

Canopy profiles of photosynthetic parameters under elevated CO₂ and N fertilization in a poplar plantation

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Photosynthetic acclimation occurred only marginally.

Abstract

A poplar plantation has been exposed to an elevated CO₂ concentration for 5 years using the free air CO₂ enrichment (FACE) technique. Even after such a long period of exposure, leaves of *Populus x euramericana* have not shown clear signs of photosynthetic acclimation. Only at the end of the growing season for shade leaves was a decrease of maximum velocity of carboxylation (V_{cmax}) observed. Maximum electron transport rate (J_{max}) was increased by FACE treatment in July. Assimilation rates at CO₂ partial pressure of 400 (A_{400}) and 600 (A_{600}) $\mu\text{mol mol}^{-1}$ were not significantly different under FACE treatment. Most notably FACE significantly decreased stomatal conductance (g_s) both on upper and lower canopy leaves. N fertilization increased N content in the leaves on mass basis (N_m) and specific leaf area (SLA) in both CO₂ treatments but did not influence the photosynthetic parameters. These data show that in poplar plantations the long-term effects of elevated CO₂ on photosynthesis do not differ considerably from the short-term ones even with N deposition.

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Abbreviations: A_{400} , rate of CO₂ uptake at CO₂ partial pressure of 400 $\mu\text{mol mol}^{-1}$; A_{600} , rate of CO₂ uptake at CO₂ partial pressure of 600 $\mu\text{mol mol}^{-1}$; C_a , CO₂ partial pressure in the air; C_i , CO₂ partial pressure in the leaf; Chl, chlorophyll; E/A, elevated/ambient, i.e. FACE effect expressed as FACE/control; FACE, free air CO₂ enrichment; F/U, fertilised/unfertilised, i.e. fertilization effect; g_s , stomatal conductance; ISF, indirect site factor; J_{max} , maximum rate of electron transport; N_a , leaf nitrogen concentration on leaf area basis; N_m , leaf nitrogen concentration on leaf mass basis; P_i , inorganic phosphate ion; PPFD, photosynthetic photon flux density; RubP, ribulose-1,5-bisphosphate; SLA, specific leaf area; τ , canopy transmittance of PPFD; V_{cmax} , maximum velocity of carboxylation.

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1. Introduction

A stimulation of photosynthetic rate under elevated CO₂ is shown in most of the recent reviews (Gielen and Ceulemans, 2001; Norby et al., 1999; Saxe et al., 1998). However increased photosynthesis may not be sustained over a long-term period under CO₂ enrichment and a down-regulation often occurs after some time. This is especially true for plants growing under limiting conditions which do not allow a maintenance of active sinks necessary to accumulate the surplus of carbohydrates produced under elevated CO₂ (Stitt, 1991). The accumulation of carbohydrates in the leaves due to an

imbalance of source–sink relationships usually induces a feedback mechanism of reduction of photosynthetic capacity (Webber et al., 1994). Sage (1994) proposed that acclimation could occur under elevated CO₂ if the capacity of carboxylation exceeds the capacity to regenerate RubP. Moreover, Harley et al. (1992) suggested that the inorganic phosphate (P_i) required for the RubP regeneration could become limiting under elevated CO₂. Nevertheless, Long and Drake (1992) assessed that there is limited evidence that acclimation of photosynthesis to elevated CO₂ can involve a change in capacity for the regeneration of RubP and suggested that variation in response might be explained by variation in sink strength. Down-regulating responses are usually observed near the end of the growing seasons when the sink strength is dramatically reduced (Sholtis et al., 2004). Moreover, seasonal changes, e.g. low temperature, enhance photosynthetic down-regulation (Tsonev et al., 2003) and the same behaviour is expected with a decline of leaf N (Herrick and Thomas, 2001). Most commonly down-regulation involves a reduction of the maximum velocity of carboxylation (V_{cmax}) and of the maximum electron transport rate (J_{max}). For instance V_{cmax} was 12–20% lower in *Populus deltoides* × *Populus nigra* in mid September, but not in early August (Curtis et al., 1995). The reduction in V_{cmax} under elevated CO₂ has been shown to increase over the course of the season from 9% to 23% (Norby et al., 1999). Down-regulating responses are often due to restrictive experimental conditions such as growth in pots which do not allow a free development of below-ground biomass or growth in chambers which affects the normal development of the aerial portion and/or indirectly modifies the microclimate inducing a well-known chamber effect (Leadley and Drake, 1993). In other cases limitations could originate from modification of processes occurring at the ecosystem level under elevated CO₂ which lead to reduced nutrient availability. A reduction of nitrogen availability during long-term exposure to elevated CO₂ is often observed due to increased removal of N from soil to sustain increased growth rates and/or due to decreased decomposition rates of litter and soil organic matter. Soil N availability usually influences N content in the leaves (Hobbie et al., 2001) which tends itself to be slightly reduced by elevated CO₂. Of course in most cases this reduction can be simply explained as dilution effect, that is as a result of increased accumulation of simple carbohydrates in leaves under elevated CO₂. Nevertheless, species that showed the greatest reductions in leaf N content at elevated CO₂, also showed reduced enhancements in assimilation rates (Nowak et al., 2004). Regardless of this down-regulation, assimilation rates at growth conditions usually remain higher under elevated CO₂ regardless of the duration of the study (Norby et al., 1999).

Leaf position within the canopy can also affect the response of photosynthesis to elevated CO₂. This can be due both to a direct effect, that is the photosynthetic response to elevated CO₂ changes at different light levels, and to an indirect effect since elevated CO₂ can change both microclimatic conditions and leaf properties even when similar crown positions are considered between treatments. Kubiske et al. (1997) observed greater photosynthetic enhancements in elevated CO₂ in the lower half of the crown, although mid-crown leaves exhibited reductions in photosynthetic capacity. It has been hypothesized that elevated CO₂ and shading can interact inducing a stronger acclimation response than either factor alone (DeLucia and Thomas, 2000; Herrick and Thomas, 1999, 2001). For example it has been observed in *Populus tremuloides* that acclimation-response to elevated CO₂ is stronger in shade than in sun leaves (Kubiske et al., 2002).

Acclimation of stomatal conductance under elevated CO₂ may also be crucial to influence assimilation rates; a reduction of about 20% in trees was usually observed (Field et al., 1995) although the responses were very variable depending on the duration of the experiment, the species and the environmental constraints (Saxe et al., 1998). In a meta-analysis by Curtis and Wang (1998), a 10% decrease in stomatal conductance was reported for woody species under elevated CO₂, although this response was not statistically significant.

The possibility to study poplars, which are characterised by unlimited growth, and to perform a CO₂ enrichment using the FACE technique, i.e. in field conditions without environmental constraints and over a long-term period, provides useful information on photosynthetic properties. Moreover, by including the N treatment, it is possible to verify how crucial is the role of this nutrient on the response of fast growing trees to elevated CO₂ over long-term periods. A key question remains, whether acclimation of photosynthesis occurs most prevalently in ecosystems where N is limiting (Nowak et al., 2004). Therefore the objectives of our study were to verify (a) if acclimation occurs in a poplar plantation after 5 years of exposure to elevated CO₂ and how this is influenced by N availability, by light regimes and by the timing within the growing season and (b) if in situ stomatal conductance is reduced under elevated CO₂.

2. Materials and methods

Measurements were carried out at the POPFACE site which is located in Tuscania, Viterbo (central Italy, 42°22' N, 11°48' E). Three of the six experimental plots selected within a 9-ha short rotation coppice culture of poplars were maintained at ambient CO₂ (control) whereas the other three were maintained at a CO₂

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