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MiniReview

Malassezia Baillon, emerging clinical yeasts

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Abstract

The human and animal pathogenic yeast genus *Malassezia* has received considerable attention in recent years from dermatologists, other clinicians, veterinarians and mycologists. Some points highlighted in this review include recent advances in the technological developments related to detection, identification, and classification of *Malassezia* species. The clinical association of *Malassezia* species with a number of mammalian dermatological diseases including dandruff, seborrhoeic dermatitis, pityriasis versicolor, psoriasis, folliculitis and otitis is also discussed.

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1. Introduction

Members of the genus *Malassezia* are opportunistic yeasts of increasing importance, due in large part to advances in detection and culture methodology which have both allowed their investigation and revealed their importance in human and animal disease [1,2]. The genus *Malassezia* belongs to the basidiomycetous yeasts and is classified in the *Malasseziales* (*Ustilaginomycetes*, *Basidiomycota*) [3–5]. The cells show a multilayered cell wall, enteroblastic budding (Fig. 1), urease activity, and

development of methods to differentiate them has

a positive staining reaction with Diazonium Blue B

⁽DBB) [3]. The genus was named in 1889 by Baillon [6] with the species *M. furfur*, to accommodate the filamentous fungus observed in scales of the human skin disease pityriasis versicolor (PV). *Pityrosporum* [7] has been proposed as an alternative generic name, but because *Malassezia* had been published earlier this name has nomenclatural priority. The genus remained limited to *M. furfur* and *M. pachydermatis* for a long time. *M. pachydermatis* is lipophilic but not lipid-dependent, and usually occurs on animals [8]. For many years all pathologies caused by *M. furfur sensu lato*, particularly disorders of the skin such as dandruff, seborrhoeic dermatitis (D/SD), pityriasis versicolor (PV), and folliculitis, were ascribed to a single species [9]. Only the recent recognition of a number of new species and the

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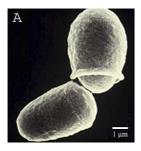




Fig. 1. Morphology of *Malassezia* cells. (A) Budding cells of *M. pachydermatis* viewed by scanning electron microscopy. (B) *M. furfur* under bright field microscopy.

changed this approach [2,10]. The 28S-rDNA sequences revealed seven distinct genetic entities [11], which are now widely accepted as species (M. furfur, M. obtusa, M. globosa, M. slooffiae, M. sympodialis, M. pachydermatis, and M. restricta) [10]. Since then, five new Malassezia species have been reported (M. dermatis [12], M. equi [13], M. japonica [14], M. nana [15], and M. yamatoensis [16]), but further biochemical and molecular characterization will be required for their acceptance as distinct species. Biochemical identification tests for Malassezia species include catalase and β -glucosidase activity, and evaluation of growth with cremophor EL and Tweens 20, 40, 60, 80, using the diffusion method in Sabouraud glucose agar (Table 1, Fig. 2) [1,4,10,17].

2. Malassezia species

M. furfur is morphologically heterogeneous with globose, oval or cylindrical yeast cells. This species can be identified by its ability to grow at 37 °C, strong catalase activity, absence or a very weak β-glucosidase activity, and equal growth in the presence of cremophor EL (=castor oil) and Tweens 20, 40, 60, 80 as sole lipid sources [4,17]. Strains of M. furfur showed two different karyotypes [18], but demonstrated high percentages of DNA/DNA reassociation and high ribosomal RNA similarity [10,11]. Some strains are able to produce filaments, either spontaneously or under particular culture conditions [19]. Strains of the species originate from various hosts, body sites and diseases. However, M. furfur was not observed in recent epidemiological surveys of healthy persons and patients with pityriasis versicolor (PV) and seborrhoeic dermatitis (SD) or with only PV [20,21]. This absence may, perhaps, be caused by the isolation protocols used, or may arise from competition between different skin-inhabiting species of Malassezia. Using the same isolation protocol, the species has been isolated from systemic and mucosal sites, such as urine, vagina and blood, or exposed sites such as nails (E. Guého, unpublished data). It has also been isolated from animals [22–24]. M. furfur is the only Malassezia species

Salient characteristics of Malassezia species [1,2,10,12,14-17,24,26-28,30-33,36,38-42,55,57,62,63,65,72,80,82-84,92,95,96,99-101]

Species	Occurrence	Cell morphology	Lipid	Tween 20	Tween 40	Tween 60	Tween 80	Cremophor	Catalase	β-Glucosidase	Tween 20 Tween 40 Tween 60 Tween 80 Cremophor Catalase β-Glucosidase Growth at 37 °C
			dependency					EL			
M. dermatis	AD	Ellipsoidal, globose	+	+	+	+	+	W, (+)	+	i	+
M. furfur	AD, AN, HS, OE,	Globose, ellipsoidal,	+	+, (-)	+, (-)	+, (-)	+,(-)	+, (-)	+, (-)	–, (w)	+
	PV low, S, SD low	cylindrical									
M. globosa	AN, AD, P, PV high,	Globose	+	I	_2	-2	I	I	+	1	–, (w)
	SD high, SD/D high										
M. japonica	AD, HS	Globose, ellipsoidal	+	1	W	+	1	٠	+		+
M. nana	AN, OE	Ellipsoidal	+	>	+	+	W	;	+	5	+
M. obtusa	AN, HS, OE, SD	Ellipsoidal, cylindrical	+	1	1	I	I	1	+	+	-, (w)
M. pachydermatis	AN, OE, S, SD/AN	Ellipsoidal	-, (w)	+	+	+	+	+	+, w	+, (-)	+
M. restricta	AD, HS scalp, P,	Globose, ellipsoidal	+	I	اء	اء	I	į	1		Λ
	SD, SD/D high										
M. slooffae	AN, HS, P, PV, SD	Ellipsoidal, cylindrical	+	+, w, (-)	+	+	–, (w)	1	+	1	+
M. sympodialis	AD, AN, HS, P, PV, SD Ellipsoidal	Ellipsoidal	+	$-, w^2 +$	+	+	+	–, (w)	+	+	+
M. vanatoensis	AD. HS. SD	Ellipsoidal	+	+	+	+	+	6	+	i	+

AD, atopic dermatitis, AN, non-human animals, HS, healthy human skin; OE, otitis externa; P, psoriasis; PV, pityriasis versicolor; S, sepsis; SD, seborrhoeic dermatitis; SD/AN, seborrhoeic dermatitis in animals; SD/D, seborrhoeic dermatitis/dandruff; SD; w, weak; v, variable; () indicate rare deviations from main pattern; 1, growth may be inhibited near the well where the substrate is

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