et al. 2015).

Rotavirus infection is a burden for the Sri Lankan health care system, as a considerable number of children suffer and die from rotavirus-associated diarrhea each year (Ahmed et al. 2010b; Chandrasena et al. 2009). Studies of rotavirus diarrhea in Sri Lanka are scarce, and our study performed during 2005–2006 showed that a diversity of rotavirus strains were circulating among children, with G9 as the dominant genotype, followed by G3, G12, G2,G1, and G4(Ahmed et al. 2010b). In that study, no strain of animal origin was detected, although the genotype could not be determined for 9% of the strains. During 2008–2009, we initiated another study for the surveillance of rotavirus diarrhea in the same locality and detected an animal-like rotavirus of genotype G4P[6] in a child with diarrhea. Genetic analyses of animal-like rotaviruses are important for the assessment of whether Download English Version:

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