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## Original Article

## Inter-observer variability in the assessment of ultrasound features of polycystic ovaries

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## ABSTRACT

**Objective:** To evaluate inter-observer variability in the assessment of ultrasound features of polycystic ovaries.

**Design:** Prospective cohort study.

**Population:** 60 females with known polycystic ovarian syndrome (PCOS).

**Setting:** Two tertiary care hospitals in Amman, Jordan; Prince Hamza Hospital and Albashir Hospital.

**Methods:** A total of 120 transvaginal scans were performed on 60 study participants with known PCOS by two gynecologists. The ovaries were evaluated for the presence or absence of PCOS criteria: The number of follicles and the volume of each ovary. The correlation coefficient was calculated. Bland-Altman plots were used to analyze any discrepancies in the measurement of ovaries between the observers.

**Main outcome measures:** Identification of ultrasound features of PCOS will be reproducible by different gynecologists.

**Results:** The mean follicle count and ovarian volume in PCOS patients reported in this study were 21 cm<sup>3</sup> follicles and 11.5 cm<sup>3</sup> respectively. The Pearson correlation coefficient was calculated and it can be seen that there was low inter-observer correlation in follicular count (0.560) and a moderate correlation in measuring ovarian volume (0.770). Bland-Altman plots show low inter-observer agreement in follicular count and high inter-observer agreement in the measurement of ovarian volume.

**Conclusion:** The inter-observer agreement in the assessment of ultrasound features of polycystic ovaries is not acceptable. The diagnostic criteria of polycystic ovarian morphology need to be revisited.

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## 1. Introduction

Polycystic ovarian syndrome was first described by Stein and Leventhal as an association of amenorrhea, obesity and an atypical appearance of polycystic and enlarged ovaries at laparotomy [1]. It is the most frequently occurring endocrine pathology among women of reproductive age, with an estimated prevalence of 5–10% of women in reproductive age [2]. The diagnostic criteria for PCOS remains an area of heated debate since the pathophysiology of PCOS is not well understood. The NIH criteria is the first definition that received consensus from the scientific community, which defined PCOS as the combined presence of hyperandrogenism and/or hyperandrogenemia, oligoovulation and exclusion of related disorders such as Cushing's syndrome, hyperprolactinemia, and congenital adrenal hyperplasia [3]. The Rotterdam criteria for PCOS

serves as the second definition to receive consensus and describes PCOS as the presence of at least two of three cardinal features: Oligoovulation, hyperandrogenism and polycystic ovary morphology [4]. Polycystic ovarian morphology was characterized as containing 12 or more follicles measuring 2–9 mm and/or an increased ovarian volume of more than 10 cm<sup>3</sup> [5]. The difference between the NIH criteria and the Rotterdam criteria is an emphasis of polycystic ovarian morphology as a separate cardinal feature in the latter.

Only two studies have examined inter- and intra-observer variability when making the ultrasound diagnosis of PCOS. Both studies have shown poor inter-observer agreement in identifying features of PCOS on ultrasound. The first was a prospective observational study undertaken to evaluate the inter-observer reliability of ultrasound diagnosis of PCOS. The authors of this study evaluated 27 women and concluded that there is significant variability between different operators when attempting to diagnose PCOS using ultrasound criteria [6]. A more recent study investigated inter-observer agreement when identifying and quantifying

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individual ultrasonographic features of polycystic ovaries. Four operators evaluated the digital recording of transvaginal ultrasound of thirty women with PCOS and found that the inter-observer agreement between different operators was low [7].

Ultrasound technology has advanced significantly in recent years. The identification of PCOS using ultrasound must be easier to apply and the recorded inter-observer variability must decrease for a reliable diagnosis to be made. In this study we want to measure inter-observer variability in the assessment of ultrasound features of polycystic ovaries and interpret and discuss the results as they are reflected on real life applications and diagnoses of PCOS. We will discuss the methods and the process of patient examination in Section 2. Section 3 will highlight the results, while Section 4 will discuss current evidence and explore the significance of our results in real life applications. Section 5 will highlight our conclusions.

## 2. Materials and methods

### 2.1. Examination process

Ethical approval was obtained from the Hashemite University Ethics Committee and from Prince Hamzah Hospital Ethics Committee.

Spanning the period from December 1, 2015 to March 1, 2016, one hundred and twenty women were recruited for this study. Women recruited for the study group were diagnosed with PCOS using the 2003 international consensus guidelines of having two of three characteristics:

- Oligoovulation or anovulation
- Clinical and/or biochemical evidence of hyperandrogenism
- Polycystic ovaries on ultrasound ( $\geq 12$  follicles measuring 2–9 mm in diameter or an ovarian volume  $>10$  cm<sup>3</sup>) [4]

Exclusion criteria were those using hormonal contraception, fertility medications and/or valproate in the two months prior to enrolment and the inability to visualize the ovaries on vaginal ultrasound. 81 participants with PCOS were approached and 8 participants were excluded. 6 participants were excluded for the use of fertility medication in the 2 month prior to the study. 2 participants were excluded due to the use of combined birth control pills. All participants were tested for prolactin levels, cortisol levels, thyroid function test, and 17 hydroxy-progesterone levels.

Women who met the inclusion criteria and agreed to participate in the study were administered two transvaginal ultrasound scans performed by two different operators within 10 min of each other. The first scan was performed by “Operator 1” and the second scan was performed by “Operator 2”. Each examiner was given 15 min to complete the exam. Only one operator was present in the exam room during patient examination. All ultrasound tests were performed using the same machine, a Samsung Medison R5 (South Korea), with a transvaginal transducer. The two gynecologists performing the examination were certified obstetricians and gynecologists with significant experience in administering transvaginal ultrasound scans. The scans were performed at a random time during the menstrual cycle of each patient. Each ovary was visualized and the anatomic orientation relative to the utero-ovarian ligament was established. Ovaries were scanned from the inner to outer margins in both transverse and sagittal planes. Gynecologists performing the scans were asked to count the total number of follicles ( $\geq 2$  mm) in each ovary. Ovarian volume was then calculated using the equation for a spheroid from measurements of the largest and widest diameters of the ovaries in the transverse and sagittal planes [8].

### 2.2. Statistical analysis

Data used for the descriptive statistics were obtained from clinical and laboratory records. Mean measurements of follicular counts and ovarian volume were compared among observers using the inter-class correlation coefficient (Pearson coefficient). Bland-Altman graphs were used to analyze the degree of agreement between observers. Guidelines for evaluating the level of agreement among scores were:  $>0.80$  for high/good,  $0.60$ – $0.80$  for moderate/fair, and  $<0.60$  for low/poor [9].

## 3. Results

A total of 72 women met the inclusion criteria and agreed to participate in the study. It was not possible to obtain acceptable images of the ovaries in 12 participants, mainly due to the degree of their obesity. Acceptable images were obtained and included in the study in 60 participants. The mean age of participants was 27.4 years with a range of 18.0–35.0 years. Participants had a mean BMI of 30.2% with a range of 20–40%. The mean menstrual cycle length was 74 days with a range of 31–211 days. Clinical and metabolic features of the women participating in the study can be seen in Table 1.

Descriptive statistics of the follicular counts and ovarian volume can be seen in Table 2. It can be seen in Figs. 1 and 2 that the number of follicles found on each ovary has poor agreement between observers and there is random distribution around the equality line indicating no bias. It can also be seen in Figs. 3 and 4 that ovarian volume measurement has good agreement between observers due to the relation of each value to the equality line. It can also be seen in Table 3 that there is poor inter-observer correlation in obtaining follicular count and moderate inter-observer correlation in measuring ovarian volume after measuring the Pearson correlation coefficient.

The Bland-Altman graph is a scatterplot of variable means plotted on the horizontal axis and the differences plotted on the vertical axis which shows the amount of disagreement between the two measurements. This plot includes approximate 95% limits. If differences observed in this plot are not deemed clinically important, this is a confirmation of agreement. Bland-Altman graphs for the differences in follicular number count measurements between

**Table 1**

The mean clinical and metabolic features of study participants.

	Mean	Range	Normal value
Age (years)	27.4	18–35	–
BMI (%)	30.2	20.0–40.0	20.0–25.0
Menstrual cycle length (days)	74.0	31.0–211	21.0–35.0
LH:FSH	2.40	0.70–6.70	<2.00
SHBG (nmol/L)	44.0	16.0–76.0	18.0–114
Fasting glucose (mmol/l)	4.90	4.00–6.40	<6.10

**Table 2**

Follicular count and ovarian volume as measured by Examiner A and Examiner B.

Examiner A	Mean	Examiner B	Mean
Rt. ovarian follicular count	22.5 ± 7.80	Rt. ovarian Follicular count	21.0 ± 6.70
Lt. ovarian follicular count	24.5 ± 7.20	Lt. ovarian Follicular count	23.3 ± 6.60
Rt. ovarian volume (cm <sup>3</sup> )	11.3 ± 1.80	Rt. ovarian Volume (cm <sup>3</sup> )	11.4 ± 1.50
Lt. ovarian volume (cm <sup>3</sup> )	11.7 ± 1.40	Lt. ovarian Volume (cm <sup>3</sup> )	11.8 ± 1.10

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