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## Transcription Factors in Regulatory and Protein Subnetworks during Generation of Neural Stem Cells and Neurons from Direct Reprogramming of Non-fibroblastic Cell Sources

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Abstract—Direct reprogramming of non-fibroblastic cells to the neuronal cell types including induced neurons 12 (iNs) and induced neural stem cells (iNSCs) has provided an alternative approach for the direct reprogramming of fibroblasts to those cells. However, to increase the efficiency of the reprogramming process the underlaying mechanisms should be clarified. In the current study, we analyzed the gene expression profiles of five different cellular conversions to understand the most significant molecular mechanisms and transcription factors (TFs) underlying each conversion. For each conversion, we found the list of differentially expressed genes (DEGs) and the list of differentially expressed TFs (DE-TFs) which regulate expression of DEGs. Moreover, we constructed gene regulatory networks based on the TF-binding sites' data and found the most central regulators and the most active part of the network. Furthermore, protein complexes were identified from constructed protein-protein interaction network for DE-TFs. Finally, we proposed a list of main regulators for each conversion; for example, in the direct conversion of epithelial-like cells (ECs) to iNSCs, combination of centrality with active modules or protein complex analyses highlighted the role of POU3F2, BACH1, AR, PBX1 SOX2 and NANOG genes in this conversion. To the best of our knowledge, this study is the first one that analyzed the direct conversion of non-fibroblastic cells toward iNs and iNSCs and we believe that the expression manipulation of identified genes may increase efficiency of the process. © 2018 Published by Elsevier Ltd on behalf of IBRO.

Key words: active modules, direct reprogramming, non-fibroblastic cells, protein complexes, transcription factors, regulatory networks.

## INTRODUCTION

Failure in function of central nervous system cells is the 14 origin of neuronal disorders that there are no effective 15 drug treatments or therapeutic strategies for most of 16 them yet. Neurons as the most abundant neuronal cells 17 and neural stem cells (NSCs) attract a lot of attention as 18 they may be used to treat neural disorders or to study 19 them in the laboratory. Acquisition of such cells from 20 human tissues has its own limitation due to the ethical 21 concerns (Breunig et al., 2011). In this regard, generation 22 of them through differentiation of stem cells or direct 23

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E-mail addresses: Moein.yaqubi@mail.mcgill.ca (M. Yaqubi), Abdulshakour.mohammadnia@mail.mcgill.ca (A. Mohammadnia). <sup>†</sup> These authors contributed equally to this work.

Abbreviations: ChEA, Chip-Enrichment Analysis; DEGs, differentially expressed gene; DE-TFs, differentially expressed TFs; ECs, epitheliallike cells; iNs, induced neurons; iNSCs, induced neural stem cells; NRPCs, neural restricted progenitor cells; NSCs, neural stem cells; PRC2, polycomb repressive complex 2; TFs, transcription factors. reprogramming of somatic cells has become a promising 24 approach to replace damaged cells, study central nervous 25 system disorders, and investigate nervous system devel-26 opment. It has been shown that fully differentiated 27 somatic cells can be reprogrammed to a pluripotent state 28 using defined transcription factors (TFs) (Takahashi and 29 Yamanaka, 2006; Takahashi et al., 2007). These pluripo-30 tent stem cells can then be differentiated to the functional 31 cells. However, differentiation of iPSCs to somatic cells is 32 a time consuming, non-efficient process, variable among 33 various lines of iPSCs (Hu et al., 2010) and harbors the 34 risk of tumor formation upon in vivo transplantation 35 (Pera, 2011). Alternatively, somatic cells can be con-36 verted to another type of somatic cells in a process called 37 direct reprogramming (see Fig. 1). 38

The direct reprogramming allows conversion of 39 somatic cells to each other through forced expression of 40 TFs, miRNA and application of small molecules. So far, 41 extensive efforts have been done for generation of 42 induced neurons (iNs) (Caiazzo et al., 2011; Marro 43

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**Fig. 1.** Overall representation of the methods which we used in our study. Firstly, transcriptomic data sets were obtained from GEO database. Following identification of differentially expressed genes (DEGs), the data of transcription factor (TF)-binding site and protein–protein interactions were recruited to construct integrated gene regulatory network (GRN) and protein–protein interaction (PPI) networks. Finally, the GRN was subjected to different analyses, including identification of the most activated modules, the most central genes and the most affected cellular processes. In addition, the PPI network was analyzed to find the top protein complexes.

et al., 2011a; Son et al., 2011; Ladewig et al., 2012; Liu
et al., 2013; Wapinski et al., 2013; Zhou et al., 2014;
Colasante et al., 2015; Hu et al., 2015; Li et al., 2015)
or NSCs (Lee et al., 2011; Han et al., 2012; Matsui

et al., 2012; Ring et al., 2012; Thier et al., 2012;48Cassady et al., 2014) from somatic cells, specially fibrob-49last. However, the direct conversion of somatic cells50toward iNs and NSCs is not restricted to the fibroblasts.51

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