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## Perspective

# Analysis of key microbial community during the start-up of anaerobic ammonium oxidation process with paddy soil as inoculated sludge

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### ABSTRACT

A sequencing batch reactor (SBR)-anaerobic ammonium oxidation (anammox) system was started up with the paddy soil as inoculated sludge. The key microbial community structure in the system along with the enrichment time was investigated by using molecular biology methods (e.g., high-throughput 16S rRNA gene sequencing and quantitative PCR). Meanwhile, the influent and effluent water quality was continuously monitored during the whole start-up stage. The results showed that the microbial diversity decreased as the operation time initially and increased afterwards, and the microbial niches in the system were redistributed. The anammox bacterial community structure in the SBR-anammox system shifted during the enrichment, the most dominant anammox bacteria were *Candidatus Jettenia*. The maximum biomass of anammox bacteria achieved  $1.68 \times 10^9$  copies/g dry sludge during the enrichment period, and the highest removal rate of TN achieved around 75%.

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### Introduction

Nitrogen removal has taken more and more attention in the field of wastewater treatment. The conventional biological nitrogen removal process has many disadvantages such as high energy and material consumptions. However, the discovery of anaerobic oxidation of ammonium (anammox) bacteria overcame these issues (Thamdrup and Dalsgaard, 2002; Dalsgaard et al., 2003; Kuypers et al., 2003). The anammox phenomenon was first observed in the denitrifying fluidized-bed reactor (Mulder et al., 1995), and anammox bacteria can oxidize ammonium to nitrogen gas ( $N_2$ ) under anoxic conditions using nitrite as the electron acceptor. The autotrophic and anaerobic properties of

microorganisms save a great deal of energy and organic carbon source consumption. The anammox bacteria are affiliated with phylum Planctomycetes (Jetten et al., 2010), and five genera of anammox bacteria have been reported, “*Candidatus Brocadia*”, “*Candidatus Kuenenia*”, “*Candidatus Scalindua*”, “*Candidatus anammoxoglobus*” and “*Candidatus Jettenia*” (Schmid et al., 2005, 2007; Kartal et al., 2007, 2008).

In recent years, autotrophic denitrification processes based on anammox have been successfully applied in high-strength ammonia wastewater treatments (Jetten et al., 1997; van Dongen et al., 2001; López et al., 2008). It is very important for the start-up of the anammox process to choose suitable seed sludge for enrichment of anammox bacteria. Generally, the seed sludge

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67 derives from anaerobic sludge, nitrifying sludge or mixed sludge  
68 (Zhang et al., 2004; Zheng et al., 2004; Chen et al., 2011).

69 Anammox bacteria has been found in many ecosystems  
70 such as wastewater plant (Bae et al., 2010), river (Zhang et al.,  
Q8 2007; Wang et al., 2012), lake (Schubert et al., 2006), constructed  
Q9 wetland (Zhu et al., 2011a, 2011b), marine (Dalsgaard et al.,  
73 2003), paddy soil, and other natural habitats with extremely  
74 cold condition and poor nutrition (Jiang et al., 2015). It shows not  
75 only that these bacteria are widely distributed in the natural  
76 environment, but also that the anammox process has great  
Q10 contribution to the global nitrogen cycle flux. Previous re-  
78 searchers indicated that anammox reaction contributed 50–70%  
79 to N<sub>2</sub> production in marine oxygen and 4–37% to N<sub>2</sub> production  
80 in paddy soil (Dalsgaard et al., 2003; Kuypers et al., 2003, 2005;  
81 Arrigo, 2005; Lam et al., 2007; Zhu et al., 2011a, 2011b).

82 Rice paddy fields make a great contribution to the nitrogen  
83 cycle in terrestrial ecosystems. The long-term fertilization of  
84 paddy soil has a high concentration of nitrogen and the soil  
85 surface is flooded seasonally, which provides a suitable growth  
86 environment for anammox bacteria. The previous report has  
87 shown that the abundance of anammox bacteria in the  
88 40–50 cm depth of paddy soil reaches to  $1.2 \times 10^7$  copies/g dry  
89 soil (Zhu et al., 2011a, 2011b). Present researches have focused  
90 on the distribution of anammox bacteria in paddy fields.  
91 However, few studies have involved the start-up of anammox  
92 process with the sludge in paddy soil and discussed the  
93 evolution of bacteria during this process.

94 Hence, the present study aimed at investigating the dynamic  
95 changes of key microbial community structure during the  
96 start-up of anammox process with paddy soil as seed sludge,  
97 and finally successfully starts up an anammox process for  
98 nitrogen removal from wastewater.

## 100 1. Materials and methods

### 101 1.1. Seed sludge

102 A long-term fertilization paddy field soil as the sampling site  
103 in Songjiang District of Shanghai in China was selected. The  
104 depth of the collected samples was 0 to 80 cm, and the

105 samples were placed in a sterile plastic bag and brought back  
106 to the lab under low temperature conditions. One part of  
107 paddy soil samples was inoculated in the sequencing batch  
108 reactor (SBR) to start up the anammox process, and another  
109 part was stored at  $-80^{\circ}\text{C}$  for the subsequent analysis.

### 110 1.2. The SBR-anammox system

111 The paddy soil was inoculated in an 8-L SBR reactor (effective  
112 volume, 6 L) for anammox system in this study (Fig. 1). The  
113 reactor was equipped with on-line detection devices for  
114 detecting some important parameters, such as temperature,  
115 pH, dissolved oxygen and oxidation reduction potential. The  
116 formulated wastewater was introduced from the bottom of  
117 the reactor via a peristaltic pump (BT100-2J, Longer, China).  
118 The stirring rate was about 65 r/min. The temperature was  
119 controlled at  $35^{\circ}\text{C}$  using a temperature controlling belt. The  
120 anammox process in the SBR was operated with 1-day cycle.  
121 Each cycle was composed of four phases: nitrogen aeration  
122 (20 min), continuous feeding period (20 min), anaerobic reac-  
123 tion period (20 hr), settling (3 hr) and withdrawal (20 min).  
124 The nitrogen aeration in the beginning of each cycle was to  
125 remove oxygen from the SBR-anammox reactor system. The  
126 pH in the influent was kept between 7.5 and 8.3. The entire  
127 reaction device is wrapped with aluminum foil to achieve the  
128 dark effect. The initial sludge concentration was 6000 mg/L.

### 129 1.3. Synthetic wastewater

130 The influent concentrations of  $\text{NH}_4\text{Cl}$  and  $\text{NaNO}_2$  were  
131 gradually increased during the start-up of the anammox  
132 system, which the concentration of  $\text{NH}_4\text{Cl}$  increased from 15  
133 to 60 mg/L, and the concentration of  $\text{NaNO}_2$  gradually  
134 increased from 15 to 78 mg/L. The composition of the mineral  
135 medium was (g/L):  $\text{NaHCO}_3$ : 1.85;  $\text{KH}_2\text{PO}_4$ : 0.00625;  $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$ :  
136 0.018;  $\text{EDTA} \cdot 2\text{H}_2\text{O}$ : 0.0125;  $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ : 0.2;  $\text{CaCl}_2 \cdot 2\text{H}_2\text{O}$ : 0.3;  
137 and 1 mL/L of trace element solution. The trace element  
138 solution contained (g/L):  $\text{MnCl}_2 \cdot 4\text{H}_2\text{O}$ , 0.99; EDTA, 15;  $\text{H}_3\text{BO}_4$ ,  
139 0.014;  $\text{ZnSO}_4 \cdot 7\text{H}_2\text{O}$ , 0.43;  $\text{CoCl}_2 \cdot 6\text{H}_2\text{O}$ , 0.24;  $\text{NaMoO}_4 \cdot 2\text{H}_2\text{O}$ ,  
140 0.22;  $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$ , 0.25;  $\text{NiCl}_2 \cdot 6\text{H}_2\text{O}$ , 0.19;  $\text{NaSeO}_4 \cdot 10\text{H}_2\text{O}$ ,  
141 0.21; and  $\text{NaWO}_4 \cdot 2\text{H}_2\text{O}$ , 0.05 (Van de Graaf et al., 1996).

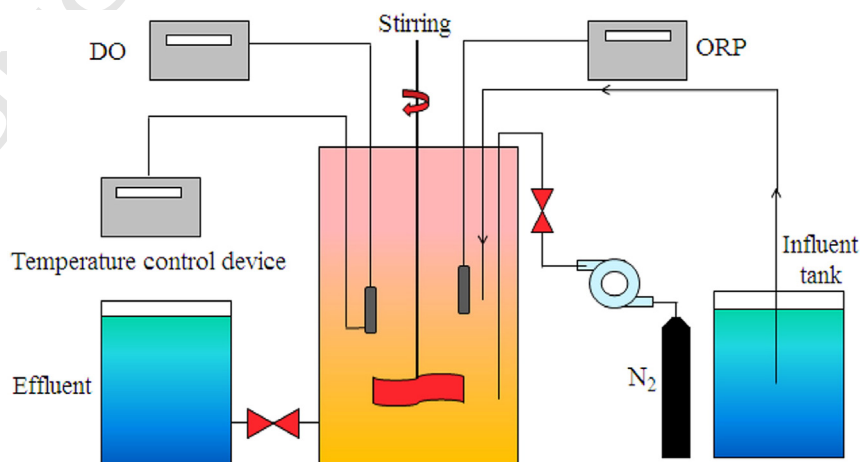


Fig. 1 – Schematic diagram of the reactor configuration. DO: dissolved oxygen; ORP: oxidation-reduction potential.

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