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# Ecotoxicity evaluation of Cu- and Fe-CNT complexes based on the activity of bacterial bioluminescence and seed germination

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#### ABSTRACT

The toxic effects of the composites of  $Fe^0$  and  $Cu^0$  with different percentages of CNTs were 17 examined based on the activity of bacterial bioluminescence and seed germination. In 18 terms of the  $EC_{50}$  values, the toxic effects of  $Cu^0$  on bacterial bioluminescence and seed 19 germination were approximately 2 and 180 times greater than that of  $Fe^0$ , respectively. The 20 toxicity increased with increasing CNT content in the Cu-CNT mixtures for both organisms, 21 whereas opposite results were observed with Fe-CNT mixtures. The mean toxic effects of 22 Cu-CNT (6%) were approximately 1.3–1.4 times greater than that of Cu-CNT (0%), whereas 23 the toxic effects of Fe-CNT (6%) were approximately 2.1–2.5 times lower than that of Fe-CNT 24 (0%) for both the bioluminescence activity and seed germination. The causes of this 25 phenomenon are unclear at this point. More research will be needed to elucidate the 26 mechanism of the toxicity of nano-mixture materials and the causes of the different 27 patterns of toxicity with Cu- and Fe-CNT mixtures. 28

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# 42 Introduction

43 Nanotechnology has emerged as an enabling technology with high potential impact on virtually all fields of mankind. Estimates 44 45 suggested that by 2015 nanotechnology will have a trillion-plus dollar global economic impact (Ge et al., 2012). Berman (2016) also 46 reported that nanotechnology will bring improvements by 47 48 addressing environmental problems using advanced nanotechnology by 2025. The implementation of unique materials and 49 devices ranging from electronics to engineered tissues is one of 50 the rapidly growing fields of nanotechnology (Shvedova et al., 51 2003). Two advanced nanomaterials that are being used increas-52 ingly for different technologies are carbon nanotubes (CNTs) and 53 54 graphene. CNTs are well-ordered, high aspect ratio allotropes of

carbon. CNTs have attracted considerable interest from both 55 scientists and industry because their unique atomic configura- 56 tion, mechanical, optical, electronic properties, high aspect ratios 57 (e.g., like fibers), strength, and remarkable physical properties 58 (Smart et al., 2006). Current global multi-wall CNTs production 59 capacity is estimated to be 13,996 tons (Zhao et al., 2017). Carbon 60 nanotube metal matrix composites are an emerging class of 61 new materials that are being developed because of the high 62 tensile strength and electrical conductivity of carbon nanotube 63 materials.

Powder metallurgy processing to prepare nanocrystalline 65 CNT composite powders with Cu and Fe have attracted 66 considerable attention in the next generation automobile 67 industry to manufacture the desired advanced automotive 68

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components because of their promising mechanical, electrical, 69 70 and thermal properties. Any exposure of humans to CNTs would 71 pose considerable danger to human health. Therefore, under-72 standing and ensuring the safety of nanomaterials are of huge importance to the tremendous commercial applications of 73 nanotechnology (Foldvari and Bagonluri, 2008; Tang et al., Q5 2012). Safety evaluation of nanomaterials should consider 75 their behaviors in various aspects, including their interactions 76 77 with proteins, DNA, lipids, membranes, organelles, cells, 78 tissues, biological fluids, and even the dissolution of metal constituents (Zhao et al., 2008; Nel et al., 2009; Liu et al., 2013). 06 80 There are different ways, in which the human body can 81 be exposed to nanomaterials, including penetration through the skin, ingestion, inhalation, and injection (Zhao and Liu, 82 83 2012).

84 A great effort in recent years has been performed to elucidate the mechanism of nanotoxicity in living organisms 85 and cells. For this purpose, scientists and researchers have 86 examined the toxicity of CNTs in several cell lines (Wang 87 et al., 2011; Pichardo et al., 2012). Their efforts suggest that 88 89 various factors, including functioning, dimensions, and char-90 acteristics, can determine the toxicity of CNTs, including environmental factors, such as temperature, humidity, and 91 92 barometric pressure, affecting rates of consumption and even 93 the occurrence of some toxic agents (Horie et al., 2012). 94 Recently, in vivo toxicology studies, by focusing on multiple 95 organ systems, have proved that CNTs can cause a toxic 96 response within these multiple organ systems. Research into 97 the ecotoxicity of nanomaterials has demonstrated different 98 levels of toxicity depending on the bioassays (Brunner et al., 99 2006; Soto et al., 2006). TiO<sub>2</sub> and ZnO nanomaterials, which are 100 used widely in sun protection products as well as self-cleaning coatings, have been shown to have toxic effects that can inhibit 101 the growth of microalgae, crustaceans, and bacteria, while 102 opposite results have been reported by other investigators 103 (Serpone et al., 2007; Heinlaan et al., 2008; Aruoja et al., 2009). 104 Ecotoxic bioassays using various test organisms (bacteria, algae, 105 protozoa, plants, and fish) and their metabolic processes have 106 107 gained widespread attention for environmental contaminants (Banks and Schultz, 2005). Knowledge of each test organism's 108 sensitivity is important for evaluating the contaminant toxicity 109 110 levels. Among these processes, seed germination and bacterial bioluminescence were adopted because of their high sensitivity, 111 simplicity, etc. Plant toxicity assays are particularly relevant 112 when the phytotoxic contaminants of nanomaterials are present 113 114 in soil (Boutin et al., 2004). Among plant processes, seed germination studies are considered short-term because they 115 assess the rapid response to acute toxicity. The root and shoot 116 elongation test is one of the simplest short-term methods used 117 in environmental biomonitoring (Di Salvatore et al., 2008). 118 Assays based on bacterial bioluminescence are a time-saving 119 and cost-effective test that are used widely as a reproducible and 120 sensitive screening method for determining the acute toxicity of 121 122 different sample types (Wang et al., 2002).

In this study, composites of Fe<sup>0</sup> and Cu<sup>0</sup> with different percentages of CNTs, which are currently used in laboratories and the automobile industry without any restriction by environmental law, were tested. The ecotoxic effects of composites of Fe and Cu with CNTs were evaluated based on the activity of bacterial bioluminescence and seed germination.

## 1. Materials and methods

## 1.1. Preparation of metals-CNT mixtures

**120** 131

CNTs were synthesized using a catalytic CVD method and 132 then dispersed in distilled water at concentration of 5 wt.% 133 CNT using a Sonosmasher. Carboxymethyl cellulose (CMC) 134 was used as the dispersant with a 9 sec working interval and 135 1 sec off continuously, totaling 6 hr in dispersion for the 136 finally dispersed product. Copper powder with a particle size 137 of up to 63  $\mu$ m was purchased from Markin Metal Powders Ltd. 138 (UK). Iron powders with a particle size of up to 150  $\mu$ m were 139 purchased from Hoganas Ltd. Each metal powder was placed 140 into a milling chamber with the CNTs solution for the attrition 141 ball milling process using SUS-316 L balls, 5 mm in diameter, 142 as the milling media. Ball milling was proceeded at 300 r/min 143 for 1 hr. The milled solution was then dried in an oven for 144 24 hr to produce the final milled and mixed metal-CNTs 145 powder. Four different conditions of the mixture samples 146 were prepared according to the CNT concentrations: 0, 1%, 3%, 147 and 6% (W/W). 148

# 1.2. Toxicity test of metals-CNT mixtures on bioluminescence 149 activity 150

The toxicity of the metal-CNT mixtures was measured based 151 on a bioluminescence activity of the Escherichia coli DH5a 152 strain RB1436, harboring a variant of the pUCD615 plasmid 153 (obtained from R. Burlage, Concordia University, USA). This 154 strain, which contains a constitutive promoter to express the 155 lux genes, produces bioluminescence in a growing culture and 156 is used to detect deleterious conditions that would cause a 157 measureable decrease in bioluminescent output, such as 158 those induced by metal-CNT mixtures. The RB 1436 strain 159 was stored at -70°C until needed, at which time, it was grown 160 overnight in Luria-Bertani<sup>ka</sup> (LB<sup>ka</sup>) medium at 27°C with 161 shaking at 130 r/min. The culture was diluted 30-fold in LB<sup>ka</sup> 162 medium and allowed to grow until the optical density (OD<sub>600</sub>) 163 reached approximately 0.6. This culture was diluted appro- 164 priately with minimum salt medium to a final  $OD_{600}$  of 0.2 for 165 the toxicity test (Ko and Kong, 2014). For the test, 1 mL of the 166 diluted bacterial suspension was mixed with 9 mL of the 167 sample, after which the bioluminescence activity was mea- 168 sured after 1 and 1.5 hr of incubation. The bioluminescence 169 activity was measured using a Turner 20/20 luminometer 170 (Turner Design, USA), which had a maximum detection limit 171 of 9999 relative light units (RLU). 172

### 1.3. Toxicity test of metals-CNT mixture on seed germination 173

The seed (Lactuca sativa L.) produced and distributed by a 174 commercial seed company (Nongwoo Bio., South Korea) were 175 purchased from a local seed store. These particular seeds 176 were employed in the test because the plants from which they 177 were obtained are important food crops in the local region. 178 Prior to the germination test, all seeds were surface-sterilized 179 with an aqueous 3% H<sub>2</sub>O<sub>2</sub> solution for 10 min and then rinsed 180 with distilled water. Filter paper was then placed in a sterilized 181 Petri dish and moistened with 5 mL of an aqueous solution 182

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