

## HSP70 Facilitates Memory Consolidation of Fear Conditioning through MAPK Pathway in the Hippocampus

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**Abstract**—Heat shock proteins of the 70-kDa (HSP70) family are cytoprotective molecular chaperones that are present in neuronal cells and can be induced by a variety of homeostatically stressful situations (not only proteostatic insults), but also by synaptic activity, including learning tasks. Physiological stimuli that induce long-term memory formation are also capable of stimulating the synthesis of HSP70 through the activation of heat shock transcription factor-1 (HSF1). In this study, we investigated the influence of HSP70 on fear memory consolidation and MAPK activity. Male rats were trained in contextual fear conditioning task and HSP70 content was analyzed by western blot in the hippocampus at different time points. We observed rapid and transient elevations in HSP70 60 min following training. Double immunofluorescence with GFAP and HSP72 revealed that astrocytes were not the site for HSP72 induction by CFC training. HSP72 distribution markedly surrounded synapses between Shaffer collateral and CA1 pyramidal cells. Infusion of recombinant HSP70 (*hspa1a*) into the dorsal hippocampus immediately after training facilitated memory consolidation and enhanced ERK activity while decreasing the activated forms of JNK and p38 in the hippocampus. Blocking endogenous extracellular HSP70 through the administration of specific antibody did not produce any further effect on memory consolidation when applied immediately after training, suggesting that it is indeed acting intracellularly. Induction of HSP70 after fear conditioning is fast and can act as a signaling molecule, modulating MAPK downstream signaling during memory consolidation in the hippocampus, which is crucial for fear memory formation. © 2018 IBRO. Published by Elsevier Ltd. All rights reserved.

**Key words:** molecular chaperone, HSP70, contextual fear conditioning, MAPK.

### 1. INTRODUCTION

Memory consolidation is a process in which new memories are transformed from a labile state to a more

stable one. This process depends on the activation of kinases, transcription factors, increased gene expression and protein synthesis in the postsynaptic neuronal cell (Elgersma and Silva, 1999; Abel and Lattal, 2001; Izquierdo et al., 2006). One of the most important pathways for long-term memory formation is the activation of protein kinases by glutamate receptor signaling that leads to the activation of CREB (cAMP responsive element-binding protein), a transcription factor responsible for the activation of different genes involved in memory consolidation (Suzuki et al., 2011; Izquierdo et al., 2006). Among those kinases, the mitogen-activated protein kinase (MAPK) family occupies a critical position. MAPKs are divided into three different subfamilies, including the extracellular signal-regulated kinases (ERK), the c-Jun amino-terminal kinases (JNK) and the p38-MAPK (Seeger and Krebs, 1995). JNK was shown to

\*Correspondence to: L. de Oliveira Alvares, Neurobiology of Memory Lab, Biophysics Department, Biosciences Institute, Federal University of Rio Grande do Sul, Porto Alegre, RS 91509-900, Brazil. E-mail address: [lucas\\_alvares@yahoo.com](mailto:lucas_alvares@yahoo.com) (L. de Oliveira Alvares). **Abbreviations:** AMPA,  $\alpha$ -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid; CAMK,  $\text{Ca}^{2+}$ /calmodulin-dependent protein kinase; CFC, Contextual Fear Conditioning; CNS, central nervous system; CREB, cAMP responsive element-binding protein; ERK, extracellular signal-regulated kinases; GluR, glutamate receptor; HSF-1, heat Shock Factor 1; HSP70, heat shock proteins of the 70 kDa; HSP72, inducible form of heat shock protein 70; JNK, c-Jun amino-terminal kinases; LTD, long-term depression; LTP, long-term potentiation; MAPK, mitogen-activated protein kinase; NMDA, N-methyl D-aspartate; p-ERK, phosphorylated ERK; p-JNK, phosphorylated JNK; PKA, protein kinase A; PKC, protein kinase C; rHSP70, recombinant human HSP70.

36 be a negative regulator of associative learning and, along- 97  
37 side p38, is involved in synaptic plasticity, inducing long- 98  
38 term depression (LTD) (Moult et al., 2008; Sherrin et al., 99  
39 2010, 2011). Alternatively, the increase of cAMP and 100  
40  $Ca^{2+}$  levels in the postsynaptic cell enhances both pro-  
41 tein kinase A (PKA) and protein kinase C (PKC) activities,  
42 respectively, leading to the activation of the ERK path-  
43 way, CREB phosphorylation and the initiation of transcrip-  
44 tion of several genes (Alberini, 2009; Kandel, 2012;  
45 Johansen et al., 2011; Roberson et al., 1999). HSP70, a  
46 member of the 70-kDa family of heat shock proteins  
47 (HSPs), is a potential target gene due to the presence  
48 of a CRE motif in its promoter region that can be activated  
49 by CREB (Choi et al., 1991; Murshid et al., 2010).

50 Inducible HSP70 (or HSP72, encoded by *HSPA1A*  
51 gene in humans) is a cytoprotective molecular  
52 chaperone (Lindquist and Craig, 1988), which is synthe-  
53 sized in the central nervous system (CNS) under a variety  
54 of homeostatically stressful situations, including heat  
55 shock, glucose and oxygen deprivation, glutamatergic  
56 excitotoxicity and psychophysiological stress (Belay and  
57 Brown, 2006; Lee et al., 2001). The synthesis of this pro-  
58 tein under these conditions protects cells against oxida-  
59 tive stress and cell death, since HSP70 is capable of  
60 blocking inflammation and apoptosis signaling (Beere  
61 et al., 2000; Garrido et al., 1999). Exogenous HSP70 is  
62 able to cross the blood–brain barrier, protect motor neu-  
63 rons from death induced by energy deprivation  
64 (Robinson et al., 2005), attenuate seizures (Ekimova  
65 et al., 2010) and is envisaged as a potential treatment in  
66 Alzheimer's disease, diminishing accumulation of  
67 amyloid- $\beta$  and protecting against spatial memory deficits  
68 in animal models of the disease (Bobkova et al., 2014).

69 In neurons, HSP70 is present in postsynaptic  
70 structures (Suzuki et al., 1999) where it can be induced  
71 by synaptic activation (Rao and Steward, 1991; Kaneko  
72 et al., 1993). Physiological stimuli that induce long-term  
73 memory formation, such as increased  $Ca^{2+}$ , PKC and  
74  $Ca^{2+}$ /calmodulin-dependent protein kinase (CAMK)  
75 levels, are also capable of stimulating the synthesis of  
76 HSP70 following on from the activation of its most impor-  
77 tant transcription factor (Heat Shock Factor-1, HSF1)  
78 (Price and Calderwood, 1991). Elevation of HSP70 by  
79 heat shock prevents the suppression of long-term poten-  
80 tiation (LTP) induced by scopolamine in hippocampal  
81 slices (Lin et al., 2004). Similar results have also been  
82 observed *in vivo*, in which heat shock pretreatment has  
83 shown to block the amnesic effect of scopolamine in  
84 the inhibitory avoidance test just 16 h following interven-  
85 tion, a time point of HSP70 peak in the hippocampus  
86 (Hung et al., 2004).

87 Increased HSP70 mRNA and protein content was  
88 found to be increased following learning, using different  
89 protocols. HSP70 is induced in the hippocampus  
90 following aversive and spatial learning (Pizarro et al.,  
91 2003; Igaz et al., 2004) and in the cerebellum following  
92 a two-way avoidance task (Ambrosini et al., 2005), which  
93 suggests that its expression is dependent on the region  
94 engaged in the task. Despite numerous assumptions  
95 regarding the involvement of HSP70 in synaptic plasticity  
96 and memory, there is no concrete evidence of its role and/

or downstream signaling in memory formation besides its  
chaperone function. Therefore, the aim of our study was  
to verify the influence of HSP70 on memory consolidation  
and its possible downstream signaling pathways.

## 2. MATERIALS AND METHODS

### Animals

Adult male Wistar rats (270–350 g) from our breeding  
colony were housed four to five per cage and  
maintained under constant temperature ( $23 \pm 1^\circ\text{C}$ ) with  
controlled photoperiods (12 h light/12 h dark; lights on at  
7:00 a.m.) and 60% relative humidity. A standard  
commercial laboratory diet (Nuvilab, Curitiba, Brazil)  
was provided *ad libitum*. All experiments were  
performed in accordance with local and national  
guidelines (Federal Law no 11.794/2008) for animal  
care and the project was approved by the Ethics  
Committee on Animal Experimentation of the Federal  
University of Rio Grande do Sul (CEUA n. 27791).

### Stereotaxic surgery and placement of cannulae

Rats were deeply anesthetized via an intraperitoneal  
injection of ketamine/xylazine (75 and 10 mg/kg,  
respectively) and bilaterally implanted with 27-gauge  
guide cannulae with respect to bregma aimed at AP  
–4.0 mm, ML  $\pm$  3.6 mm, DV –1.6 mm (from brain  
surface), positioned 1.0 mm above the CA1 area of the  
dorsal hippocampus (Paxinos and Watson, 1998). The  
animals were exposed to behavioral procedures one  
week after the surgery. Following behavioral  
experiments, the rats were euthanized and brains  
dissected and preserved in 10% formaldehyde to verify  
cannula position. Only animals with correct cannula  
placements were included.

### Drugs

Recombinant mouse low-endotoxin heat shock protein 70  
(hsp72, inducible form of HSP70, encoded by the *hspa1a*  
gene, Enzo, ADI-ESP-502) was diluted in Dulbecco's  
PBS pH 7.4, containing 8.1 mM sodium phosphate, 1.5  
mM potassium phosphate, 2.7 mM potassium chloride  
and 137 mM sodium chloride at a total concentration of  
either 0.25  $\mu\text{g}/\mu\text{L}$ , 0.5  $\mu\text{g}/\mu\text{L}$  or 1.1  $\mu\text{g}/\mu\text{L}$ . Anti-Heat  
Shock Protein 70 monoclonal antibody produced in  
mouse (Sigma, H5147, clone BRM–22) was diluted in  
PBS containing 15 mM sodium azide to a total  
concentration of 1  $\mu\text{g}/\mu\text{L}$  or 0.1  $\mu\text{g}/\mu\text{L}$ . The vehicle used  
was the buffer in which the drugs were diluted. Drugs  
were infused bilaterally into the dorsal hippocampus  
either immediately, 1 h or 6 h after the training session.

### Intrahippocampal infusion

At the time of infusion, a 30-gauge infusion needle was  
fitted into the guide cannula, with its tip protruding 1.0  
mm beyond the end of the guide cannula. A volume of  
1  $\mu\text{L}$  was infused bilaterally at a slow rate (20  $\mu\text{L}/\text{h}$ ) and  
the needle was removed 30 s following complete  
administration of the drug.

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