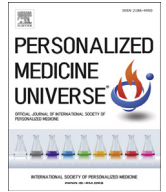




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Review

Cloning and expansion of antigen-specific T cells using iPSC cell technology: Possible use of regenerated T cells in personalized medicine

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Recent advances in adoptive immunotherapy using cytotoxic T lymphocytes (CTLs) have demonstrated that CTLs are effective in killing tumor cells *in vivo* for some tumor types. However, a critical issue that CTLs collected from patients are easily exhausted during expansion culture is yet to be addressed. Therefore, we have been developing a strategy that utilizes induced pluripotent stem cell (iPSC) technology, based on the idea that iPSCs produced from antigen-specific CTLs would regenerate CTLs with the same antigen specificity as the original CTLs. We previously succeeded in regenerating melanoma antigen MART1-specific CTLs, and more recently in producing potent CTLs expressing the CD8 $\alpha\beta$ heterodimer. We are now developing a novel method by which non-T derived iPSCs are transduced with exogenous T cell receptor (TCR) genes. If this method is applied to the allogeneic transfusion setting wherein HLA haplotype-homozygous iPSC stocks are used as the cell source, it will be possible to prepare “off-the-shelf” T cells. We are also considering incorporation of a personalized medicine approach to this allogeneic setting. In such a scheme, genes encoding TCRs specific for neoantigens will be collected from patients and HLA-homo iPSCs will be transduced with these TCR genes. Using such iPSCs, it will be possible to produce allogeneic CTLs expressing autologous TCRs originating from patients.

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1. Current status and recent issues in cancer immunotherapy

Cancer immunotherapy has evolved remarkably during the past several years. Immune checkpoint blockade drugs such as anti-CTLA-4 monoclonal antibody (mAb) and anti-PD-1 mAb have shown therapeutic effects [1–3]. However, their therapeutic efficacy reaches only 20–30% and autoimmune-related adverse events are frequently observed as side effects [4].

Delivery of autologous CTLs directly into patients has also shown good results. For example, Rosenberg and colleagues have demonstrated that transfusion of *ex-vivo* expanded tumor-infiltrating lymphocytes (TILs) was effective in melanoma patients [5]. Transfer of T cell receptor (TCR) genes such as NY-ESO1 antigen-specific one, into patients' peripheral T cells also achieved good clinical outcomes

[6–8], indicating that targeting a single antigen can be effective for some types of cancer. In addition, T cells enforced to express the chimeric antigen receptor (CAR) have shown enormous efficacy in the treatment of B cell leukemia [9].

In both TCR and CAR engineering, peripheral T cells are transduced by a retrovirus, introducing the risk of tumorigenicity due to the random genomic integration of the transfected gene. Moreover, in the autologous setting, it would be costly to produce T cells, and it is not always easy to obtain potent CTLs from patients.

2. Application of reprogramming technologies for the cloning and expansion of T cells

To address the aforementioned problems, we have thought of a method by which T cells can be expanded. When embryonic stem cells (ESCs) or iPSCs are generated from T cells, the genomic structure of rearranged T cell receptor (TCR) genes should be passed on to the resulting ESCs/iPSCs, and T cells regenerated from these ESCs/iPSCs should express the same TCR as the original T cells

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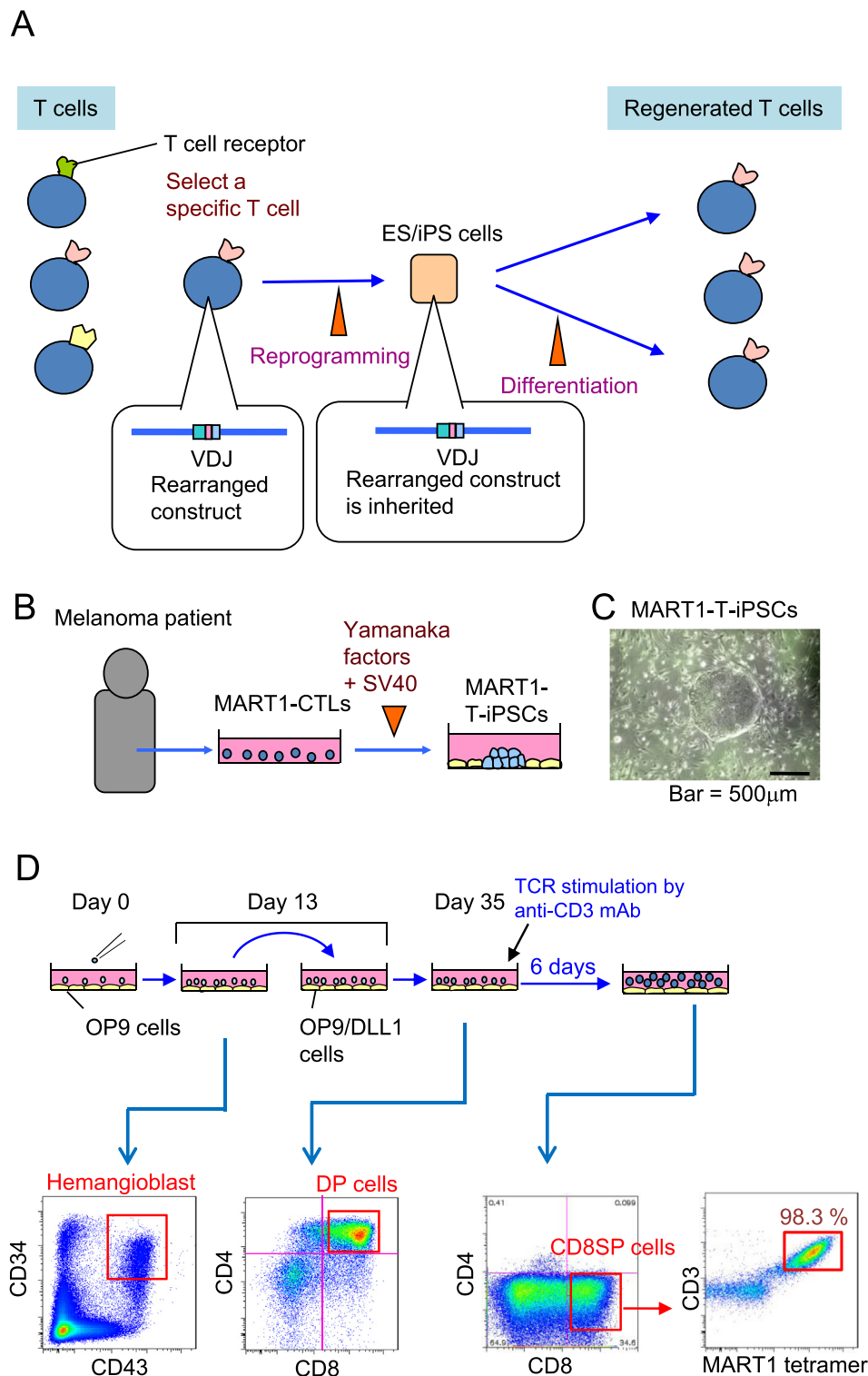


Fig. 1. Regeneration of T cells using iPSC technology. **A.** The concept of T-iPSC therapy. When T-iPSCs are established from tumor antigen-specific T cells, the rearranged configuration of TCR genes is passed on to iPSCs. T cells regenerated from these T-iPSCs express the same tumor antigen-specific TCRs. **B.** We established MART1-specific T-iPSCs by transducing MART1-specific T cells derived from a melanoma patient with Yamanaka factors and SV40 using the Sendai virus vector system. **C.** A colony of MART1-T-iPSCs. **D.** MART1-iPSCs were sequentially cultured with two types of feeder cells, OP9 and OP9/DLL1. On Day 13, CD34⁺CD43⁺ hemangioblasts were generated, and on Day 35, CD4/CD8 double-positive (DP) cells were generated, when anti-CD3 mAb was added to induce the generation of mature T cells. CD8 single-positive cells were generated 6 days after anti-CD3 mAb stimulation. Virtually all CD8 single-positive cells expressed a TCR specific for MART1 antigen.

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