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Advanced Powder Technology xxx (2018) xxx-xxx

Contents lists available at ScienceDirect

# Advanced Powder Technology

journal homepage: www.elsevier.com/locate/apt

## Original Research Paper

# Development of cerium and silicon co-doped hydroxyapatite nanopowder and its *in vitro* biological studies for bone regeneration applications

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#### ARTICLE INFO

Article history:
Received 24 February 2018
Received in revised form 25 July 2018
Accepted 31 July 2018
Available online xxxx

21 Keywords:

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3 5

- 22 Ce-HAP
- 23 Ce/Si-HAP 24 Erythrocyte
- 24 Erythrocytes25 Bioactivity
- 26 MG-63 Osteoblast

#### ABSTRACT

Multi-ion, co-substituted bioactive glass ceramics play a significant role in the stimulation of physical and biological properties for outstanding effects in biomedical application. The following work attempts to develop HAP as a parent material doped with a combination of cerium ( $Ce^{4+}$  @1.25 wt%) and silicon ( $Si^{4+}$  @1, 3 and 5 wt%) by refluxing based sol-gel technique. The anti-bacterial tests exhibit *E. coli* showing higher inhibition efficiency, *in vitro* hemolytic test exhibit good compatible nature of dual doped HAP with erythrocytes (<5% of hemolytic). *In vitro* bioactivity assay confirms that the developed dual doped HAP possesses excellent bone-like apatite layer formation on their surfaces. *In vitro* cellular study was performed for Ce/Si-HAP@5% powder against MG-63 cells, which demonstrated the good cell viability at higher concentrations (up to 800 µg/ml). Further, dual doped HAP powders were characterized by various analytical techniques such as ATR-FTIR, Powder-XRD, TGA-DTA, SEM-EDS, TEM and XPS analysis. The studies confirm that the synthesized dual doped HAP will act as better bioactive glass ceramics for potential orthopedic and dentistry applications.

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### 46 **1. Introduction**

Natural bone is a nanocomposite material with the combination 47 of inorganic apatite and organic collagen which is embedded into 48 the extracellular matrix (ECM) for rigid structural maintenance of 49 bone [1]. The biological apatite consists of the major amount of cal-50 cium phosphate and a minor amount of anionic, cationic metal ions 51 such as Cl<sup>-</sup>, F<sup>-</sup>, SiO<sub>4</sub><sup>4-</sup>, CO<sub>3</sub><sup>2-</sup>, Mg<sup>2+</sup>, Na<sup>+</sup>, Al<sup>3+</sup>, K<sup>+</sup>, Zn<sup>2+,</sup> and Sr<sup>2+</sup>, etc. 52 53 respectively [2,3]. Nowadays, synthetic hydroxyapatite [HAP:  $Ca_{10}(PO_4)_6(OH)_2$  is widely used as an alternative bone graft in var-54 ious clinical surgeries due to its physical and chemical structural 55 similarities with natural apatite mineral [4,5]. HAP has extraordi-56 nary properties such as biocompatibility, non-toxicity, osteoinduc-57 tion, osteogenesis, osteointegrity and direct active chemical bond 58 formation with hard tissues [6]. However, pure phase synthetic 59 60 HAP exhibits lower biomineralization when compared with the 61 other bioactive glass ceramics due to rather minimal bioresorption 62 properties, lack of microbial restriction behaviour and lower frac-63 ture toughness with higher modulus strength [7,8]. These chal-64 lenges can be overcome by the doping of bioactive metal ions in

pure HAP crystal structure to improve the biological and mechanical properties for outstanding clinical applications [9]. In HAP crystal lattice, Ca<sup>2+</sup> ions can be substituted or doped by various biofunctional metal ions such as Ag<sup>+</sup>, Cu<sup>2+</sup>, Zn<sup>2+</sup>, Ti<sup>4+</sup>, Al<sup>3+</sup>, Mg<sup>2+</sup>, Na<sup>+</sup>, Co<sup>2+</sup>, Fe<sup>2/3+</sup>, Si<sup>4+</sup>, Sr<sup>2+</sup> and Ce<sup>3+/4+</sup>, etc. [10,11]. However, these ions lead to changes in the physical properties of pure HAP, in terms of solubility, thermal stability, surface charge with surface area, the degree of crystallinity and slight modification of lattice parameters [12,13]. In recent years, many researchers have been focusing on the development of a customized approach to tailor the characteristics of synthetic bioceramics with the doping of dual bioactive metal ions. Hence, these metal ions play a crucial role in the enhancement of biological properties of pure HAP for improved clinical applications [14].

For the past few decades, pathogen restriction behaviour of pure HAP has been created by the addition of transition and rare-earth elements [15,16]. Nevertheless, among these elements, Cerium (Ce) was selected to include into the crystal lattice of HAP due to its similar nature as Calcium ions and also accumulation in a natural bone at the lower level of concentrations [17,18]. Generally, Ce has been widely used as an antibacterial agent for long-term medical applications owing to its thermal stability, low cost and excellent antimicrobial efficacy [19]. The

https://doi.org/10.1016/j.apt.2018.07.028

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Please cite this article in press as: B. Priyadarshini, U. Vijayalakshmi, Development of cerium and silicon co-doped hydroxyapatite nanopowder and its *in vitro* biological studies for bone regeneration applications, Advanced Powder Technology (2018), https://doi.org/10.1016/j.apt.2018.07.028

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88 ceramic materials with Ce ions were tremendously used in cancer 89 treatments, Alzheimer diseases, drug delivery, dental fillings, 90 catheters, and burn wound healings [20]. It is well known that 91 Ce exists as +3 and +4 oxidation states because of its partial occupancy of 4f and 5d subshells of electrons and lead to excitation 92 state by a change in the chemical environment [21]. Yingguang 93 et al. have evaluated the antibacterial activity of Ce<sup>3+</sup> substituted 94 HAP using S. aureus, E. coli, and L. casei pathogens and found the 95 excellent inhibition efficiency by the release of Ce<sup>3+</sup> ions [22]. Cio-96 banu et al. have synthesized Ce4+ substituted HAP using co-97 98 precipitation method and examined its antibacterial activity against E. coli and S. aureus [23]. Gopi et al. have developed the 99 Ce<sup>3+</sup>/Eu<sup>3+</sup> dual substituted HAP coatings on 316L SS by electrode-100 position and studied the antibacterial activity, cytotoxicity, and 101 102 anti-corrosion property. These studies have proved that coated 103 implants play a vital role in biomedical applications [24]. Sanval 104 et al. employed the sol-gel technique to synthesize  $Zr^{4+}/Ce^{3+}$  co substituted HAP and fluorohydroxyapatite also evaluated its struc-105 tural formation and bacterial restriction behaviour for clinical 106 applications [25]. Gopi et al. prepared Sr<sup>2+</sup>/Ce<sup>3+</sup> co substituted 107 108 HAP by microwave irradiation technique and confirmed the 109 enhancement of bioactivity and antibacterial activity for better tissue engineering applications [26]. 110

Silicon (Si) is the important trace element that plays a crucial 111 112 role in the osteoblast differentiation by increasing metabolic activ-113 ity of human osteoblast cells and is also involved in the develop-114 ment of connective tissues with the production of collagen type I 115 [27,28]. Si has the ability to enhance the bioactivity of biological apatite and accelerates the bio-mineralization of natural bone 116 117 and cartilage to repair the injuries in human bone [29]. Incorpora-118 tion of Si ions into HAP crystal lattice improves the physicochemical properties of HAP with enhancement of the proteins-cell 119 120 interactions to improve the bone regeneration and osteointegra-121 tion properties [30]. It has been reported that Si-HAP coatings 122 increase the biological activity and human osteoblast cell adhesion 123 [31]. Gibson et al. synthesized Si-substituted HAP and examined its 124 cellular response which demonstrated the prominent activity of 125 osteoblast cells on Si-HAP surface [32]. Bang et al. employed the 126 precipitation method to prepare the Si-HAP with different concen-127 trations such as 0.4, 0.8 and 1.6 wt% and found slight changes in the crystal structure of HAP by incorporation of Si in terms of 128 increase in the lattice parameters with reduction of grain size 129 [33]. Friederichs et al. have synthesized Zn and  $SiO_4^{4-}$  co substi-130 131 tuted HAP using wet precipitation method and performed the experimental chemistry, atomic remodeling studies to investigate 132 133 the presence of Zn and Si atoms [34]. Jianyong et al. have devel-134 oped the individual and combination of Si and Sr co-doped HAP 135 by the adoption of the hydrothermal method and found to increase 136 in the biological activity of osteoblast cells when compared with 137 the undoped HAP [35]. Kim et al. reported Si and Mg co-138 substituted HAP using the precipitation method and observed as single phase HAP formation with improved biocompatibility beha-139 viour for implantation and bone augmentation applications [36]. 140 Zhang et al. studied the preparation of Si and Sr co-substituted 141 142 HAP nanowires using Sr containing calcium silicate and trisodium phosphate sources by hydrothermal treatment and proposed them 143 as multifunctional bioactive ceramics towards regeneration of hard 144 tissue applications [37]. Hence, these findings suggest that dual ion 145 incorporated HAP can act as a promising biomaterial for biomedi-146 147 cal applications.

Therefore, in the present investigation, we have attempted to synthesize 1.25% of Ce along with 1, 3, and 5% of Si co-doped HAP by sol-gel method. The Ce and Si co-doped HAP was characterized by ATR-FTIR, XRD, TGA/DTA, SEM-EDS, HR-TEM and XPS analysis. In addition to this, we have performed the *in vitro* biological studies such as antibacterial activity, hemocompatibility, bioactivity and cytocompatibility to prove them as the suitable 154 bioactive glass material for bone and dental applications. 155

#### 2. Materials and method

Ceric (IV) ammonium nitrate  $[(NH_4)_2 \text{ Ce } (NO_3)_6\text{-SDFCL }98.5\%],$ 157Calcium nitrate tetrahydrate  $[Ca(NO_3)_2\text{-}4H_2O\text{-}SDFCL 99\%],$  Triethyl158phosphite  $[P(OC_2H_5)_3\text{-}Sigma \text{ Aldrich }98\%],$  Tetraethyl orthosilicate159 $(Si(OC_2H_5)_4 \text{ Sigma Aldrich }98\%),$  Double distilled water  $[DD \cdot H_2O],$ 160Phosphate buffer solution (PBS) and Dimethyl sulfoxide (DMSO).161

#### 2.1. Synthesis of Ce and Si-doped HAP

Based on the previous report, we found that 1.25% of Ce doped 163 HAP exhibits excellent antibacterial activity, hemocompatibility 164 and cytocompatibility properties [38]. Hence, the following work 165 attempts to synthesize 1.25% of Ce doped HAP co-substituted with 166 Silica using different concentrations such as 1, 3 and 5% by sol-gel 167 method via refluxing process. Firstly, we have maintained the Ca 168 (0.9875 M) to Ce (0.0125 M) ratio as "1 M" and mixed these 169 sources in DD H<sub>2</sub>O to obtain the clear solution. On the hand, the 170 ratios of P to Si were maintained as "0.6 M" and precursors were 171 dissolved in DD H<sub>2</sub>O. The mixture of Ca/Ce solution was added 172 dropwise to the P/Si solution and stirred for 1 h followed by aging 173 at room temperature for 24 h. Further, the aged solution was 174 refluxed at 85 °C for 16 h to induce the interaction between the 175 ions. After refluxing process, the mixture of the solution was kept 176 for evaporation at 80 °C for 8 h to obtain the gel formation by poly-177 condensation. The formed gel was dried in an oven for 8 h at 100 °C 178 followed by sintering at 900 °C for 2 h. In addition, Si-doped HAP 179 was synthesized by following the above-mentioned procedure 180 using various concentrations of Si such as 1, 3 and 5% respectively. 181 Further, heat treated pure phase and dual doped Ce/Si-doped HAP 182 powders were characterized by various techniques. Hereafter, 1, 3 183 and 5% of Si content incorporation into Ce doped HAP are men-184 tioned as Ce/Si-HAP@1%, Ce/Si-HAP@3% and Ce/Si-HAP@5% 185 respectively. 186

#### 2.2. In vitro hemocompatibility

In vitro hemocompatibility study is done to evaluate the bio-188 compatibility of prepared material and examined to evaluate the 189 blood compatible behaviour with 1.25% of Ce-HAP, 5% of Si-HAP, 190 Ce/Si-HAP@1%, Ce/Si-HAP@3%, and Ce/Si-HAP@5% powders. To 191 perform the hemocompatibility assay, we collected 5 ml of fresh 192 human blood from healthy volunteers with age of 26 years (Health 193 Center, VIT University: Ref. No. VIT/IECH/01/Aug.2016) and trans-194 ferred to the falcon tube containing an anticoagulant agent. The 195 obtained blood sample was centrifuged at 4 °C for 2 min using 196 10,000 rpm to separate the plasma from erythrocytes. After separa-197 tion of erythrocytes from plasma, for three repetitive times, it was 198 diluted with 5 ml of PBS solution followed by centrifugation for 2 199 min at 10,000. The settled erythrocytes were mixed with 15 ml of 200 freshly prepared PBS solution and kept in the refrigerator. From 15 201 ml of the blood sample, 0.2 ml was taken and separately mixed 202 with 5 mg of Ce-HAP, Si-HAP, and Ce/Si-HAP powders which were 203 dispersed in 0.8 ml of PBS solution. On the other hand, 0.2 ml of 204 blood with 0.8 ml of DD H<sub>2</sub>O was treated as a positive control 205 whereas with 0.8 ml of PBS acts as negative control respectively. 206 These mixtures of blood samples were incubated for 1 h at room 207 temperature in shaking conditions followed by centrifugation for 208 3 min using 10,000 rpm. After this step, the obtained supernatant 209 was used for readings at 570/655 nm by Biorad Elisa. 210

The following formula was used to determine the hemolytic ratio of Ce-HAP, Si-HAP, and Ce/Si-HAP powders.

Please cite this article in press as: B. Priyadarshini, U. Vijayalakshmi, Development of cerium and silicon co-doped hydroxyapatite nanopowder and its *in vitro* biological studies for bone regeneration applications, Advanced Powder Technology (2018), https://doi.org/10.1016/j.apt.2018.07.028

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