REFERENCES

- Fridkin SK, Steward CD, Edwards JR et al. Surveillance of antimicrobial use and antimicrobial resistance in United States hospitals: project ICARE phase 2. Project Intensive Care Antimicrobial Resistance Epidemiology (ICARE) hospitals. Clin Infect Dis 1999; 29: 245–252.
- Fluit AC, Verhoef J, Schmitz FJ et al. Frequency of isolation and antimicrobial resistance of gram-negative and grampositive bacteria from patients in intensive care units of 25 European university hospitals participating in the European arm of the SENTRY Antimicrobial Surveillance Program 1997–1998. Eur J Clin Microbiol Infect Dis 2001; 20: 617–625.
- 3. Hsueh PR, Chen ML, Sun CC *et al.* Antimicrobial drug resistance in pathogens causing nosocomial infections at a university hospital in Taiwan, 1981–1999. *Emerg Infect Dis* 2002; **8**: 63–68.
- Jones RN, Kirby JT, Beach ML et al. Geographic variations in activity of broad-spectrum beta-lactams against Pseudomonas aeruginosa: summary of the worldwide SENTRY Antimicrobial Surveillance Program (1997–2000). Diagn Microbiol Infect Dis 2002; 43: 239–243.
- Gales AC, Jones RN, Turnidge J et al. Characterization of Pseudomonas aeruginosa isolates: occurrence rates, antimicrobial susceptibility patterns, and molecular typing in the global SENTRY Antimicrobial Surveillance Program, 1997–1999. Clin Infect Dis 2001; 32(suppl 2): S146–S155.
- Nordmann P, Poirel L. Emerging carbapenemases in gramnegative aerobes. Clin Microbiol Infect 2002; 8: 321–331.
- Harris AD, Smith D, Johnson JA et al. Risk factors for imipenem-resistant Pseudomonas aeruginosa among hospitalized patients. Clin Infect Dis 2001; 34: 340–345.
- 8. Gulay Z, Atay T, Amyes SG. Clonal spread of imipenemresistant *Pseudomonas aeruginosa* in the intensive care unit of a Turkish hospital. *J Chemother* 2001; **13**: 546–554.
- Sorin M, Segal-Maurer S, Mariano N et al. Nosocomial transmission of imipenem-resistant Pseudomonas aeruginosa following bronchoscopy associated with improper connection to the Steris System 1 processor. Infect Cont Hosp Epidemiol 2001; 22: 409–413.
- Hsueh PR, Teng LJ, Yang PC et al. Persistence of a multidrug-resistant Pseudomonas aeruginosa clone in an intensive care burn unit. J Clin Microbiol 1998; 36: 1347–1351.
- National Committee for Clinical Laboratory Standards. Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically, 5th edn. Approved standard M7-A4. Wayne, PA: NCCLS, 2000.
- National Committee for Clinical Laboratory Standards. Performance standards for antimicrobial susceptibility testing, twelfth informational supplement. M100–S14. Wayne, PA: NCCLS, 2004.
- Yan JJ, Hsueh PR, Ko WC et al. Metallo-β-lactamases among clinical isolates of *Pseudomonas* in Taiwan and identification of VIM-3, a novel variant of the VIM-2 enzyme. *Antimicrob Agents Chemother* 2001; 45: 2224– 2228.
- 14. Lee K, Lee WG, Uh Y *et al.* VIM- and IMP-type metalloβ-lactamase-producing *Pseudomonas* spp. and *Acinetobacter* spp. in Korean hospitals. *Emerg Infect Dis* 2003; **9**: 868–870.
- 15. Maguire AJ, Brown DF, Gray JJ et al. Rapid screening technique for class 1 integrons in Enterobacteriaceae and nonfermenting gram-negative bacteria and its use in

- molecular epidemiology. Antimicrob Agents Chemother 2001; 45: 1022–1029.
- Sardelic S, Pallecchi L, Punda-Polic V et al. Carbapenemresistant Pseudomonas aeruginosa carrying VIM-2 metalloβ-lactamase determinants, Croatia. Emerg Infect Dis 2003; 9: 1022–1023.
- 17. Hsueh PR, Teng LJ, Chen CY *et al*. Pandrug-resistant *Acinetobacter baumannii* causing nosocomial infections in a university hospital, Taiwan. *Emerg Infect Dis* 2002; **8**: 827–832
- 18. Hirakata Y, Yamaguchi T, Nakano M *et al.* Clinical and bacteriological characteristics of IMP-type metallo-β-lactamase-producing *Pseudomonas aeruginosa. Clin Infect Dis* 2003; **37**: 26–32.
- Bantar C, Di Chiara M, Nicola F, Relloso S, Smayevsky J. Comparative in vitro bactericidal activity between cefepime and ceftazidime, alone and associated with amikacin, against carbapenem-resistant *Pseudomonas* aeruginosa strains. Diagn Microbiol Infect Dis 2000; 37: 41– 44.
- Li J, Nation RL, Milne RW, Turnidge JD, Coulthard K. Evaluation of colistin as an agent against multi-resistant Gram-negative bacteria. *Int J Antimicrob Agents* 2005; 25: 11–25.

RESEARCH NOTE

The relationship between serotypes and PFGE genotypes in isolates of *Streptococcus* pneumoniae from Hungary

O. Dobay^{1,2}, F. Rozgonyi², E. Hajdú³, E. Nagy³, M. Knausz⁴ and S. G. B. Amyes¹

¹Medical Microbiology, Medical School, University of Edinburgh, UK, ²Institute of Medical Microbiology, Semmelweis University, Budapest, ³Department of Clinical Microbiology, Faculty of Medicine, University of Szeged, Szeged and ⁴Aladár Petz County Teaching Hospital, Györ, Hungary

ABSTRACT

The relatedness of 112 penicillin-non-susceptible isolates of *Streptococcus pneumoniae* from Hungary was determined by pulsed-field gel electrophor-

Corresponding author and reprint requests: S. G. B. Amyes, University of Edinburgh, Medical School, Medical Microbiology, Teviot Place, Edinburgh EH8 9AG, UK E-mail: S.G.B.Amyes@ed.ac.uk esis (PFGE), serotyping and antibiotic susceptibility tests. The differences in PFGE patterns closely mirrored the changes in resistance. Some genotypes comprised multiple serotypes, and the genetic diversity among certain serotypes was considerable. Generally, serotyping was insufficient for epidemiological mapping of pneumococcal isolates. There was considerable serotype diversity, but the five most frequent international serotypes (6, 9, 14, 23, 19) were the most prevalent. In addition, the presence of some well-defined resistant international pneumococcal clones in the Hungarian population was identified.

Keywords Genotypes, Hungary, penicillin-non-susceptible, pneumococci, serotypes, *Streptococcus pneumoniae*

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Streptococcus pneumoniae causes a wide range of serious diseases, and it is essential to determine the epidemiological and genetic relatedness of isolates at the local, national and international levels to control their spread and to develop new vaccines [1]. Phenotypic and genotypic typing methods can be used for the characterisation and epidemiological tracking of these bacteria. The present study used pulsed-field gel electrophoresis (PFGE) for molecular characterisation of penicillin-non-susceptible Hungarian isolates of pneumococci, and then compared the genotypic PFGE results with phenotypic results based on serotypes and antibiotic resistance patterns.

Penicillin-non-susceptible, non-invasive S. pneumoniae isolates (n = 112), obtained from five clinical laboratories in Hungary during 2000-2002 [2], were included in the study. The identities of all isolates were also confirmed by *lytA* PCR [2,3]. Antibiotic susceptibility testing was performed by the agar dilution method on Mueller–Hinton (Oxoid, Basingstoke, UK) blood agar plates. The sensitivity and resistance breakpoints used were, where available, those recommended by the 2002 NCCLS guidelines [4]. For telithromycin, the following breakpoints were used: susceptible (S), $\leq 0.5 \text{ mg/L}$; resistant ≥2 mg/L [5,6]. All antibiotics were purchased from Sigma (Poole, UK), except moxifloxacin (Bayer, Leverkusen, Germany) and telithromycin (Aventis Pharma, Bridgewater, NJ, USA).

Serotyping was performed with *S. pneumoniae* typing antisera (Mast Group, Bootle, UK). Chromosomal DNA for PFGE was prepared as described by Hall *et al.* [7], with slight modifications. After digestion with ApaI (Promega, Southampton, UK), the fragments were separated in a CHEF-DR II apparatus (Bio-Rad, Hercules, CA, USA), with pulse times of 2–30 s. The NO34OS λ ladder (New England Biolabs, Hitchin, UK) was used as a molecular size marker. The PFGE profiles were analysed with the BioNumerics program v. 2.5 (Applied Maths, Sint-Martens-Latem, Belgium). A PFGE genotype was defined as isolates that showed $\geq 90\%$ identity in the dendrogram created by the UPGMA/Dice coefficients with a band position tolerance of 2%, in accordance with published interpretive criteria [1]. The presence of the *erm* and *mef* macrolide resistance determinants, when relevant for the international comparison, was tested by PCR with the primers described by Sutcliffe et al. [8]. The mef(E) and mef(A) determinants were distinguished with the BamHI restriction method of Oster *et al.* [9].

The penicillin MICs for the isolates ranged from 0.125 to 16 mg/L, but there were only eight penicillin-resistant isolates (MIC \geq 2 mg/L). The frequency of macrolide resistance (MIC \geq 1 mg/L) was 50–54%. All isolates were sensitive to telithromycin, moxifloxacin, vancomycin and linezolid.

The correlation between the serotypes and antibiotic susceptibilities of these isolates has been reported previously [2], but it was important to establish the genetic stability of individual serotypes. Although there were 25 different PFGE genotypes, 70.5% of the isolates belonged to only five genotypes (Table 1). Twelve genotypes contained only one isolate each. While the other major genotypes comprised only one serogroup, the two largest genotypes, A and B, comprised two serogroups each (9/14 and 6A/23F, respectively), and the isolates were clearly sub-clustered according to the serotypes. Additionally, there was a close correlation between the genotypes and the penicillin and macrolide susceptibility levels.

Pneumococcal isolates from Hungary have rarely been serotyped routinely. Instead of the previously reported predominance of serotype

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