

Research report

Acquisition of ischemic tolerance by repetitive transcranial magnetic stimulation in the rat hippocampus

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Abstract

We investigated the acquisition of ischemic tolerance in the rat hippocampus using repetitive transcranial magnetic stimulation (rTMS). Rats received 1000 pulses/day for 7 days, and the field excitatory postsynaptic potentials were measured in the hippocampal CA1. After slices were exposed to ischemic conditions, long-term potentiation (LTP) was induced. The LTP of the stimulated group was enhanced compared with the LTP of the sham control group in each ischemic condition, suggesting that rTMS has the potential to protect hippocampal function from ischemia.

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Topic: Ischemia

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Transcranial magnetic stimulation (TMS) is a non-invasive technique that stimulates the brain using magnetically induced eddy currents through a coil positioned on the surface of the head [1,22]. Repetitive TMS (rTMS) has become an increasingly important therapeutic tool for the treatment of neurological and psychiatric disorders, such as depression and Parkinson's disease [13,15,21]. There is a possibility that rTMS might have therapeutic effects on ischemia; however, few studies have described the effects of rTMS on brain ischemia.

CA1 pyramidal neurons in the hippocampus are highly vulnerable to cerebral ischemia. Brief periods (minutes) of severe ischemia cause neuronal degeneration in the CA1 region 3 to 7 days after the ischemia by apoptosis (so-called "delayed neuronal death, DND") [12]. Notable increases in

extracellular glutamate concentration during cerebral ischemia, excessive excitation of neurons by the accumulation of glutamate, and Ca^{2+} influx during ischemia are thought to cause neuronal hyperactivity and trigger neuronal injury (so-called "excitotoxicity") [5].

Long-term potentiation (LTP) is a long-lasting increase in synaptic efficacy resulting from the high-frequency stimulation of afferent fibers [3,4]. LTP in the hippocampus is thought to be representative of the type of synaptic plasticity that is related to learning and memory [3,16]. There are many mechanisms associated with LTP induction; for example, the enhancement of transmitter release, the activation of α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA) and *N*-methyl-D-aspartate (NMDA) receptors, changes in the number of synaptic-spine contacts and alterations in the shape of the spine heads [2,3,14].

Previously, we reported that the effect of rTMS depends on the stimulus intensity and that rTMS administered at the appropriate intensity (0.75 T) enhanced LTP in area CA1 in the rat hippocampus. These data suggest that 0.75 T rTMS

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might activate hippocampal neurons [19,20]. It was also reported that rTMS protected neurons in the hippocampal CA1 from delayed neuronal death induced by transient ischemia [8]. Therefore, it is possible that TMS might prove to be a useful therapeutic treatment for ischemia. In this study, we investigated the ability of 0.75 T rTMS to protect the hippocampal function of the rat from ischemic injury.

All experimental procedures performed in this study were approved by the Animal Ethics Committee of the University of Tokyo. Experiments were performed on male Wistar rats (4 weeks old, weight = 60 to 80 g; Saitama Experimental Animals Supply, Saitama, Japan). Pairs of rats (one stimulated and one sham control) were housed in individual cages with free access to food and water at room temperature. Rats were magnetically stimulated using a round coil (inner diameter = 15 mm, outer diameter = 75 mm, thickness = 10 mm) positioned over the head of each rat. Conscious rats were held beneath the coil by the nape of the neck during the stimulation delivery. The stimulator (NIHON KOHDEN, Tokyo, Japan) delivered biphasic cosine current pulses for 238 μ s. The peak magnetic field was set to 0.75 T at the center of the coil (80% of the motor threshold), which induces an eddy current of approximately 9 A/m² maximum in the brain [20]. Ten 1-s trains of 25 pulses/s with a 1-s intertrain interval were applied to the rats four times per day for 7 days. The coil was cooled during the intertrain intervals. The sham control rats were treated with a sham coil (that is, non-stimulated) and exposed to the same noise as that produced during the stimulation. After rTMS for 7 days in vivo, ischemia experiments were performed in vitro using electrophysiological methods.

Approximately 15 h after the final stimulation, the rats were anesthetized with diethyl ether and decapitated. The brain was quickly removed from the skull and placed on an ice-cold filter paper dampened with artificial cerebrospinal fluid (ACSF) (composition (mM): NaCl 125, KCl 3, NaH₂PO₄ 1.2, NaHCO₃ 26, CaCl₂ 2.0, MgCl₂ 1.0 and glucose 10). The hippocampus was dissected and cut with a microslicer (D.S.K, Kyoto, Japan) into 400 μ m transverse slice sections. The slices were incubated and then allowed to recover in ACSF bubbled with 95% O₂/5% CO₂ (pH 7.4) at room temperature for a minimum of 1 h before recording. The slices were then transferred to a recording chamber and continuously perfused (approximately 2 ml/min) with ACSF at 30 °C. After stimulation of Schaffer collaterals with a tungsten bipolar stimulating electrode, field excitatory postsynaptic potentials (fEPSP) were recorded from the dendrites of CA1 pyramidal cells using a tungsten recording electrode. A single stimulus was administered at 20-s intervals. The stimulus intensity was set to generate an fEPSP with a slope that was approximately 30% of the maximum, as determined from the input–output curve. After obtaining stable fEPSP recordings for 20 min, the ACSF was replaced by ischemic ACSF (composition (mM): NaCl 125, KCl 3, NaH₂PO₄ 1.2, NaHCO₃ 26, CaCl₂ 2.0, MgCl₂

1.0 and sucrose 10) bubbled with 95% N₂/5% CO₂ (pH 7.4). Ischemic periods were set to 5, 10, 30, 40 and 50 min. After 5, 10, 30, 40 and 50 min, ischemic ACSF was replaced by normal ACSF and fEPSP recordings were continued. fEPSP peaks were continuously measured before and after ischemia. When fEPSPs did not recover after ischemia, the recording was stopped. We defined the viability of the slices as the percentage of the slices with fEPSPs that recovered after ischemia (the number of recovered slices/the total number of slices). When the fEPSP recovered and became stable after ischemia, LTP was induced by tetanus stimulation (100 Hz for 1 s; 0.1 ms duration). fEPSP recordings continued for 60 min after tetanus stimulation. LTP is achieved if the potentiation is maintained for at least 1 h. Pre-tetanus and post-tetanus fEPSPs were compared using their maximum leftward slopes, and the fEPSP slope data for each experiment were expressed as percentages of the pre-tetanus EPSP baseline average. Each slice was used for only one experiment and then discarded. Data were analyzed using pCLAMP software (Axon Instrument). LTP without ischemia was also induced for each rat. All LTP data for each group were averaged and expressed as the mean \pm standard error of the mean (S.E.M.). The average value of the maintenance phases of LTP (ranging from 10 min after tetanus stimulation to 60 min) were statistically analyzed by two-way ANOVA followed by post-hoc Dunnett's test. A probability (*P*) level of less than 0.05 was considered to be statistically significant.

Fig. 1 shows the viability of the slices after ischemia. The viabilities of the slices in the sham control groups after 5,

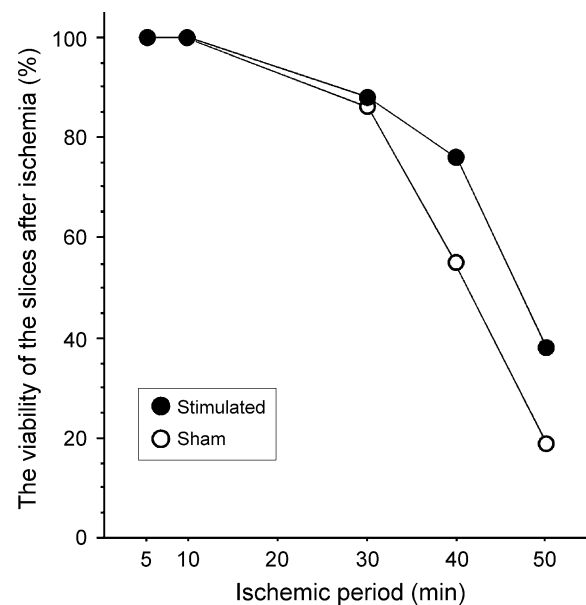


Fig. 1. The viability of the slices after ischemia (the number of recovered slices/the total number of slices). The viabilities in the sham control groups after 5, 10, 30, 40 and 50 min of ischemia were 100% (22/22), 100% (17/17), 86% (19/22), 55% (11/20) and 19% (3/16), respectively (open circle). The viabilities of the slices in the stimulated groups after 5, 10, 30, 40 and 50 min of ischemia were 100% (21/21), 100% (17/17), 88% (15/17), 76% (13/17) and 38% (5/13), respectively (closed circle).

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