

Molecular phylogeny of the genus *Pseudomonas* based on *rpoB* sequences and application for the identification of isolates

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Received 5 January 2005; accepted 25 February 2005

Available online 31 March 2005

Abstract

Phylogenetic relationships within the genus *Pseudomonas* were examined by comparing partial (about 1000 nucleotides) *rpoB* gene sequences. A total of 186 strains belonging to 75 species of *Pseudomonas* sensu stricto and related species were studied. The phylogenetic resolution of the *rpoB* tree was approximately three times higher than that of the *rrs* tree. Ribogroups published earlier correlated well with *rpoB* sequence clusters. The *rpoB* sequence database generated by this study was used for identification. A total of 89 isolates (79.5%) were identified to a named species, while 16 isolates (14.3%) corresponded to unnamed species, and 7 isolates (6.2%) had uncertain affiliation. *rpoB* sequencing is now being used for routine identification of *Pseudomonas* isolates in our laboratory.

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Keywords: *Pseudomonas*; Phylogeny; *rpoB*; Sequencing

1. Introduction

For over a century, the genus *Pseudomonas* Migula 1894 served as a repository for straight, strictly aerobic Gram-negative rods that were motile by one or several polar flagella. The nutritional studies of Stanier et al. [24] showed that the ability of *Pseudomonas* strains to utilize different compounds as sole source of carbon and energy provided essential phenotypic characterization. Groups of species differed considerably in their nutritional versatility.

Later, DNA–DNA [20] and rRNA–DNA hybridization [7,21] split the genus *Pseudomonas* into five groups called rRNA groups I to V.

Finally, sequencing 16S rRNA [28] or its gene (*rrs*) [27] supported the transfer of many species to other (often new) genera in the alpha subclass (e.g., *Aminobacter*, *Brevundimonas*, *Devosia*, *Oligotrophia*, *Sphingomonas*, *Zavarzinia*), beta subclass (e.g., *Acidovorax*, *Burkholderia*, *Comamonas*, *Hydrogenophaga*, *Ralstonia*, *Telluria*), gamma subclass (e.g., *Chryseomonas*, *Flavimonas*, *Pseudomonas* sensu stric-

to), or between the beta- and gamma-subclasses (e.g., *Stenotrophomonas*) of the Proteobacteria (reviewed by Kersters et al.) [13].

The genus *Pseudomonas* sensu stricto contains all species which corresponded to the RNA group I, i.e., fluorescent pseudomonads and related bacteria [19]. The list of species (145 when this paper was written) and subspecies of the genus *Pseudomonas* which have standing in nomenclature is available on the WorldWide Web (<http://www.bacterio.cict.fr/p/pseudomonas.html>).

Although the taxonomy of the genus *Pseudomonas* has progressed steadily thanks to rRNA sequencing (for gross phylogeny) and DNA–DNA hybridization (as species delimiter), identification of species is often a nightmare. The finest phenotypic systems (e.g., Biotype-100 strips) cannot resolve species within the *P. fluorescens*, *P. putida*, or *P. syringae* complexes [10].

Siderophore typing has been proposed for the identification of *Pseudomonas* species producing fluorescent pyoverdine [14].

Ribotyping, using endonucleases *Sma*I and *Hinc*II, showed high resolution in separating DNA hybridization groups

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[3]. However, DNA sequencing is becoming less expensive, less tedious, and far more portable than ribotyping.

The high degree of conservation of the *rrs* gene (an advantage for its universality) led to a small number of informative sites in its sequence. Its utility has been questioned because of its heterogeneity [5], and it often fails to reveal precise and statistically supported phylogeny at the species level [9,11,25].

Therefore, other genes have been used to aim at a more precise phylogeny, such as *gyrB*, *rpoD* and *oprI* for the genus *Pseudomonas* [6,29].

The *rpoB* gene, which codes for the RNA polymerase beta subunit, has been used as a signature for bacterial identification as well as a locus for phylogenetic analysis [15]. Moreover, *rpoB* is a highly conserved housekeeping gene, and one copy is present in all bacteria because of its essential role in cellular metabolism [22].

Due to its discriminatory power, the *rpoB* gene has been used for developing probes for specific detection and phylogenetic analysis of several bacterial groups [4,8,16,22].

In this study, *rpoB* gene partial sequences of 900–1200 bp were determined for examining the phylogenetic structure

of the genus *Pseudomonas* and the results were compared to those obtained with the *rrs* gene. Isolates submitted to *rpoB* sequencing were identified against the sequence database built in this study.

2. Materials and methods

2.1. Bacterial strains and DNA preparation

A total of 186 strains belonging to 75 species of the genus *Pseudomonas* were studied. These strains are listed in Table 1 and Table 2. They were obtained from the Laboratorium voor Microbiologie Gent Culture Collection (LMG), Ghent, Belgium; Collection de l'Institut Pasteur (CIP), Paris, France; Collection de la Faculté de Médecine de Lille (CFML), Lille, France; American Type Culture Collection (ATCC), Manassas, VA; or received for identification in our laboratory. Strains were grown in appropriate media and temperatures following the recommendations of the culture collections, and stored at -80°C in brain–heart infusion supplemented with 50% (vol/vol) glycerol.

Table 1

GenBank accession numbers of *rrs* and *rpoB* gene sequences and ribogroup of type strains studied

Species	Strain	Ribogroup <i>rrs</i>	Accession no.	<i>rpoB</i> accession no.
<i>Marinobacterium georgiense</i>	CIP 106746T		AB021408	AJ717489
<i>Marinobacterium stanieri</i>	LMG 6847T		AB021367	AJ717490
<i>Oceanimonas doudoroffii</i>	CIP 74.9T		AB021371	AJ717491
<i>P. abietaniphila</i>	CIP 106708T		AJ011504	AJ717416
<i>P. alcaliphila</i>	CIP 108031T		AB030583	AJ717463
<i>P. aeruginosa</i>	LMG 1242T	R50	Z76651	AJ717442
<i>P. agarici</i>	LMG 2112T	R40	Z76652	AJ717477
<i>P. alcaligenes</i>	LMG 1224T	R52	Z76653	AJ717475
<i>P. amygdali</i>	LMG 2123T	R18	Z76654	AJ717462
<i>P. anguilliseptica</i>	CIP 106711T		AB21376	AJ717417
<i>P. asplenii</i>	LMG 2137T	R33	Z76655	AJ717432
<i>P. aurantiaca</i>	CIP 106718T		AB021412	AJ717421
<i>P. aureofaciens</i>	LMG 1245T	R30	Z76656	AJ717426
<i>P. avellanae</i>	CIP 105176T		U49384	AJ717469
<i>P. azotoformans</i>	CIP 106744T		D84009	AJ717458
<i>P. balearica</i>	CIP 105297T		U26418	AJ717480
<i>P. brassicacearum</i>	CIP 107059T		AF100321	AJ717436
<i>P. brenneri</i>	CIP 106646T		AF268968	AJ717482
<i>P. cannabina</i>	CIP 106140T		AJ492827	AJ717453
<i>P. caricapapaye</i>	LMG 2152T	R14	D84010	AJ717437
<i>P. cedrina</i>	CFML 96-198T		AF064461	AJ717424
<i>P. chlororaphis</i>	LMG 5004T	R30	D84011	AJ717478
<i>P. cichorii</i>	LMG 2162T	R25	Z76658	AJ717418
<i>P. citronellolis</i>	CIP 104381T		Z76659	AJ717460
<i>P. coronafaciens</i>	LMG 13190T	R24	Z76660	AJ717443
<i>P. corrugata</i>	LMG 2172T	R10	D84012	AJ717487
<i>P. cremoricolorata</i>	CIP 107616T		AB060137	AJ717476
<i>P. ficuserectae</i>	LMG 5694T	R17	AB021378	AJ717457
<i>P. flavescens</i>	CIP 104204T		U01916	AJ717468
<i>P. fluorescens</i> bv. 1	LMG 1794T	R42	D11188	AJ717451
<i>P. fragi</i>	LMG 2191T	R34	AB021413	AJ717444
<i>P. frederiksborgensis</i>	CIP 106887T		AJ249382	AJ717465
<i>P. fulva</i>	CIP 106765T		D84015	AJ717419
<i>P. fuscovaginae</i>	LMG 2158T	R32	AB021381	AJ717433

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