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At what stage in metazoan evolution did leukotriene generation first appear?-key insights from cartilaginous fish

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Abstract

Leukotriene (LT) B₄ is a key player in inflammatory responses in mammals. During the generation of this derivative of arachidonic acid, the unstable product of 5-lipoxygenase, termed LTA₄, is converted to LTB₄ by LTA₄ hydrolase. Invertebrates do not generate LTs yet all vertebrates from bony fish onwards synthesize this compound. As cartilaginous fish are the most primitive living jawed vertebrates, we investigated if the leukocytes from such a fish, the Thornback ray (Raja clavata) could generate LTB₄. Supernatants from ionophore-challenged leukocytes generated the 5-lipoxygenase products, 6-trans-LTB₄ and 6-trans-12-epi-LTB₄ but were unable to synthesize LTB₄. To determine if these cells contained an active LTA₄ hydrolase, LTA₄ was incubated with lysates from ray leukocytes. Such preparations did not contain any demonstrable LTA₄ hydrolase activity. Our findings imply at the stage of cartilaginous fish evolution over 350 million years ago that the evolution of an active LTA₄ hydrolase had yet to occur.

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Keywords: Ray; Leukotriene A hydrolase; Lipid mediators; Inflammation

1. Introduction

Leukotrienes (LT) in particular LTB₄, are key pro-inflammatory molecules that have been shown to be implicated in many disease processes including asthma and rheumatoid arthritis [1]. During the biosynthesis of LTB₄, the C20 polyunsaturated fatty acid, arachidonic acid, is converted by 5-lipoxygenase (5-LO) to the unstable epoxide termed LTA₄, which in turn is subject to enzymatic hydrolysis by LTA₄ hydrolase (EC 3.3.2.6; Fig. 1). In the absence of LTA₄ hydrolase activity, the unstable LTA₄ ($t_{1/2} < 10$ s in physiological conditions) undergoes rapid non-enzymatic hydrolysis to give rise to a number of di-hydroxyeicosatetraenoic acids (di-HETEs) including 6-trans-LTB₄, 6-trans-12-epi LTB₄ and various 5,6 diHETEs (Fig. 1). These non-enzymatic products are largely without the potent biological activity of LTB₄ [2]. LTA₄ hydrolase is a 69 kDa bi-functional, zinc metalloenzyme with both aminopeptidase and LTA₄ hydrolase activities [3-5]. It is widely distributed in mammalian tissues [6] and has been found in all leukocyte types as well as mature erythrocytes [3,7].

Abbreviations: HETE, hydroxyeicosatetraenoic acid; LO, lipoxygenase; LT, leukotriene; PG, prostaglandin; RP-HPLC, reverse phase high performance liquid chromatography.

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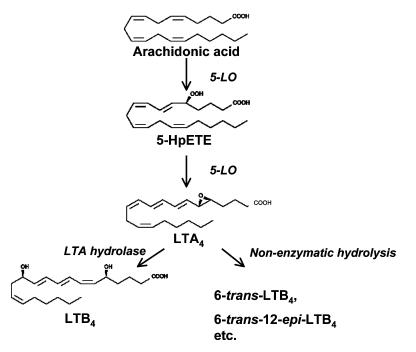


Fig. 1. Schematic diagram showing the major biosynthetic routes of leukotriene (LT) and dihydroxyeicosatetraenoic acid generation from the unstable epoxide, LTA₄. 5-HpETE, 5 hydroperoxyeicosatetraenoic acid; LO, lipoxygenase.

Although invertebrates exhibit potent inflammatory reactions, there is no convincing evidence for the generation of LTs in these animals [8]. This raises the question about the evolution of both 5-LO and LTA₄ hydrolase, and at what stage their products first appeared. Recently, an aminopeptidase has been cloned from the nematode worm, Caenorhabditis elegans that although showing strong homology to human LTA4 hydrolase is unable to convert LTA_4 to LTB_4 [9]. One interpretation of this finding is that in 'lower' animals, such as C. elegans, this enzyme originally functioned as an aminopeptidase and, at a later stage in evolution of more complex multicellular animals, the lipid binding site became altered such that the active site could bind LTA₄ and catalyse the biosynthesis of LTB₄ [5]. As leukocytes from bony fish and amphibians generate LTB₄ [10,11] and such animals have been shown to have an active LTA₄ hydrolase [12,13], this implies that the evolution of LT generation probably came about early in vertebrate evolution with the appearance of bony fish. This current study determines whether cartilaginous fish (sharks, rays, etc.) that represent the ancestors of some of the most primitive jawed vertebrates, also generate LTB_4 and express functional LTA_4 hydrolase activity. This was achieved by examining the LT generating capacity of leukocytes from the hemopoietic Leydig organ located in the mucosa of the oesophagus of the Thornback ray, *Raja clavata*.

2. Materials and methods

2.1. Fish

Thornback rays, *Raja clavata*, (25–58 cm length) were collected by trawling from the *R. V. Noctiluca* around the Gower Peninsula, South Wales and either used immediately or maintained in an aquarium overnight. Rays were not fed during this time and were terminally anaesthetised by placing in 3-aminobenzoic acid ethyl ester methanesulfonate salt (MS-222, 0.1 g ml⁻¹, Sigma Aldrich, UK).

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